



US 20070128224A1

(19) **United States**(12) **Patent Application Publication** (10) **Pub. No.: US 2007/0128224 A1**
(43) **Pub. Date: Jun. 7, 2007**
Van Der Werf et al.(54) **NOVEL STRAIN OF SARS-ASSOCIATED
CORONAVIRUS AND APPLICATIONS
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Paris (FR); **Valerie Lorin**, Montrouge
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Les Lilas (FR); **Chantal Combredet**,
Paris (FR); **Jean-Francois Delagneau**,
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WASHINGTON, DC 20001-4413 (US)(21) Appl. No.: **10/581,356**(22) PCT Filed: **Dec. 2, 2004**(86) PCT No.: **PCT/FR04/03106**§ 371(c)(1),
(2), (4) Date: **Feb. 8, 2007**(30) **Foreign Application Priority Data**Dec. 2, 2003 (FR)..... 0314151
Dec. 2, 2003 (FR)..... 0314152**Publication Classification**(51) **Int. Cl.**
A61K 39/215 (2006.01)
C12Q 1/70 (2006.01)
C07H 21/04 (2006.01)
C07K 14/165 (2006.01)
C07K 16/10 (2006.01)
C12N 5/06 (2006.01)
(52) **U.S. Cl.** **424/221.1**; 435/5; 435/69.3;
435/326; 435/456; 530/350;
530/388.3; 536/23.72; 977/802(57) **ABSTRACT**The invention relates to a novel strain of severe acute
respiratory syndrome (SARS)-associated coronavirus,
resulting from a sample collected in Hanoi (Vietnam),
reference number 031589, nucleic acid molecules originat-
ing from the genome of same, proteins and peptides coded
by said nucleic acid molecules and, more specifically, pro-
tein N and the applications thereof, for example, as diag-
nostic reagents and/or as a vaccine.

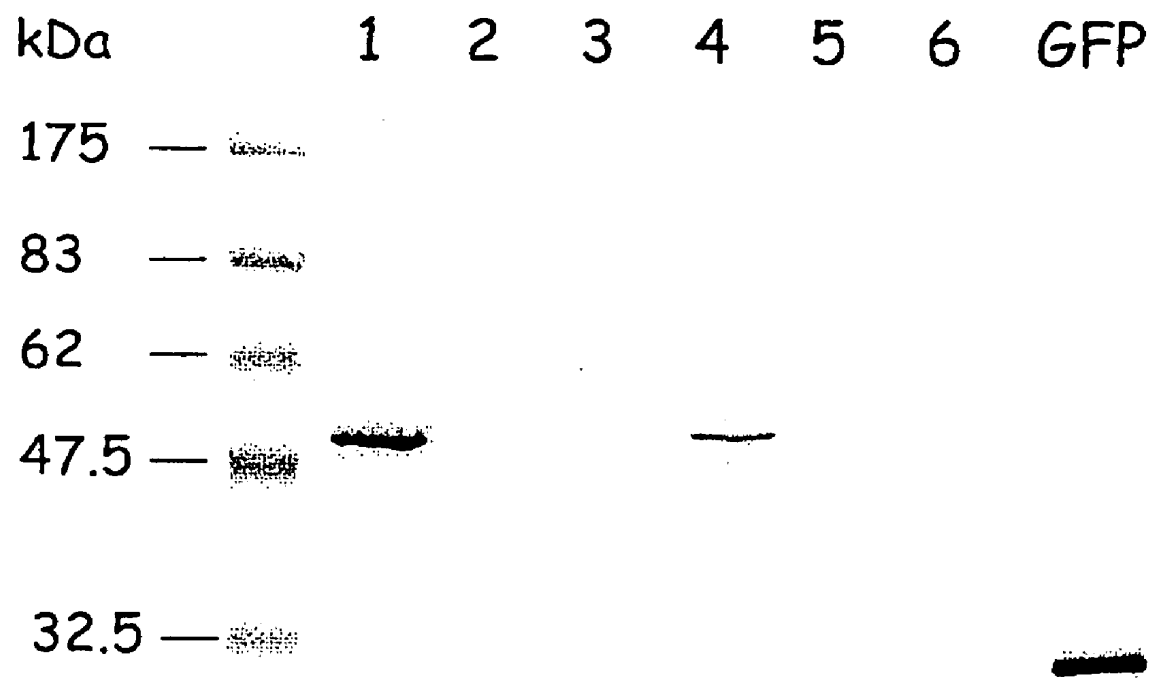


FIGURE 1

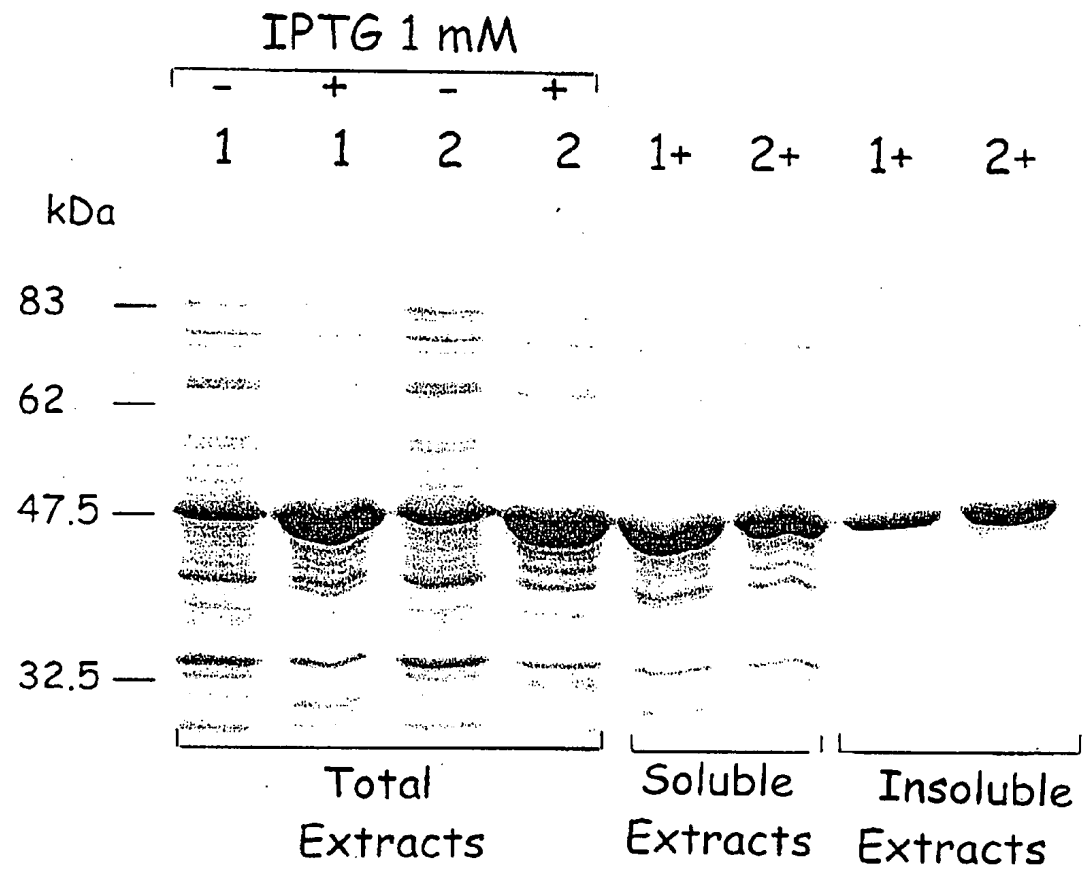


FIGURE 2

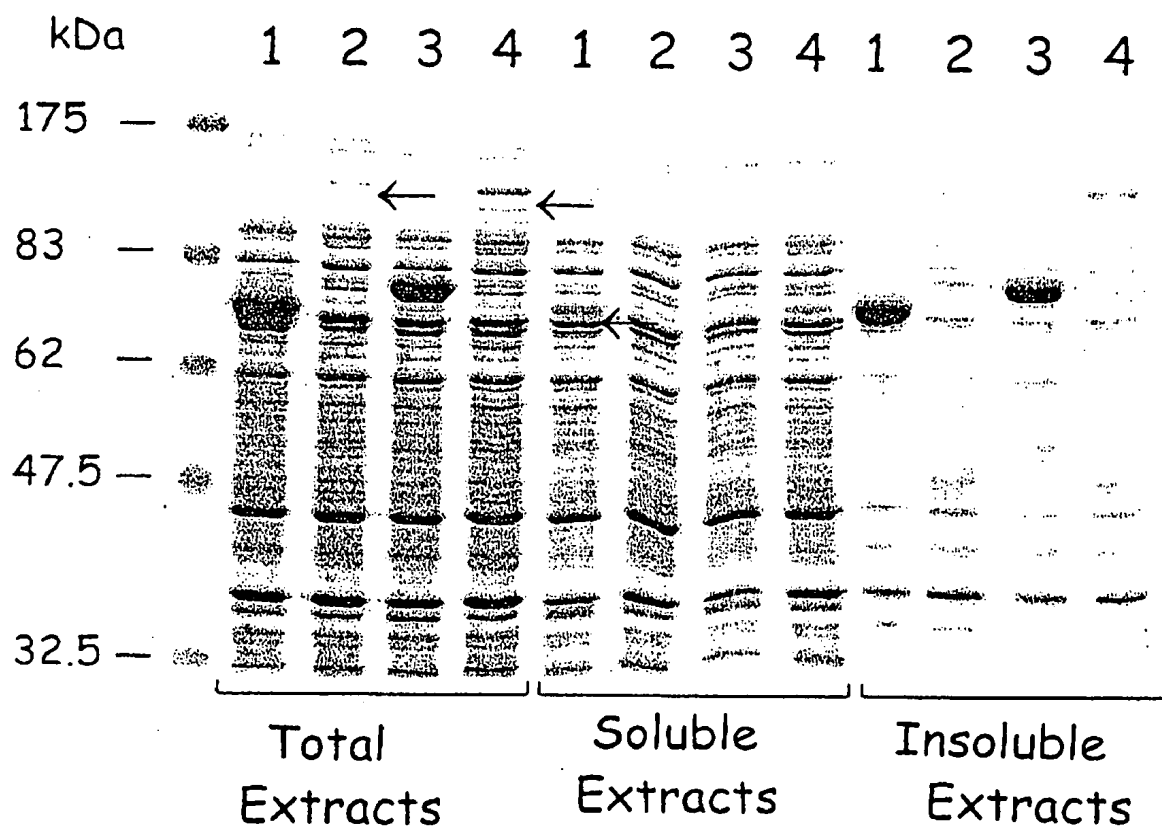


FIGURE 3

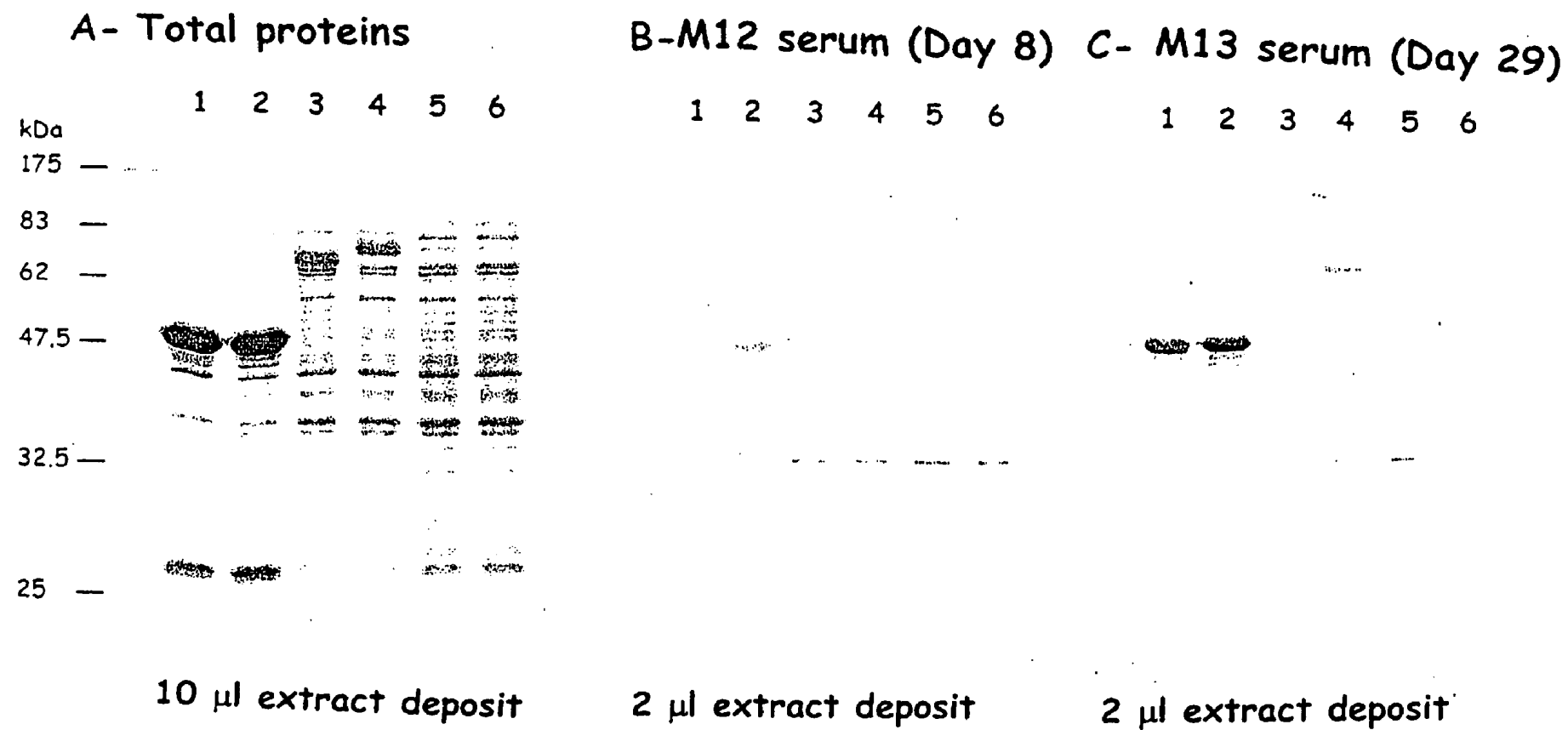


FIGURE 4

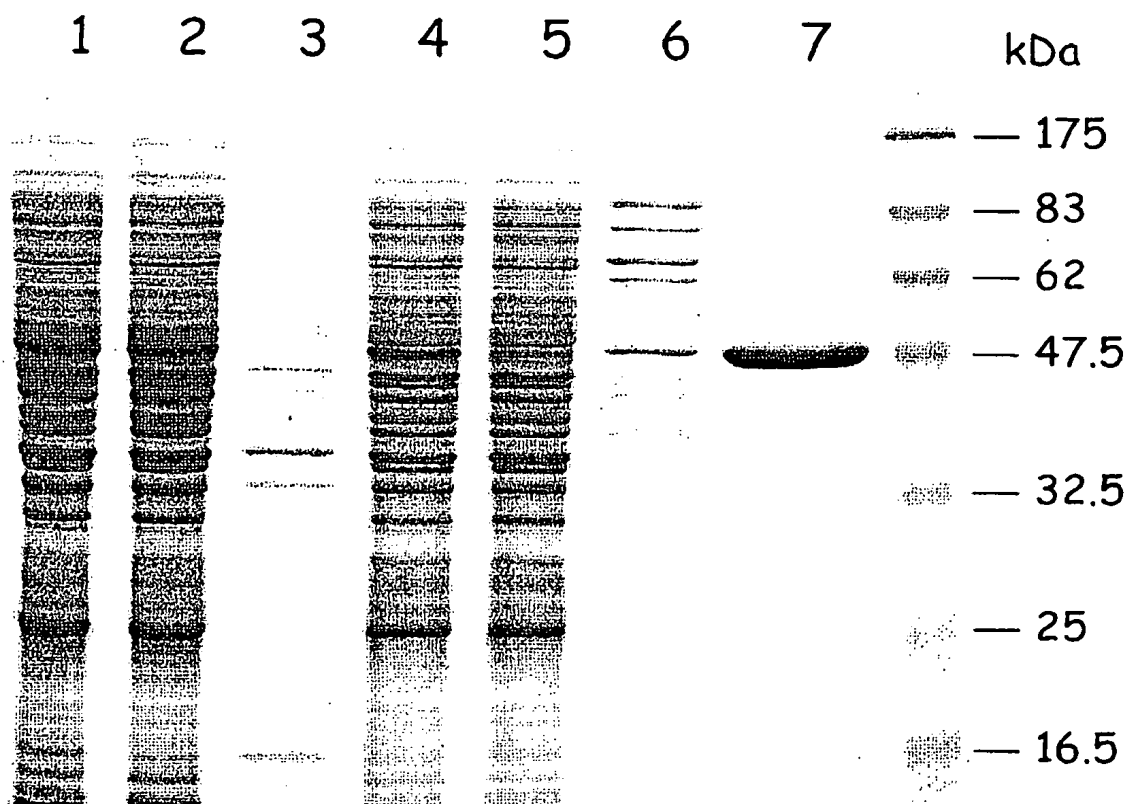


FIGURE 5

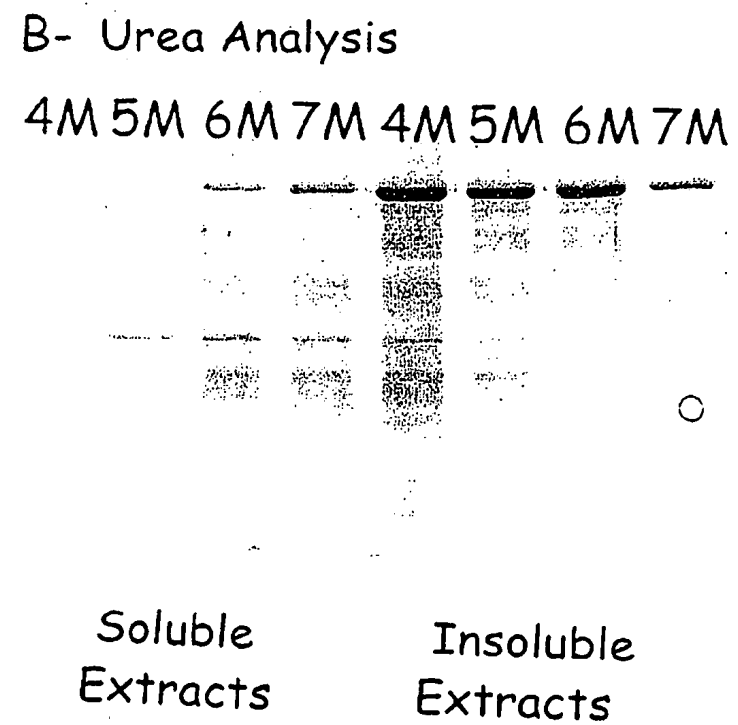
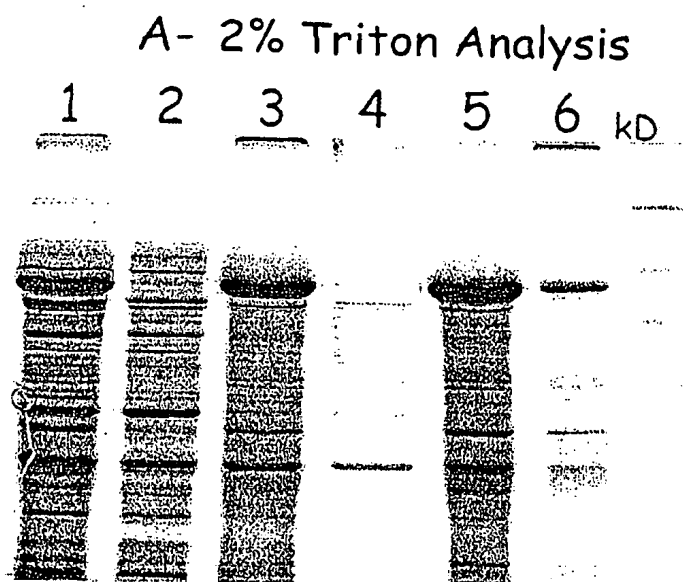


FIGURE 6

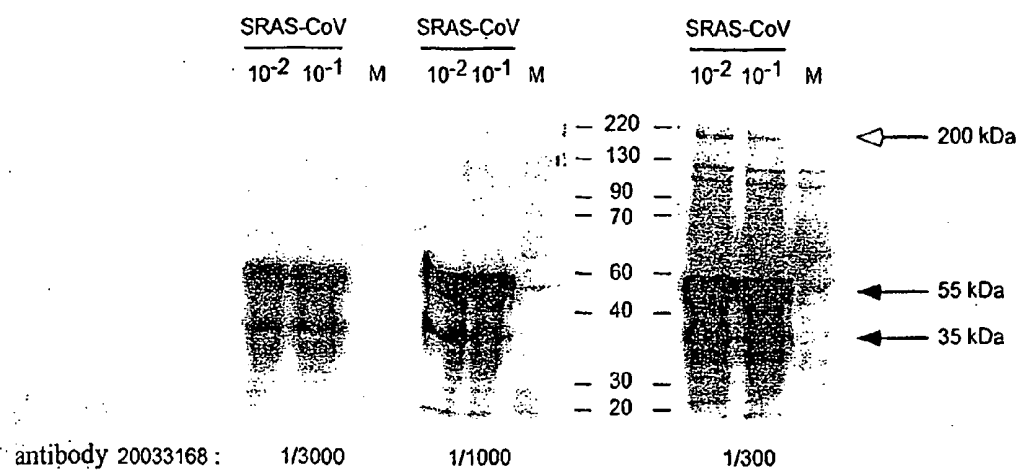


FIGURE 7

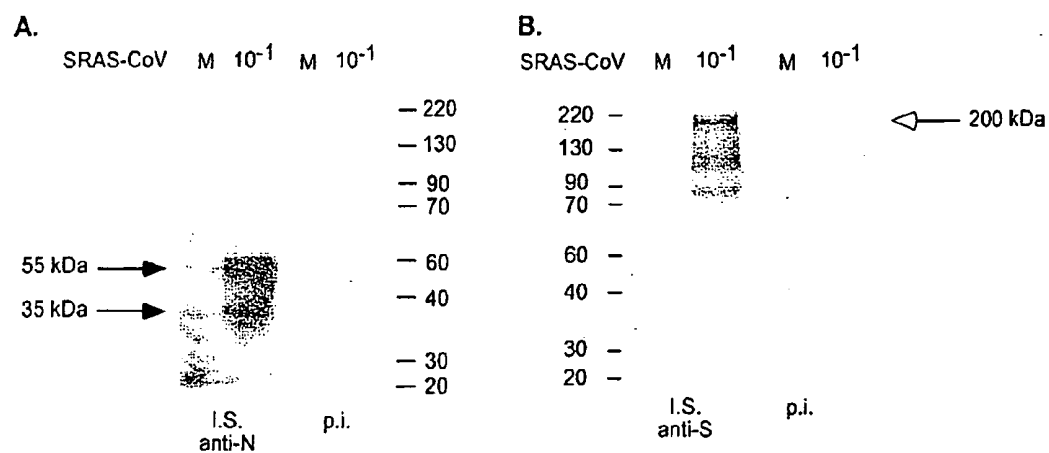
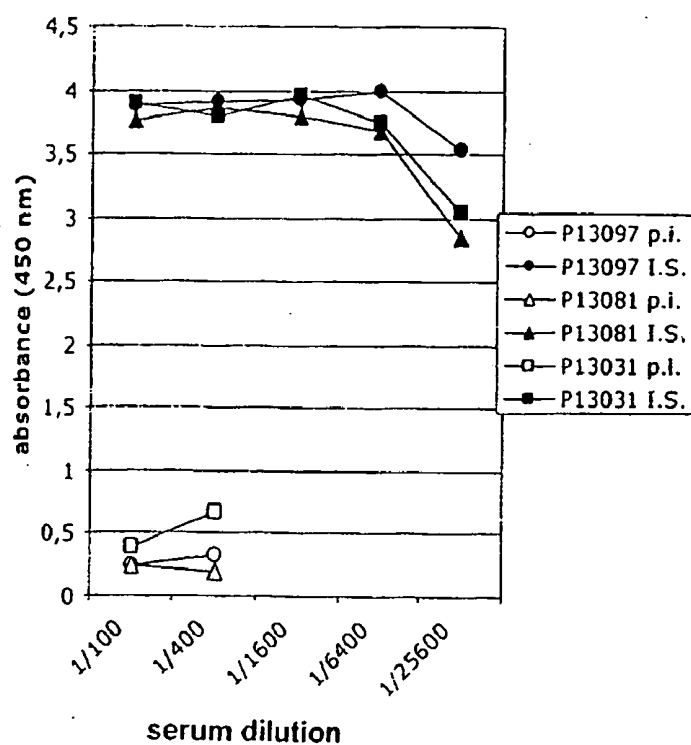


FIGURE 8

A



B

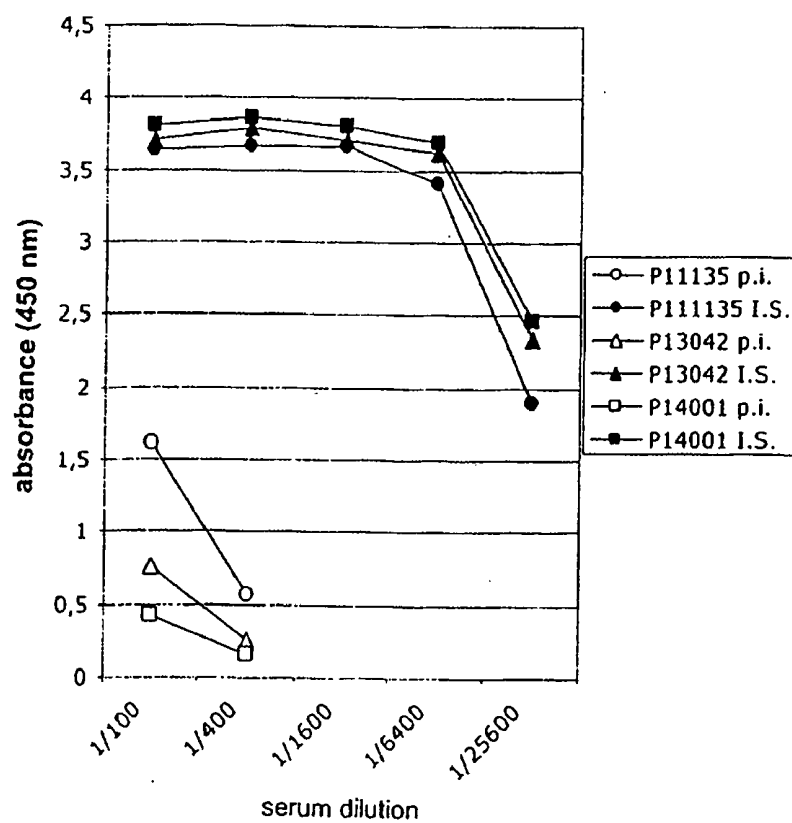


FIGURE 9

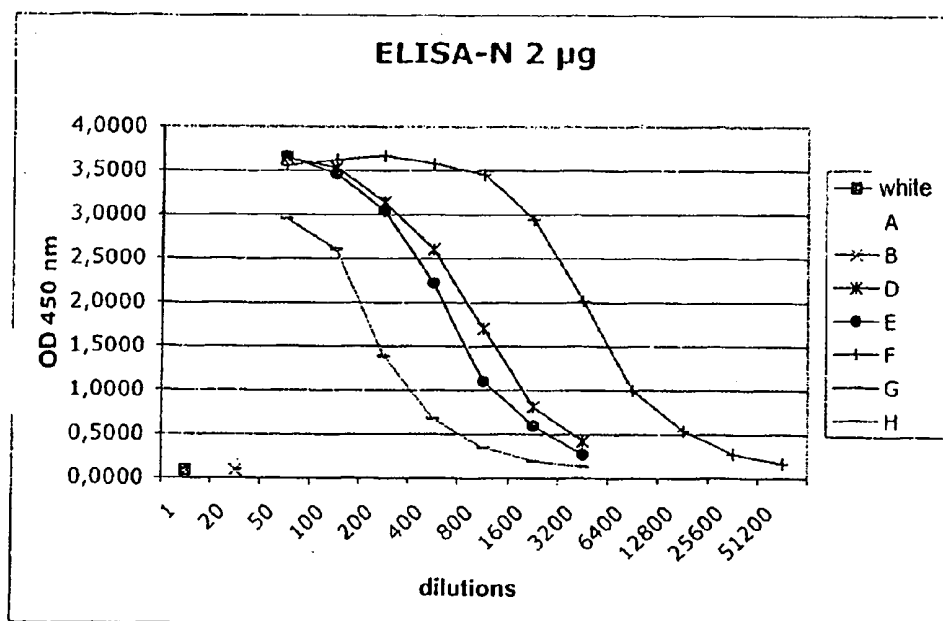
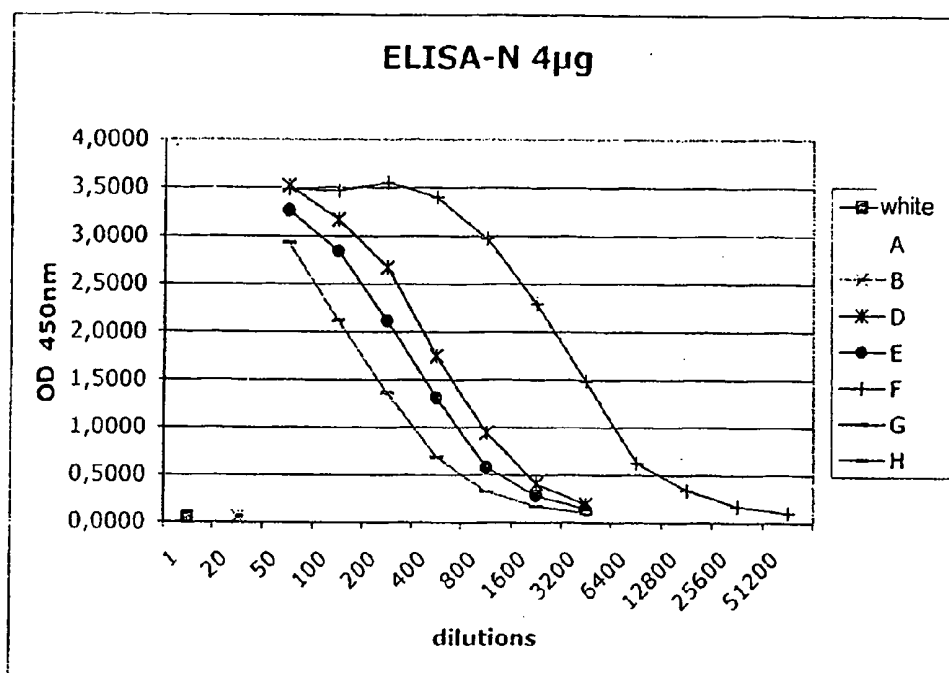


FIGURE 10a

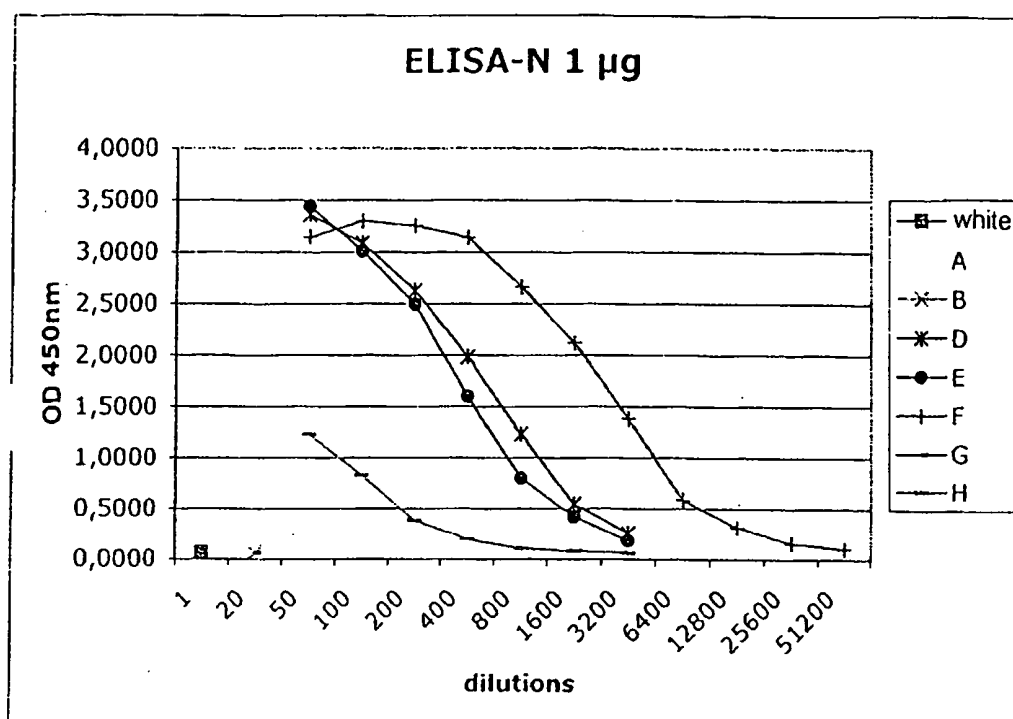


FIGURE 10b

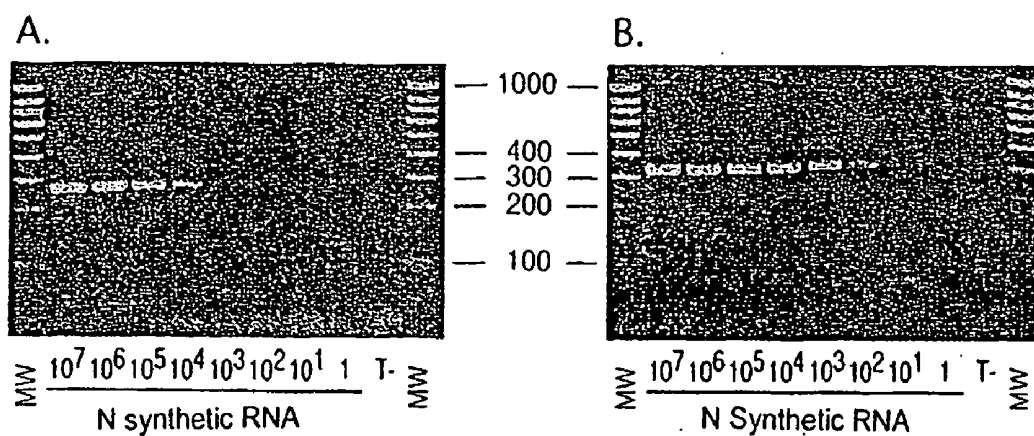


FIGURE 11

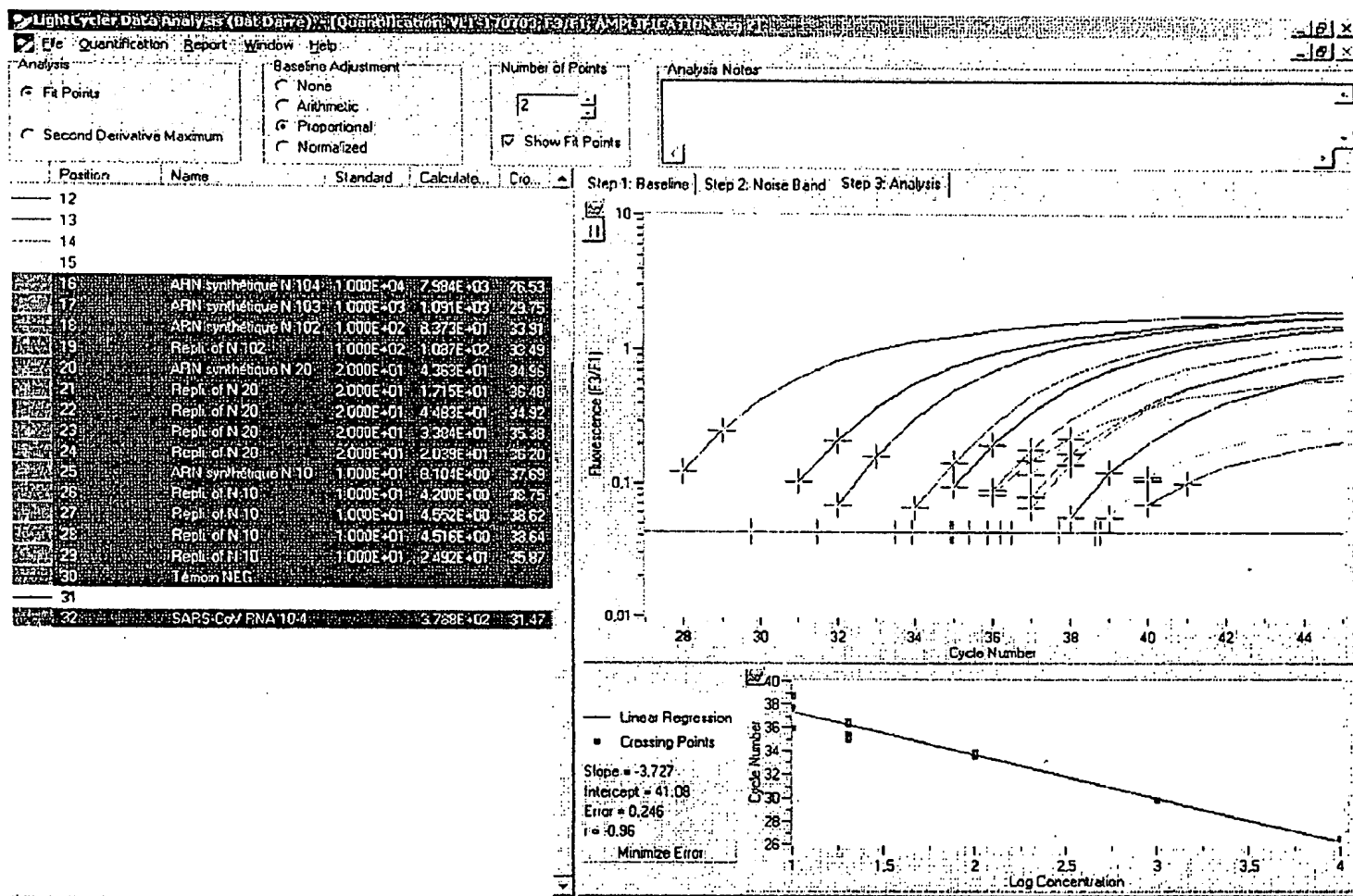


FIGURE 12

```

                >> ScrFI                                >> XhoII
                >> MvaI                                > < TthHB8I >> Sau3AI
                >> EcoRII                              > < TaqI >> NdeII
                >> Ecl136I                            >> Sau3AI >> MflI
                >> DsaV                                >> NdeII >> MboI
                >> BstOI                              >> MboI>< MnlI>< DpnI
                >> BstNI                              >> DpnII >> BstYI
                >> BsiLI                              >< DpnI >> BspAI
                >> BsaJI                              >> BspAI >> Bsp143I
                >> ApyI                                >> Bsp143I>< BglII
ATATTAGGTT TTTACCTACC CAGGAAAAGC CAACCAACCT CGATCTCTTG TAGATCTGTT CTCTAAACGA
    10          20          30          40          50          60          70

                >> VneI
                >> SphI
                >> SnaI
                >> RmaI
                >> PaeI >> SduI
                >> NspI >> NspII
                >> NspHI >> HgiAI
                >> NlaIII >> Bsp1286I
                >> MaeI >> BmyI
                >> ApaLI
                >> Alw44I
                >> DraI >> AluI > < Fnu4HI >> Alw21I
ACTTTAAAT CTGTGTAGCT GTCGCTCGGC TGCATGCCTA GTGCACCTAC GCAGTATAAA CAATAATAAA
    80          90          100          110          120          130          140

                >> SfcI
                >> PstI
                >> MnlI
                >> Ksp632I
                >> HindII > < MboII >> EarI
                >> HincII >> MaeIII >> Eam1104I
TTTTACTGTC GTTGACAAGA AACGAGTAAC TCGTCCCTCT TCTGCAGACT GCTTACGGTT TCGTCCGTGT
    150          160          170          180          190          200          210

                >> TthHB8I >> StyI
                >> TaqI >> RmaI >> ScrFI
                >> Sau3AI >> MaeI >> NciI
                >> NdeII >> EcoT14I >> MspI
                >> MboI >> Eco130I >> MaeIII
                >> DpnII >> BssTII >> HpaII
                >> DpnI >> BsaJI >> HapII
                >> BspAI >> BlnI >> DsaV
                >> Bsp143I >> AvrII >> BcnI
TGCAGTCGAT CATCAGCATA CCTAGGTTTC GTCCGGGTGT GACCGAAAGG TAAGATGGAG AGCCTTGTTT
    220          230          240          250          260          270          280

                >> RmaI
                >> HindII >> MaeII> < Eco57I >> Esp3I >> MaeII
                >> HincII > < AflIII > < DdeI >> BsmAI >> MaeI
                >> AflIII >> Alw26I >> BsmBI
TTGGTGTCAA CGAGAAAACA CACGTCCAAC TCAGTTTGCC TGTCTTCAG GTTAGAGACG TGCTAGTGCC
    290          300          310          320          330          340          350

```

FIGURE 13.1

```

>< Sau96I
  >< PssI
  >< Pali
  >< NspIV
  >< MnlI
  >< HaeIII
  >< EcoO109I
  >< DraII>< MboII >< PmlI
  >< MnlI >< Cfr13I >< PmaCI
  >< Ksp632I >< BsuRI >< MaeII
  >< HinfI >< BsiZI>< EcoNI >< Eco72I
  >< EarI >< BshI >< BslI >< BsaAI
  >< PleI >< Eam1104I>< AsuI >< BsiYI>< BbrPI >< MnlI
TGGCTTCGGG GACTCTGTGG AAGAGGCCCT ATCGGAGGCA CGTGAACACC TCAAAAATGG CACTTGTGGT
  360      370      380      390      400      410      420

  >< Tru9I
  >< SfaNI
  >< RmaI >< Csp6I >< BspWI >< MseI
  >< MaeI >< AluI >< AfaI >< AluI >< MaeII
CTAGTAGAGC TGGAAAAAGG CGTACTGCCC CAGCTTGAAC AGCCCTATGT GTTCATTAAA CGTTCTGATG
  430      440      450      460      470      480      490

  >< Pali
  >< HaeIII
  >< Tru9I >< GdiII >< RsaI
  >< MseI >< EaeI >< McrI ><
  >< Esp4I >< BsuRI >< BsmI BsiEI ><
  >< AflIII >< BshI >< AluI >< BscCI >< AfaI
CCTTAAGCAC CAATCACGGC CACAAGGTCG TTGAGCTGGT TGCAGAAATG GACGGCATTC AGTACGGTCG
  500      510      520      530      540      550      560

  >< NspI
  >< ScaI >< NspHI
  >< RsaI >< NlaIII
  >< Csp6I >< BslI
  >< BsrI >< BsiYI >< MboII
  >< AciI >< AfaI >< AflIII >< MunI >< AciI
TAGCGGTATA AACTGGGAG TACTCGTGCC ACATGTGGGC GAAACCCCAA TTGCATACCG CAATGTTCTT
  570      580      590      600      610      620      630

  >< TthHB8I
  >< TaqI
  >< Sau3AI
  >< NdeII
  >< MboI
  >< DpnII
  >< DpnI
  >< ClaI
  >< Bsu15I
  >< BspDI
  >< BspAI
  >< NlaIV
  >< MspI
  >< HpaII
  >< HapII
  >< Cfr10I
  >< BscBI >< AluI >< BanIII >< MaeIII >
  >< Bsp143I
  >< Bsp106I
  >< BsiXI
  >< BscI>< SfaNI Ddel ><
  >< BfrI ><
CTTCGTAAGA ACGGTAATAA GGGAGCCGGT GGTCTAGCT ATGGCATCGA TCTAAAGTCT TATGACTTAG
  640      650      660      670      680      690      700

```

FIGURE 13.2

```

>> Sau3AI
>> NdeII
>> MboI
>> HphI
>> DpnII
>> BspAI
>> AlwI>> DpnI
>> AluI >> Bsp143I >> MboII >> BsrI >> DdeI
VneI >>
SnoI >>
> < NlaIII
ApaLI >>
Alw44I >>
GTGACGAGCT TGGCACTGAT CCCATTGAAG ATTATGAACA AACTGGAAC ACTAAGCATG GCAGTGGTGC
710 720 730 740 750 760 770
>> SstI
>> SduI
>> SacI
>> NspII
>> MnlI
>> HgiAI
>> Eco24I
>> TthHB8I
Sau96I >>
>> SduI >> Eco24I >> TagI Pali >>
>> NspII >> Ecl136II >> Sall NspIV >>
>> HgiAI >> Bsp1286I >> RtrI HaeIII >>
>> DraIII >> BmyI >> HindII BsuRI >>
>> Bsp1286I >> BanII >> HincII BsiZI >>
>> BmyI >> Alw21I >> BsgI BshI >>
>> Alw21I >> AluI >> MaeIII >> AccI AsuI >>
ACTCCGTGAA CTCACTCGTG AGCTCAATGG AGGTGCAGTC ACTCGCTATG TCGACAACAA TTTCTGTGGC
780 790 800 810 820 830 840
>> ThaI
>> ThaI
>> MvnI
>> MvnI
>> HinPII
>> Hin6I >> VneI
>> HhaI >> SnoI
>> CfoI >> SduI
>> BstUI NspII >>
>> BstUI HgiAI >>
>> Bsp50I Bsp1286I >>
>> Bsp50I >> BmyI
>> AciI >> ApaLI
>> AccII >> Alw44I
>> Acc65I >> MnlI >> SfaNI >> AccII Alw21I >>
CCAGATGGGT ACCCTCTTGA TTGCATCAAA GATTTTCTCG CACGCGCGGG CAAGTCAATG TGCACTCTTT
850 860 870 880 890 900 910
>> TthHB8I
>> TthHB8I
>> TaqI
>> TaqI
>> MnlI
>> Ksp632I
>> HinfI>> PleI
>> Eam1104I >> MboII >> MaeIII EcoRII >>
>> EarI >> BbvI>> AccI >> Fnu4HI DsaV >>
CCGAACAACT TGATTACATC GAGTCGAAGA GAGGTGTCTA CTGCTGCCGT GACCATGAGC ATGAAATTGC
920 930 940 950 960 970 980
>> TthHB8I
>> TaqI
>> SfuI
>> NspV>> Tru9I
>> ScrFI >> HinPII >> LspI>> MseI

```

FIGURE 13.3

```

>< MvaI      >< Hin6I      >< SduI      >< Csp45I
>< Ecl136I   >< HhaI      >< NspII     >< BstBI
>< BstOI     >< HaeIII    >< HgiAI     >< Bsp119I
>< BstNI     >< Eco47III   >< Bsp1286I  >< BsiCI
>< BsiLI     >< CfoI      >< BmyI      >< Bpu14I
>< ApyI >< DdeI >< Bsp143II >< AluI   >< Alw21I   >< AsuII
CTGGTTCAC T GAGCGCTCTG ATAAGAGCTA CGAGCACCAG ACACCCTTCG AAATTAAGAG TGCCAAGAAA
    990      1000      1010      1020      1030      1040      1050

                                >< Tru9I
                                >< MseI
                                >< BsmI
                                >< BscCI
                                >< MnlI
TTTGACACTT TCAAAGGGGA ATGCCCAAAG TTTGTGTTTC CTCTTAAGTC AAAAGTCAAA GTCATTCAAC
    1060      1070      1080      1090      1100      1110      1120

>< PmlI
>< PmaCI
>< MaeII
>< Eco72I
>< BsaAI
>< BbrPI
>< AflIII    >< MnlI>< DdeI    >< NlaIII    >< Bst1107I >< RsaI
>< AflIII    >< MnlI>< DdeI    >< AccI      >< Csp6I
CACGTGTTGA AAAGAAAAAG ACTGAGGGTT TCATGGGGCG TATACGCTCT GTGTACCTG TTGCATCTCC
    1130      1140      1150      1160      1170      1180      1190

>< SfaNI
>< MaeIII    >< AccI      NlaIII ><
ACAGGAGTGT AACAATATGC ACTTGCTAC CTTGATGAAA TGTAATCATT GCGATGAAGT TTCATGGCAG
    1200      1210      1220      1230      1240      1250      1260

                                >< SinI
                                >< Sau96I
                                PssI ><
                                >< Psp5II
                                >< PpuMI
                                >< NspIV
                                >< NspHII
                                >< Eco47I
                                >< DraII
                                >< Cfr13I
                                >< BsiZI
                                >< Bme18I
                                >< AvaII
                                >< AsuI
>< MaeII
ACGTGCGACT TTCTGAAAGC CACTTGTAAC CATTGTGGCA CTGAAAATTT AGTTATTGAA GGACCTACTA
    1270      1280      1290      1300      1310      1320      1330

                                EcoO109I ><AflIII >
                                Van91I ><
                                SinI ><
                                Sau96I ><
                                PflMI ><
                                NspIV ><
                                NspHII >
                                Eco47I ><
                                Cfr13I ><
                                BslI ><
                                BsiZI ><
                                BsiYI ><
                                Bme18I ><
                                AvaII ><
                                AsuI ><

>< RsaI
>< NspI
>< NlaIV
>< NlaIII
>< NspHI>< KpnI
>< Eco64I
>< Csp6I
>< BscBI
>< BanI
>< Asp718
>< AfaI
>< AccBII

```

FIGURE 13. 4

```

    >< Acc65I          >< SfcI          >< NlaIII          AccB7I ><
CATGTGGGTA CCTACCTACT AATGCTGTAG TGAATATGCC ATGTCCTGCC TGTCAAGACC CAGAGATTGG
    1340          1350          1360          1370          1380          1390          1400

                                >< TthHB8I
                                >< TaqI>< MnlI
                                >< HinfI

    >< DdeI          >< PleI          >< AciI
ACCTGAGCAT AGTGTTCGAG ATTATCACAA CCACTCAAAC ATTGAACTC GACTCCGCAA GGGAGGTAGG
    1410          1420          1430          1440          1450          1460          1470

    >< RmaI          NlaIV ><
    >< MnlI          >< BsrI
    >< MaeI          >< BbvI          >< Fnu4HI          BscBI ><
ACTAGATGTT TTGGAGGCTG TGTGTTTGGC TATGTTGGCT GCTATAATAA GCGTGCCTAC TGGGTTCTTC
    1480          1490          1500          1510          1520          1530          1540

                                XhoII ><
                                Sau3AI ><
                                NdeII ><
                                MflI ><
                                MboI ><

                                >< MaeIII
                                >< Eco3II          DpnII ><
                                >< Pali          >< HaeIII          >< BsrI          >< MnlI DpnI >
    >< RmaI          >< BsuRI          >< BsrI          >< BsmAI          BstYI ><
    >< MnlI          >< DdeI          >< BspWI          >< BsaI>< HphI          BspAI ><
    >< MaeI          >< BshI>< BglI          >< Alw26I          Bsp143I >
GTGCTAGTGC TGATATTGGC TCAGGCCATA CTGGCATTAC TGGTGACAAT GTGGAGACCT TGAATGAGGA
    1550          1560          1570          1580          1590          1600          1610

                                >< Tru9I
                                >< MseI
                                >< MaeII          >< Tru9I
                                >< HpaI          >< MnlI
                                >< HindII          >< Ksp632I
                                >< HinfI >< PleI >< HincII          >< EarI
    >< AlwI          >< DdeI          >< AflIII          >< MseI          >< Eam1104I
TCTCCTTGAG ATRACTGAGTC GTGAACGTGT TAACATTAAC ATTGTTGGCG ATTTTCATTT GAATGAAGAG
    1620          1630          1640          1650          1660          1670          1680

    >< MboII          PleI ><
    >< BstXI          >< SfaNI          >< HinfI
GTTGCCATCA TTTTGGCATC TTTCTCTGCT TCTACAAGTG CCTTTATTGA CACTATAAAG AGTCTTGATT
    1690          1700          1710          1720          1730          1740          1750

                                >< StyI
                                >< MaeIII
                                >< EcoT14I
                                >< Eco130I
                                >< PleI          >< BssT1I          BslI ><
                                >< MaeIII          >< BsaJI          BsiYI ><
    >< HinfI>< AciI          >< BsaJI          BsiYI ><
ACAAGTCTTT CAAAACCATT GTTGAGTCCT GCGGTAAC TAAGTTACC AAGGGAAAGC CCGTAAAGG
    1760          1770          1780          1790          1800          1810          1820

                                >< Sau3AI          >< Van9II
                                >< NdeII          >< PflMI
                                >< MboI          >< DraIII
                                >< DpnII          >< BslI
                                >< DpnI >< Tru9I          >< BsiYI
    >< BspAI          >< MseI          >< BbvI          >< MnlI
    >< Bsp143I          >< AccB7I          Fnu4HI ><

```

FIGURE 135

```

TGCTTGAAC ATTGGACAAC AGAGATCAGT TTTAACACCA CTGTGTGGTT TTCCCTCACA GGCTGCTGGT
 1830      1840      1850      1860      1870      1880      1890

      >< ThaI
      >< SfaNI
      >< MvnI
      >< HinPII
    >< HinPII
      >< Hin6I
    >< Hin6I
      >< HhaI
    >< Sau3AI >< HhaI
    >< NdeII >< CfoI
    >< MboI >< CfoI
    >< DpnII >< BstUI
      >< DpnI >< BssHII
    >< BspAI >< Bsp50I
      >< Bsp143I >< AccII >< Fnu4HI >< BbvI
GTTATCAGAT CAATTTTTCG GCGCACACTT GATGCAGCAA ACCACTCAAT TCCTGATTG CAAAGAGCAG
 1900      1910      1920      1930      1940      1950      1960

      >< TthHBBI
      >< StyI
      >< NcoI
      >< HindII
      >< HincII
      >< HinfI
      >< EcoT14I
      >< Eco57I
      >< TaqI>< Eco130I
    >< SalI >< DsaI
    >< RtrI >< BssTII
      >< BsaHI
      >< BbiII>< NlaIII
      >< AclI >< HgaI
    >< MaeIII
      >< BbvI
      >< MaeII >< AccI>< BsaJI
CTGTCACCAT ACTTGATGGT ATTTCTGAAC AGTCATTACG TCTTGTGCGAC GCCATGGTTT ATACTTCAGA
 1970      1980      1990      2000      2010      2020      2030

      >< RsaI
      >< Csp6I
    >< BspMI
      >< NdeI
      >< MaeIII >< BsrI >< AfaI >< DdeI
CCTGCTCACC AACAGTGTCA TTATTATGGC ATATGTAAGT GGTGGTCTTG TACAACAGAC TTCTCAGTGG
 2040      2050      2060      2070      2080      2090      2100

      >< StuI
      >< PstI
      >< MaeIII
      >< Eco147I
      >< SduI >< DdeI
      >< NspII >< BsuRI
      >< Bsp1286I >< BshI
      >< BmyI >< AatI >< MnlI
TTGTCTAATC TTTTGGGCAC TACTGTTGAA AAACCTCAGGC CTATCTTTGA ATGGATTGAG GCGAAACTTA
 2110      2120      2130      2140      2150      2160      2170

      >< TfiI
      >< HinfI
      >< SfaNI >< BsgI >< FokI
GTGCAGGAGT TGAATTTCTC AAGGATGCTT GGGAGATTCT CAAATTTCTC ATTACAGGTG TTTTGTGACAT
 2180      2190      2200      2210      2220      2230      2240

```

FIGURE 13.6


```

Tru9I >>
MseI >>
HpaI >
HindII >
HincII >
>< Eco57I
CGTCAAGGGT CAAATACAGG TTGCTTCAGA TAACATCAAG GATTGTGTAA AATGCTTCAT TGATGTTGTT
2250      2260      2270      2280      2290      2300      2310

>< Sau3AI
>< NdeII
>< MboI
> < MaeIII
>< FbaI
>< DpnII
>< DpnI
>< BspAI
>< Bsp143I
>< TthHB8I
>< TaqI
AACAAAGGCAC TCGAAATGTG CATTGATCAA GTCACATATCG CTGGCGCAAA GTTGCGATCA CTCAACTTAG
2320      2330      2340      2350      2360      2370      2380

>< PvuII
>< MaeII
>< Bst1107I
>< BsaAI
>< BbvI
>< HphI
>< DrdI
>< AccI
GTGAAGTCTT CATCGCTCAA AGCAAGGGAC TTTACCGTCA GTGTATACGT GGCAAGGAGC AGCTGCAACT
2390      2400      2410      2420      2430      2440      2450

>< Tru9I
>< NlaIV
>< MseI
>< MnlI
>< Esp4I
>< Eco64I
>< BscBI
>< NlaIII >< BanI
>< AflIII
>< BbvI
>< AccBII
>< MaeIII
ACTCATGCCT CTTAAGGCAC CAAAAGAAGT AACCTTTCTT GAAGGTGATT CACATGACAC AGTACTTACC
2460      2470      2480      2490      2500      2510      2520

> < XhoI
>< TthHB8I
>< TthHB8I>> TaqI
> < SlaI
> < Pae7I
> < NspIII
>< HphI >< HinII
> < Eco88I
> < CcrI
>< Esp3I >< BsaHI
> < BcoI
>< BsmAI >< BbiII
> < Aval >< HgaI
>< TaqI > < Ama87I>< BsmBI
>< Alw26I >< AcyI
>< DdeI>< MnlI
TCTGAGGAGG TTGTTCTCAA GAACGGTGAA CTCGAAGCAC TCGAGACGCC CGTTGATAGC TTCACAAATG
2530      2540      2550      2560      2570      2580      2590

```

FIGURE 13.7

```

                                >< PstI >< NlaIII
                                >< HaeIII >< MnlI
                                >< BsuRI >< DdeI >< Tru9I
                                >< BshI >< BfrI >< MseI
>< AluI >< BsrI
GAGCTATCGT TGGCACACCA GTCTGTGTAA ATGGCCTCAT GCTCTTAGAG ATTAAGGACA AAGAACAATA
2600 2610 2620 2630 2640 2650 2660

                                >< VneI
                                Tru9I ><
                                >< SnaI
                                >< SduI
                                >< NspII
                                MseI ><
                                >< HgiAI
                                Bsp1286I >< BslI ><
                                BsiYI ><
                                >< BmyI
                                >< ApaLI
                                >< Tru9I >< Alw44I
                                >< MseI >< Alw21I
CTGCGCATTG TCTCCTGGTT TACTGGCTAC AAACAATGTC TTTCGCTTAA AAGGGGGTGC ACCAATTAAA
2670 2680 2690 2700 2710 2720 2730

                                >< TfiI
>< MaeIII >< MboII >< MaeIII >< HinfI AluI ><
GGTGTAACTT TTGGAGAAGA TACTGTTTGG GAAGTTCAAG GTTACAAGAA TGTGAGAATC ACATTGAGC
2740 2750 2760 2770 2780 2790 2800

                                >< RsaI
                                >< NlaIV
                                MaeIII ><
                                >< MspI>< KpnI
                                >< HpaII
                                >< HapII
                                >< Eco64I
                                >< SduI >< Csp6I
                                >< NspII >< TfiI >< BscBI
                                >< HgiAI >< BanI
                                >< Bsp1286I >< Asp718
                                >< BmyI >< HinfI >< AfaI
                                >< Alw21I >< AccB1I
                                >< AccI >< Acc65I
TTGATGAACG TGTTGACAAA GTGCTTAATG AAAAGTGCTC TGTCTACACT GTTGAATCCG GTACCGAAGT
2810 2820 2830 2840 2850 2860 2870

                                >< Sau3AI
                                >< NdeII
                                >< MboI
                                >< DpnII
                                >< DpnI
                                >< NspI
                                >< NspHI
                                >< NlaIII
                                >< MboII >< BspAI
                                >< BsrI >< Bsp143I
>< DdeI >< MslI >< AlwNI >< BbsI >< AlwNI
TACTGAGTTT GCATGTGTTG TAGCAGAGGC TGTGTGAAG ACTTTACAAC CAGTTTCTGA TCTCCTTACC
2880 2890 2900 2910 2920 2930 2940

                                >< Sau3AI
                                >< NdeII
                                >< MboI
                                >< DpnII
                                >< DpnI
                                >< BspAI

```

FIGURE 13.8

```

    >> NlaIII>> Bsp143I          >> AluI          >> SfaNI
AACATGGGTA TTGATCTTGA TGAGTGGAGT GTAGCTACAT TCTACTTATT TGATGATGCT GGTGAAGAAA
    2950          2960          2970          2980          2990          3000          3010

                                >> SfaNI
                                >> MnlI
    >> MboII          >> GsuI          >> Ksp632I          >> MnlI
                                >> BsaAI          >> EarI          >> MboII
    >> HphI >> MaeII>> BpmI          >> MnlI          >> Eam1104I          >> MboII
ACTTTTCATC ACGTATGTAT TGTTCCTTTT ACCCTCCAGA TGAGGAAGAA GAGGACGATG CAGAGTGTGA
    3020          3030          3040          3050          3060          3070          3080

                                >> RsaI
                                >> RsaI
    >> NlaIII
                                >> MnlI          >> FokI
                                >> Csp6I          Eco31I >>
                                >> Csp6I          >> MamI BsmAI >>
    >> MboII          >> AfaI          >> BsiBI BsaI >>
    >> MboII          >> AfaI          >> BsaB1Alw26I >>
GGAAGAAGAA ATTGATGAAA CCTGTGAACA TGAGTACGGT ACAGAGGATG ATTATCAAGG TCTCCCTCTG
    3090          3100          3110          3120          3130          3140          3150

    >> NlaIV>> PvuII>> XmnI
    >> Eco64I >> Psp5I          >> TthHB8I
    >> MnlI >> DdeI          >> TaqI          >> MnlI          >> MboII
    >> BscBI>> NspBII >> MnlI          >> Ksp632I          >> MboII >> MboII
    >> BanI          >> MnlI          >> EarI          >> BsrI
    >> AccB1I >> AluI >> Asp700I          >> Eam1104I >> MboII>> BbsI
GAATTTGGTG CCTCAGCTGA AACAGTTCGA GTTGAGGAAG AAGAAGAGGA AGACTGGCTG GATGATACTA
    3160          3170          3180          3190          3200          3210          3220

                                >> Tru9I
    >> FokI          >> MseI          >> Eco57I
    >> DdeI          >> BsrI>> MboII BsrI >>
CTGAGCAATC AGAGATTGAG CCAGAACCAG AACCTACACC TGAAGAACCA GTTAATCAGT TTACTGGTTA
    3230          3240          3250          3260          3270          3280          3290

    >> Tru9I          >> MnlI
    >> MseI          >> Tru9I >> HindII>> Tru9I          >> DraIII
    >> DraI          >> MseI >> HincII>> MseI          >> BspWI
TTTAAACTT ACTGACAATG TTGCCATTAA ATGTGTTGAC ATCGTTAAGG AGGCACAAAG TGCTAATCCT
    3300          3310          3320          3330          3340          3350          3360

                                >> VneI
                                >> SnaI
                                >> SduI
                                >> NspII
                                >> HqiAI
                                >> Bsp1286I
                                >> BmyI
                                >> ApaLI
    >> HphI          >> NlaIII          >> Alw44I
    >> BbvI          >> Fnu4HI          >> BspMI          >> Alw21I
ATGGTGATTG TAAATGCTGC TAACATACAC CTGAAACATG GTGGTGGTGT AGCAGGTGCA CTCAACAAGG
    3370          3380          3390          3400          3410          3420          3430

                                >> Sau96I
                                >> Pali
                                >> NspIV
                                >> HaeIII
    >> NlaIV          >> Cfr13I

```

FIGURE 13.9

```

>< Eco64I                >< BsuRI
>< BscBI                > < Tru9I    >< BsiZI
>< BaniI                > < MseI    >< BshI    >< MnlI
>< AccBII>< NlaIII      >< AluI    >< AsuI    >< MnlI
CAACCAATGG TGCCATGCAA AAGGAGAGTG ATGATTACAT TAAGCTAAAT GGCCCTCTTA CAGTAGGAGG
3440      3450      3460      3470      3480      3490      3500

>< SinI
>< Sau96I
>< NspIV
>< NspHI>< NspHII
>< Eco47I
>< Cfr13I
>< NlaIII    >< BspMI
>< BsiZI
>< Bmel8I
>< AvaII MnlI ><
> < DdeI      >< NspI>< AsuI FokI ><
GTCTGTTTG CTTTCTGGAC ATAATCTTGC TAAGAAGTGT CTGCATGTTG TTGGACCTAA CCTAAATGCA
3510      3520      3530      3540      3550      3560      3570

> < Tru9I
>< HphI> < MseI
>< Esp4I
>< AluI      > < NdeI
>< AflII>< Fnu4HI >< BbvI
GGTGAGGACA TCCAGCTTCT TAAGGCAGCA TATGAAAATT TCAATTCACA GGACATCTTA CTTGCACCAT
3580      3590      3600      3610      3620      3630      3640

RsaI ><
Csp6I ><
AfaI ><
>< Eco57I                >< BcgI
TGTGTGCAGC AGGCATATTT GGTGCTAAAC CACTTCAGTC TTTACAAGTG TGCGTGCAGA CGGTTTCGTAC
3650      3660      3670      3680      3690      3700      3710

>< BsqI                >< BspMI
>< BcgI/a                >< AluI                >< NlaIII
ACAGGTTTAT ATTGCAGTCA ATGACAAAGC TCTTTATGAG CAGGTTGTCA TGGATTATCT TGATAACCTG
3720      3730      3740      3750      3760      3770      3780

>< RmaI                > < MnlI    >< NlaIV    >< TfiI    >< MboII
>< MaeI                >< Eco57I    >< BscBI    >< HinfI    >< DdeI
AAGCCTAGAG TGGAAGCACC TAAACAAGAG GAGCCACCAA ACACAGAAGA TTCCAAAACT GAGGAGAAAT
3790      3800      3810      3820      3830      3840      3850

>< Tru9I
>< StuI
>< Pali
>< MseI    >< MnlI    >< MaeIII
>< HaeIII    >< EcoO65I
>< Eco147I    >< Eco91I
>< BsuRI                BstXI ><
>< BshI                >< BstPI
>< AatI                >< BstEII
CTGTCGTACA GAAGCCTGTC GATGTGAAGC CAAAAATTAA GGCCTGCATT GATGAGGTTA CCACAACACT
3860      3870      3880      3890      3900      3910      3920

TfiI ><
NlaIII ><
HinfI ><
>< DdeI                >< EcoRV    >< HindIII

```

FIGURE 13.10

```

    >< BsrI      >< MboII      >< MaeIII      >< Eco32I      >< AluI
GGAAGAACT AAGTTTCTTA CCAATAAGTT ACTCTTGTTC GCTGATATCA ATGGTAAGCT TTACCATGAT
    3930      3940      3950      3960      3970      3980      3990

    >< NspI
    >< NspHI
    >< NlaIII
    >< MnlI      >< SfaNI
    >< DdeI      >< EcoNI
    >< MboII >< BslI      >< NlaIII
    >< DdeI      >< BfrI      >< HphI      >< BsiYI      >< FokI
TCTCAGAACA TGCTTAGAGG TGAAGATATG TCTTTCCTTG AGAAGGATGC ACCTTACATG GTAGGTGATG
    4000      4010      4020      4030      4040      4050      4060

    >< SpeI
    >< RmaI
    >< MaeI      >< EcoRV>< HphI      >< SfaNI
    >< HphI      >< Eco32I      >< MnlI      >< DdeI
TTATCACTAG TGGTGATATC ACTTGTGTTG TAATACCCTC CAAAAGGCT GGTGGCACTA CTGAGATGCT
    4070      4080      4090      4100      4110      4120      4130

    >< ScrFI
    >< RsaI
    >< MvaI
    >< EcoRII
    >< Ecl136I
    >< DsaV
    >< Csp6I >< EcoNI
    >< BstOI
    >< BstNI
    >< BsiLI
    >< BsaJI
    >< BsaAI      >< BslI
    >< MaeII>< ApyI
    >< AfaI      >< BsiYI
CTCAAGAGCT TTGAAGAAAG TGCCAGTTGA TGAGTATATA ACCACGTACC CTGGACAAGG ATGTGCTGGT
    4140      4150      4160      4170      4180      4190      4200

    >< Tru9I
    >< MseI
    >< DdeI      >< Esp4I      >< RsaI
    >< MnlI      >< BspWI      >< Csp6I
    >< FokI      >< AluI      >< AflII      >< Eco57I >< AfaI
TATACACTTG AGGAAGCTAA GACTGCTCTT AAGAAATGCA AATCTGCATT TTATGTACTA CCTTCAGAAG
    4210      4220      4230      4240      4250      4260      4270

    >< ScrFI
    >< MvaI
    >< EcoRII
    >< XmnI      >< Ecl136I      NlaIII ><
    >< Ksp632I >< RmaI      >< DsaV      Ksp632I ><
    >< EarI      >< TfiI>< MboII      >< BstOI      >< EarI
    >< Eam1104I >< MaeI      >< BstNI      Eam1104I ><
    >< DdeI      >< HinfI      >< BsiLI      BsmAI ><
    >< BspWI      >< Asp700I      >< ApyI      Alw26I ><
CACCTAATGC TAAGGAAGAG ATTCTAGGAA CTGTATCCTG GAATTTGAGA GAAATGCTTG CTCATGCTGA
    4280      4290      4300      4310      4320      4330      4340

    >< VspI      >< Zsp2I
    >< Tru9I      >< Ppu10I
    >< MseI      >< NsiI
    >< MboII      >< NlaIII      >< FokI
    >< Eco57I      >< Mph1103I >< FokI

```

FIGURE 13. 11

```

                >< AsnI          >< EcoT22I          >< BspWI
                >< AseI          >< AvaIII          >< BglI          >< MaeII
AGAGACAAGA AAATTAATGC CTATATGCAT GGATGTTAGA GCCATAATGG CAACCATCCA ACGTAAGTAT
      4350      4360      4370      4380      4390      4400      4410

                >< SfaNI
                > < HindII          >< TfiI          >< SpeI
                >< MseI          > < HincII>< MboII          >< RmaI
                >< MnlI          >< DrdI >< HinfI          >< MaeI
AAAGGAATTA AAATTCAAGA GGGCATCGTT GACTATGGTG TCCGATTCTT CTTTATACT AGTAAAGAGC
      4420      4430      4440      4450      4460      4470      4480

                >< MaeIII
>< SfcI          >< Fnu4HI          >< MunI
                >< AluI          >< AluI          >< AciI          MaeIII ><
CTGTAGCTTC TATTATTACG AAGCTGAACT CTCTAAATGA GCCGCTTGTC ACAATGCCAA TTGGTTATGT
      4490      4500      4510      4520      4530      4540      4550

                >< ThaI
                >< MvnI
                >< MboII
                >< HinPII
                >< HinPII
                >< Hin6I
                >< Hin6I
                >< HhaI
                >< HhaI
                >< Tru9I          >< HhaI
>< NlaIII          >< Fnu4HI
                >< MseI          >< CfoI
                >< MnlI          >< CfoI
                >< Ksp632I          >< BstUI
                >< EarI          >< BssHII>< BspWI          >< Tru9I
                >< Eam1104I          >< Bsp50I          >< MseI
                >< BbvI          >< AccII          >< AluI          HphI ><
GACACATGGT TTTAATCTTG AAGAGGCTGC GCGCTGTATG CGTTCTCTTA AAGCTCCTGC CGTAGTGTC
      4560      4570      4580      4590      4600      4610      4620

                >< MaeIII
>< SfaNI          >< AlwNI          >< MnlI >< MnlI>< DdeI
GTATCATCAC CAGATGCTGT TACTACATAT AATGGATACC TCACTTCGTC ATCAAAGACA TCTGAGGAGC
      4630      4640      4650      4660      4670      4680      4690

                >< SinI
                >< Sau96I
                >< NspIV
                >< NspHII
>< SduI          >< Eco47I
>< NspII          >< Cfr13I
>< HgiAI          >< BsiZI
>< Bsp1286I          >< Bme18I          >< RsaI
>< BmyI          >< AvaII          >< Csp6I
>< Alw21I          >< AsuI          >< AfaI
ACTTTGTAGA AACAGTTTCT TTGGCTGGCT CTTACAGAGA TTGGTCCTAT TCAGGACAGC GTACAGAGTT
      4700      4710      4720      4730      4740      4750      4760

                > < TthHB8I
                > < TaqI
                >< SduI
                >< Van91I          >< NspII
                >< RsaI          >< PflMI          >< Eco24I
                >< HphI          >< BslI          >< Bsp1286I
                >< Csp6I          >< BsiYI          >< BmyI          >< GsuI ><

>< Tru9I
>< MseI
>< Esp4I

```

FIGURE 13.12

```

    >> AflIII >> MaeIII >> AfaI >> AccB7I >> BaniIBpmI >>
AGGTGTTGAA TTTCTTAAGC GTGGTGACAA AATTGTGTAC CACACTCTGG AGAGCCCCGT CGAGTTTCAT
    4770      4780      4790      4800      4810      4820      4830

    >> Tru9I
    >> PleI >> EcoNI
    >> MnlI >> BslI
    >> BsmAI >> BsiYI
    >> MnlI >> HphI >> HinfI >> Alw26I >> AciI >> MseI
CTTGACGGTG AGGTTCCTTC ACTTGACAAA CTAAGAGTC TCTTATCCCT GCGGGAGGTT AAGACTATAA
    4840      4850      4860      4870      4880      4890      4900

    >> AluI >> NdeI
AAGTGTTCAC AACTGTGGAC AACACTAATC TCCACACACA GCTTGTGGAT ATGTCTATGA CATATGGACA
    4910      4920      4930      4940      4950      4960      4970

    >> SniI
    >> Sau96I
    >> NspIV
    >> NspHII
    >> Eco47I
    >> CfrI3I
    >> Bsi2I
    >> BmeI8I
    >> AvaII
    >> AsuI
    >> MaeIII >> Tru9I >> MnlI
    >> FokI >> MseI >> BspHI
GCAGTTTGGT CCAACATACT TGGATGGTGC TGATGTTACA AAAATTAAAC CTCATGTAAA TCATGAGGGT
    4980      4990      5000      5010      5020      5030      5040

    >> RsaI
    >> RmaI
    >> MaeI
    >> Csp6I
    >> AfaI
    >> SnaBI
    >> MaeII >> HindIII >> RsaI
    >> Eco105I >> Csp6I
    >> BsaAI >> AluI >> AfaI
AAGACTTTCT TTGTACTACC TAGTGATGAC ACACTACGTA GTGAAGCTTT CGAGTACTAC CATACTCTTG
    5050      5060      5070      5080      5090      5100      5110

    >> RsaI
    >> NspI
    >> NspHI
    >> NlaIII
    >> Csp6I >> Tru9I
    >> AflIII >> MseI
    >> AfaI >> DraI
    >> MnlI >> BslI >> BsiYI >>
ATGAGAGTTT TCTTGCTAGG TACATGTCTG CTTTAAACCA CACAAAGAAA TGGAATTTTC CTCAAGTTGG
    5120      5130      5140      5150      5160      5170      5180

    >> Tru9I >> Tru9I
    >> MseI >> MseI
    >> MunI >> RmaI
    >> MaeI >> AluI >>
TGGTTTAACT TCAATTAAAT GGGCTGATAA CAATTGTTAT TTGTCTAGTG TTTTATTAGC ACTTCAACAG
    5190      5200      5210      5220      5230      5240      5250

    >> SfaNI
    >> SduI
    >> NspII
    >> Eco24I
    >> Bsp1286I
    >> BmyI >> HphI >>
    >> BbvI >> Fnu4HI >>
    >> MnlI >> BanII >> BspWI

```

FIGURE 13.13

```

CTTGAAGTCA AATTCAATGC ACCAGCACTT CAAGAGGCTT ATTATAGAGC CCGTGCTGGT GATGCTGCTA
5260      5270      5280      5290      5300      5310      5320

>< VneI
>< SnaI
    >< SduI
    >< NspII
    >< HgiAI
    >< Bsp1286I
    >< BmyI
>< ApaLI
>< Alw44I
    >< Alw21I
    >< AluI
    MboII ><
    >< HphI
ACTTTTGTGC ACTCATACTC GCTTACAGTA ATAAACTGT TGGCGAGCTT GGTGATGTCA GAGAACTAT
5330      5340      5350      5360      5370      5380      5390

    > < SphI
    > < PaeI
    > < NspI
    > < NspHI >< TfiI
    >< Tru9I
    >< SfcI > < NlaIII> < HinfI
    >< MseI
GACCCATCTT CTACAGCATG CTAATTTGGA ATCTGCAAAG CGAGTTCTTA ATGTGGTGTG TAAACATTGT
5400      5410      5420      5430      5440      5450      5460

    >< RsaI
    >< Tru9I
    > < Csp6I
    Esp4I >
    >< MseI
    >< AluI
    >< AfaI
    AflIII >
GGTCAGAAAA CTACTACCTT AACGGGTGTA GAAGCTGTGA TGTATATGGG TACTCTATCT TATGATAATC
5470      5480      5490      5500      5510      5520      5530

    >< RsaI
    >< MboII
    >< RmaIHinfI ><
    >< Csp6I
    >< MaeI >< BbsI
    >< AfaI
>< Tru9I
    >< SfaNI
>< MseI
    >< NlaIII
TTAAGACAGG TGTTCATT CCATGTGTGT GTGGTCGTGA TGCTACACAA TATCTAGTAC AACAGAGTC
5540      5550      5560      5570      5580      5590      5600

    >< RsaI
    >< Csp6I
    >< AfaI
    >< PleI
    > < DdeI
    >< BspWI >< BspMI
    >< BsgI
    >< BspI
TTCTTTTGTG ATGATGTCTG CACCACCTGC TGAGTATAAA TTACAGCAAG GTACATTCTT ATGTGCGAAT
5610      5620      5630      5640      5650      5660      5670

    >< RsaI
    >< Eco31I
    >< DdeI
    >< BsmAI
    >< BsaI
    MnlI ><
    >< AfaI >< BsrI
    >< Alw26I
    HphI >
GAGTACACTG GTAACATCA GTGTGGTCAT TACACTCATA TAACTGCTAA GGAGACCCCTC TATCGTATTG
5680      5690      5700      5710      5720      5730      5740

    >< SstI
    >< SnuI
    >< SacI
    >< NspII
    >< HgiAI
    >< Eco24I
    >< Eco47I
    >< Ecl136II
    >< Bsp1286I
    >< BsiZI
    >< BmyI
    >< SinI
    >< Sau96I
    >< NspIV
    >< NspHII
    > < RsaI
    >< MaeIII
    >< Cfr13I
    >< Bme18I

```

FIGURE 13. 14


```

    >< BanII
    >< Alw21I
    >< AluI
    ACGGAGCTCA CCTTACAAAG ATGTCAGAGT ACAAAGGACC AGTGACTGAT GTTTTCTACA AGGAAACATC
    5750      5760      5770      5780      5790      5800      5810

    >< TthHB8I
    >< TaqI >< MaeIII
    TTACACTACA ACCATCAAGC CTGTGTCGTA TAAACTCGAT GGAGTTACTT ACACAGAGAT TGAACCAAAA
    5820      5830      5840      5850      5860      5870      5880

    >< RsaI
    >< Csp6I
    >< SfcI >< BbvI
    >< FokI >< Fnu4HI >< AfaI
    TTGGATGGGT ATTATAAAAA GGATAATGCT TACTATACAG AGCAGCCTAT AGACCTTGTA CCAACTCAAC
    5890      5900      5910      5920      5930      5940      5950

    Tru9I ><
    SwaI ><
    MseI ><
    >< NspI
    >< NspHI
    >< NlaIII
    >< AflIII
    CATTACCAAA TGCGAGTTT GATAATTCA AACTCACATG TTCTAACACA AAATTTGCTG ATGATTTAAA
    5960      5970      5980      5990      6000      6010      6020

    >< MboII
    >< AluI >< AluI >< MaeIII
    TCAAATGACA GGCTTCACAA AGCCAGCTTC ACGAGAGCTA TCTGTCACAT TCTTCCGAGA CTTGAATGGC
    6030      6040      6050      6060      6070      6080      6090

    >< SfcI
    GATGTAGTGG CTATTGACTA TAGACACTAT TCAGCGAGTT TCAAGAAAGG TGCTAAATTA CTGCATAAGC
    6100      6110      6120      6130      6140      6150      6160

    >< Tru9I
    >< ScrFI
    >< MvaI
    >< MseI
    >< EcoRII
    >< Ecl136I
    >< DsaV
    >< BstOI
    >< BstNI
    >< BsiLI
    >< MunI
    >< BstXI
    >< ApyI
    >< MaeII
    >< BstXI
    CAATTGTTTG GCACATTAAC CAGGCTACAA CCAAGACAAC GTTCAAACCA AACACTTGGT GTTTACGTTG
    6170      6180      6190      6200      6210      6220      6230

    >< RsaI
    >< Csp6I
    >< AfaI >< BsrI
    TCTTTGGAGT ACAAAGCCAG TAGATACTTC AAATTCATTG GAAGTTCTGG CAGTAGAAGA CACACAAGGA
    6240      6250      6260      6270      6280      6290      6300

    >< HindII
    >< HincII
    >< MboII
    >< MnlI
    >< Eco57I
    ATGGACAATC TTGCTTGTGA AAGTCAACAA CCCACCTCTG AAGAAGTAGT GGAAAATCCT ACCATACAGA
    6310      6320      6330      6340      6350      6360      6370

```

FIGURE 13.15

```

    >< MaeIII
    >< MaeII
AGGAAGTCAT AGAGTGTGAC GTGAAACTA CCGAAGTTGT AGGCAATGTC ATACTTAAAC CATCAGATGA
6380      6390      6400      6410      6420      6430      6440

    >< XhoII
    >< Sau3AI
    >< NlaIII
    >< NdeII
    >< MflI
    >< MboI
    >< DpnII
    >< DpnI
    >< BstYI
    >< BspAI
    >< Tru9I
    >< MseI
    >< BspHI >< Bsp143I>< Fnu4HI
    >< MaeIII >< MnlI >< BbvI >< AlwI
AGGTGTTAAA GTAACACAAG AGTTAGGTCA TGAGGATCTT ATGGCTGCTT ATGTGGAAAA CACAAGCATT
6450      6460      6470      6480      6490      6500      6510

    >< SauI
    >< RmaI
    >< MstII
    >< MaeI
    >< Eco81I
    >< DdeI
    >< CvnI
    >< Bsu36I
    >< Bse21I
    >< BfrI> < Tru9I
    >< Tru9I >< AxyI> < MseI>< MunI >< NlaIII
    >< MseI >< AluI >< AocI >< DraI >< BbvI Fnu4HI ><
ACCATTAAGA AACCTAATGA GCTTTCAC TAAGGCTT TAAAAACAAT TGCCACTCAT GGTATTGCTG
6520      6530      6540      6550      6560      6570      6580

    >< VspI >< StyI
    >< Tru9I >< EcoT14I
    >< MseI >< Eco130I
    >< AsnI >< BssT1I
    >< AseI >< BsaJI
    >< BfrI >< Fnu4HI
CAATTAATAG TGTTCCTTGG AGTAAATTT TGGCTTATGT CAAACCATTC TTAGGACAAG CAGCAATTAC
6590      6600      6610      6620      6630      6640      6650

    >< HinPII
    >< Hin6I
    >< HhaI
    >< DdeI
    >< BbvI
    >< CfoI
    >< Tru9I
    >< MaeII>< MseI
    >< DraIII
    >< AflIII
AACATCAAAT TGCCTAAGA GATTAGCACA ACGTGTGTTT AACAATTATA TGCCTTATGT GTTTACATTA
6660      6670      6680      6690      6700      6710      6720

    >< RsaI
    >< Csp6I
    >< MunI >< AfaI
    >< RsaI>< XbaI
    >< Csp6I >< RmaI
    >< AfaI >< MaeI
    >< AluI
TTGTTCCAAT TGTGTACTTT TACTAAAAGT ACCAATTCTA GAATTAGAGC TTCCTACCT ACAACTATTG
6730      6740      6750      6760      6770      6780      6790

    >< VspI
    >< Tru9I
    >< NaeI
    >< MspI
    >< MseI

```

FIGURE 13. 16

```

                                >< HpaII
                                >< HapII
                                >< Cfr10I >< FokI
                                >< AsnI
                                >< Tru9I
                                >< DdeI   MaeIII >
                                >< MseI   >< BfrI   >< BbvI
                                >< SfaNI   >< AseI>< HphI>< MaeIII
CTAAAAATAG TGTTAAGAGT GTTGCTAAAT TATGTTTGGG TGCCGGCATT AATTATGTGA AGTCACCCAA
6800      6810      6820      6830      6840      6850      6860

                                >< Tru9I   >< DdeI   MaeIII >
                                >< MseI   >< BfrI   >< BbvI
ATTTTCTAAA TTGTTACAA TCGCTATGTG GCTATTGTTG TTAAGTATTT GCTTAGGTTT TCTAATCTGT
6870      6880      6890      6900      6910      6920      6930

                                >< SduI
                                >< NspII
                                >< HgiAI
                                >< Bsp1286I
                                >< BmyI
                                >< Alw21I
                                > < RsaI
                                >< Csp6I
                                >< Fnu4HI > < AfaI
GTAAGTGTG CTTTGGTGT ACTCTTATCT AATTTTGGTG CTCCTTCTTA TTGTAATGGC GTTAGAGAAT
6940      6950      6960      6970      6980      6990      7000

                                Tru9I ><
                                MseI ><
                                >< Tru9I   > < MaeIII
                                >< MseI   >< MaeII
                                >< Fnu4HI
                                >< BbvI >
TGTATCTTAA TTCGTCTAAC GTTACTACTA TGGATTCTG TGAAGGTCTT TTCCTTGCA GCATTTGTTT
7010      7020      7030      7040      7050      7060      7070

                                > < TfiI
                                >< MamI
                                >< HinfI
                                >< BsiBI
                                >< XmnI>< MaeIII
                                >< Asp700I
                                >< AluI >
                                >< PleI>< HinfI
                                >< BsaBI >< AluI
                                >< Asp700I
                                >< AfaI ><
AAGTGGATTA GACTCCCTTG ATTCTTATCC AGCTCTTGAA ACCATTGAGG TGACGATTTC ATCGTACAAG
7080      7090      7100      7110      7120      7130      7140

                                >< PstI
                                >< NspBII
                                >< HaeIII
                                >< GdiII
                                >< Fnu4HI
                                >< EaeI
                                >< DdeI
                                >< BsuRI
                                >< BshI >< BslI
                                >< AciI>< BsiYI
>< RmaI
>< MaeI
CTAGACTTGA CAATTTTAGG TCTGGCCGCT GAGTGGGTTT TGGCATATAT GTTGTTTACA AAATTCTTTT
7150      7160      7170      7180      7190      7200      7210

                                >< BspMI
                                >< RmaI
                                >< AluI
                                >< MaeI
ATTATTAGG TCTTTCAGCT ATAATGCAGG TGTTCTTTGG CTATTTTGCT AGTCATTTC TCAGCAATTC
7220      7230      7240      7250      7260      7270      7280

                                RsaI ><
                                >< MboII
                                >< NlaIV
                                >< Eco64I
                                >< RsaI >< BscBI
                                >< Csp6I >< Bani
                                > < NlaIII
                                > < AfaI>< AccBII
                                >< Bsp6I ><
                                >< BsiBI ><
                                >< BsaBI ><
                                >< AfaI ><

```

FIGURE 13.17

```

TTGGCTCATG TGGTTTATCA TTAGTATTGT ACAAATGGCA CCCGTTTCTG CAATGGTTAG GATGTACATC
7290      7300      7310      7320      7330      7340      7350

                                TthHB8I >>
                                >> TaqI
                                MnlI >>
                                >> NdeI
                                >> Ksp632I
                                >> EarI
                                >> MboII EarI >>
                                >> FokI
                                >> Eam1104I >> AluI >> MboII >> NlaIII Eam1104I >>
>> FokI
TTCTTTGCTT CTTTCTACTA CATATGGAAG AGCTATGTTT ATATCATGGA TGGTTGCACC TCTTCGACTT
7360      7370      7380      7390      7400      7410      7420

                                XhoII >>
                                Sau3AI >>
                                NlaIII >>
                                NdeII >>
                                MflI >>
                                MboI >>
                                >> ThaI
                                >> MvnI
                                >> Ksp632I
                                >> EarI
                                >> Eam1104I
>> HinPII >> MluI >> BstUI >> DpnII >>
>> Hin6I >> Bst50I >> RsaI >> BstYI >>
>> HhaI >> AflIII >> Csp6I >> Tru9I BspAI >>
>> NlaIII >> CfoI >> AccII >> AfaI >> MseI BglII >>
>> BspWI >> BspWI
GCATGATGTG CTATAAGCGC AATCGTGCCA CACGCGTTGA GTGTACAAC ATTGTTAATG GCATGAAGAG
7430      7440      7450      7460      7470      7480      7490

                                >> Pali
                                >> HaeIII
                                >> DsaI
                                >> MnlI
>> MboII >> BsuRI >> BshI >> MunI >> MaeIII >>
>> DpnI >> Bsp143I >> MnlI >> BsaJI >> PleI >> HinfI >> Alw26I >>
ATCTTTCTAT GTCTATGCAA ATGGAGGCCG TGGCTTCTGC AAGACTCACA ATTGGAATTG TCTCAATTGT
7500      7510      7520      7530      7540      7550      7560

                                >> RsaI
                                >> Csp6I
                                >> BsrI
                                >> AfaI
                                >> GsuI
                                >> BpmI
                                >> MaeIIIDraI >>
                                >> BsrI
                                >> Tru9I >>
                                >> MseI >>
                                >> BsrI
                                >> BssHII
                                >> Bsp50I >>
                                >> BsrI
                                >> AccII
GACCAATCAA CCCTACTGAC CAGTCATCGT ATATTGTTGA TAGTGTTGCT GTGAAAAATG GCGCGCTTCA
7640      7650      7660      7670      7680      7690      7700

```

FIGURE 13. 18

```

                                >< FokI
                                >< BsmAI
                                >< MnlI
                                >< Alw26I
                                >< AclI
CCTCTACTTT GACAAGGCTG GTCAAAAGAC CTATGAGAGA CATCCGCTCT CCCATTTTGT CAATTTAGAC
7710      7720      7730      7740      7750      7760      7770

                                >< VspI
                                >< Tru9I
                                >< MseI
                                >< AsnI
                                >< AseI
                                >< BcgI/a
> < AluI
AATTTGAGAG CTAACAACAC TAAAGGTTCA CTGCCTATTA ATGTCATAGT TTTTGATGGC AAGTCCAAAT
7780      7790      7800      7810      7820      7830      7840

                                >< SfcI
                                >< PvuII
                                >< RsaI
                                >< Psp5I
                                >< Csp6I
                                >< NspBII
                                >< PleI
                                >< DdeI
                                >< BcgI
                                >< AfaI
                                >< AluI
GCGACGAGTC TGCTTCTAAG TCTGCTTCTG TGTACTACAG TCAGCTGATG TGCCAACCTA TTCTGTTGCT
7850      7860      7870      7880      7890      7900      7910

                                TthHB8I ><
                                TaqI ><
                                SalI ><
                                RtrI ><
                                HindII >
                                HincII >
                                >< ScaI
                                >< RsaI
                                >< Tru9I
                                >< Csp6I
                                >< SfaNI >< Eco57I
                                >< AluI
                                >< MaeII
                                >< AfaI
                                >< MseI
                                >< AccI ><
TGACCAAGCT CTTGTATCAG ACGTTGGAGA TAGTACTGAA GTTCCGTTA AGATGTTTGA TGCTTATGTC
7920      7930      7940      7950      7960      7970      7980

                                >< Tru9I
                                >< MseI
                                > < Esp4I
                                > < AflIII
                                >< SfcI
                                >< BspWI
                                >< AluI
GACACCTTTT CAGCAACTTT TAGTGTTCCT ATGGAAAAAC TTAAGGCACT TGTTGCTACA GCTCACAGCG
7990      8000      8010      8020      8030      8040      8050

                                >< PvuII
                                >< Psp5I
                                >< NspBII
                                >< Fnu4HI
                                >< AluI
                                >< BbvI
AGTTAGCAAA GGGTGTAGCT TTAGATGGTG TCCTTTCTAC ATTCTGTGTA GCTGCCCGAC AAGGTGTTGT
8060      8070      8080      8090      8100      8110      8120

                                >< HindII
                                >< BsmAI
                                >< DdeI
                                >< HincII
                                >< FokI >< Alw26I
                                >< BfrI
TGATACCGAT GTTGACACAA AGGATGTTAT TGAATGTCTC AAACCTTCAC ATCACTCTGA CTAGAAGTG
8130      8140      8150      8160      8170      8180      8190

                                >< XhoII
                                Sau3AI ><
                                >< NdeII
                                >< MflI
                                >< MboI
                                >< NlaIII >< HgaI
                                >< HinII >< DpnII
                                DpnI ><

```

FIGURE 13.19

```

                                Bsp143I ><
                                >< BsaHI >< BstYI
                                >< BbiII >< BspAI
                                >< AcyI >< BglII
                                >< MaeIII>< HphI
                                >< MaeIII >< HphI >< NlaIII
ACAGGTGACA GTTGTAACAA TTTCATGCTC ACCTATAATA AGGTTGAAAA CATGACGCCC AGAGATCTTG
      8200      8210      8220      8230      8240      8250      8260

                                >< NspI
                                >< NspHI
                                >< NlaIII
>< HinfII
>< HinfI
                                >< HhaI
                                >< CfoI
                                >< BspWI >< MaeIII
GCGCATGTAT TGA CTGTAAT GCAAGGCATA TCAATGCCCA AGTAGCAAAA AGTCACAATG TTTCACATCAT
      8270      8280      8290      8300      8310      8320      8330

                                >< NspI
                                >< NspHI
                                >< NlaIII
                                >< PvuII
                                >< Psp5I
                                >< Eam1105I
                                >< NspBII
                                >< BbvI
                                >< Fnu4HI
                                >< AflIII
                                >< AluI >< BbvI >< Fnu4HI
CTGGAATGTA AAAGACTACA TGTCTTTATC TGAACAGCTG CGTAAACAAA TTCGTAGTGC TGCCAAGAAG
      8340      8350      8360      8370      8380      8390      8400

                                >< RmaI
                                >< MaeI >< Eam1105I
                                >< MboII
AACACATAC CTTT TAGACT AACTTGTGCT ACAACTAGAC AGGTGTCAA TGTCATACT ACTAAATCT
      8410      8420      8430      8440      8450      8460      8470

                                >< Tru9I
                                >< Pali
                                >< MseI
                                >< HaeIII
                                >< ScaI
                                >< Esp4I
                                >< RsaI >< Tru9I
                                >< BsuRI
                                >< Csp6I >< MseI
                                >< BshI
                                >< AfaI >< DraI >< AflIII >< BbvI
CACTCAAGGG TGTAAGATT GTTAGTACTT GTTTAACT TATGCTTAAG GCCACATTAT TGTGCGTTCT
      8480      8490      8500      8510      8520      8530      8540

                                >< RsaI
                                >< Csp6I
                                >< BsrI
                                >< NlaIII
                                >< Fnu4HI
                                >< AfaI
                                >< MaeIII
TGCTGCATTG GTTTGTTATA TCGTTATGCC AGTACATACA TTGTCAATCC ATGATGGTTA CACAAATGAA
      8550      8560      8570      8580      8590      8600      8610

                                >< MaeIII
                                >< MaeIII
                                >< FokI
ATCATTGGTT ACAAAGCCAT TCAGGATGGT GTCACCTCGTG ACATCATTTT TACTGATGAT TGTTTTGCAA
      8620      8630      8640      8650      8660      8670      8680

                                SfcI >
                                Fnu4HI ><
                                BbvI ><
                                >< NspI
                                >< NspHI
                                >< NlaIII
                                >< HgaI
                                >< BstXI
                                >< BbvI
                                >< AluI
ATAAACATGC TGGTTTGGAC GCATGGTTTA GCCAGCGTGG TGGTTCATAC AAAAATGACA AAAGCTGCCC
      8690      8700      8710      8720      8730      8740      8750

```

FIGURE 13. 20

```

                                >< ScrFI
                                >< ScrFI >< RsaI
                                >< MvaI >< MspI
                                >< EcoRII >< HpaII
                                >< Ecl136I>< NciI
                                >< DsaV >< HapII
                                >< BstOI>< DsaV
                                >< BstNI >< Csp6I
                                >< BsiLI >< BcnIDdeI ><
                                >< ApyI >< AfaI
                                >< Fnu4HI
                                >< AluI
TGTAGTAGCT GCTATCATT CAAGAGAGAT TGGTTTCATA GTGCCTGGCT TACCGGGTAC TGTGCTGAGA
8760 8770 8780 8790 8800 8810 8820

                                > < MaeIII >< HphI >< MnlI >< BspWI
GCAATCAATG GTGACTTCTT GCATTTTCTA CCTCGTGTTC TTAGTGCTGT TGGCAACATT TGCTACACAC
8830 8840 8850 8860 8870 8880 8890

                                Tru9I >
                                SfaNI ><
                                >< RsaI
                                MseI >
                                >< BspWI >< Fnu4HI >< Csp6I
                                >< BbvI>< MnlI >< DdeI >< AfaI
CTTCCAACT CATTGAGTAT AGTGATTTTG CTACCTCTGC TTGCGTTCCT GCTGCTGAGT GTACAATTTT
8900 8910 8920 8930 8940 8950 8960

                                > < RmaI
                                >< MnlI
                                >< FokI
                                > < MaeI
TAAGGATGCT ATGGGCAAAC CTGTGCCATA TTGTATGAC ACTAATTTGC TAGAGGGTTC TATTTCTTAT
8970 8980 8990 9000 9010 9020 9030

                                ScrFI >
                                MvaI >
                                MnlI ><
                                EcoRII ><
                                Ecl136I >
                                DsaV ><
                                BstOI >
                                >< NlaIV BstNI >
                                >< FokI BsiLI >
                                >< BscBI ApyI >
                                >< AluI
AGTGAGCTTC GTCCAGACAC TCGTTATGTG CTTATGGATG GTTCCATCAT ACAGTTTCCT AACACTTACC
9040 9050 9060 9070 9080 9090 9100

                                >< RsaI
                                >< SfcI >< NspI
                                >< ScaI >< NspHI
                                >< RsaI >< NlaIII
                                > < MaeIII >< Csp6I >< NlaIII
                                >< GsuI >< AfaI >< Csp6I
                                >< BpmI >< DdeI >< AccI >< AfaI
TGGAGGGTTC TGTTAGAGTA GTAACAACTT TTGATGCTGA GTACTGTAGA CATGGTACAT GCGAAAGGTC
9110 9120 9130 9140 9150 9160 9170

                                >< SstI
                                >< SduI
                                >< SacI
                                NspII ><
                                HgiAI ><
                                Eco24I ><
                                Bspl286I ><

```

FIGURE 13.21

```

                                Ecl136II ><>< BmyI
                                BanII ><
                                >< Tru9I
                                Alw21I ><
                                >< BsrI
                                >< MseI
                                >< AluI
AGAAGTAGGT ATTTGCCTAT CTACCAGTGG TAGATGGGTT CTTAATAATG AGCATTACAG AGCTCTATCA
  9180          9190          9200          9210          9220          9230          9240

                                >< TfiI
                                >< SfaNI
                                >< HinfI
                                >< AluI
                                >< MnlI
GGAGTTTTCT GTGGTGTGA TGCGATGAAT CTCATAGCTA ACATCTTTAC TCCTCTTG TG CAACCTGTGG
  9250          9260          9270          9280          9290          9300          9310

                                >< MaeIII
                                HphI ><
                                >< Eco57I
                                >< BbvI
                                Fnu4HI ><
GTGCTTTAGA TGTGTCTGCT TCAGTAGTGG CTGGTGGTAT TATTGCCATA TTGGTGACTT GTGCTGCCTA
  9320          9330          9340          9350          9360          9370          9380

                                >< RsaI
                                >< Csp6I
                                >< NlaIII
                                >< MaeII
                                >< BbvI
                                >< Fnu4HI
                                >< AflIII
                                >< AfaI>< HphI
                                >< BspWI
CTACTTTATG AAATTCAGAC GTGTTTTTGG TGAGTACAAC CATGTTGTG CTGCTAATGC ACTTTGTTT
  9390          9400          9410          9420          9430          9440          9450

                                >< RsaI
                                >< NlaIV
                                >< KpnI
                                >< Eco64I
                                >< Csp6I
                                >< BscBI
                                >< Asp718
                                >< BanI >< AluI
                                >< AfaI
                                >< AccB1I
                                >< Acc65I
                                >< ScrFI
                                >< NciI
                                >< MspI
                                >< HpaII
                                >< HinfI
                                >< HapII
                                >< PleI
                                >< BcnI
                                >< DdeI
                                >< AluI>< DsaV
                                >< AccI
TTGATGTCTT TCACTATACT CTGTCTGGTA CCAGCTTACA GCTTTCTGCC GGGAGTCTAC TCAGTCTTTT
  9460          9470          9480          9490          9500          9510          9520

                                >< RsaI
                                >< Csp6I
                                >< AfaI
                                >< HphI
                                >< HphI
                                NlaIII ><
ACTTGTA CTTT GACATTCTAT TTCACCAATG ATGTTTCATT CTTGGCTCAC CTTCAATGGT TTGCCATGTT
  9530          9540          9550          9560          9570          9580          9590

TTCTCCTATT GTGCCTTTTT GGATAACAGC AATCTATGTA TTCTGTATTT CTCTGAAGCA CTGCCATGG
  9600          9610          9620          9630          9640          9650          9660

                                >< TthHB8I
                                >< RsaI
                                >< MnlI
                                >< MnlI
                                >< Csp6I
                                >< Tru9I
                                >< PleI
                                >< BcgI/a
                                >< TaqI
                                >< MseI
                                >< DdeI
                                >< NlaIII
                                >< BbvI
                                >< Eco57I
                                >< BfrI
                                >< HinfI
                                >< MseI
                                >< MaeIII
                                >< AfaI
                                Fnu4HI ><
TTCTTTAACA ACTATCTTAG GAAAAGAGTC ATGTTTAAATG GAGTTACATT TAGTACCTTC GAGGAGGCTG
  9670          9680          9690          9700          9710          9720          9730

                                >< RsaI
                                >< Csp6I
                                >< BcgI
                                >< RsaI
                                >< Csp6I
                                >< BsmAI

```

FIGURE 13.22


```

    >> AfaI          >> AfaI          >> Alw26I
CTTTGTGTAC CTTTGTGCTC AACAAGGAAA TGTACCTAAA ATTGCGTAGC GAGACACTGT TGCCACTTAC
    9740          9750          9760          9770          9780          9790          9800

                                >> NlaIV
                                >> DdeI
                                >> BscBI
                                >> BfrI   AluI >>
ACAGTATAAC AGGTATCTTG CTCTATATAA CAAGTACAAG TATTTCAAGT GAGCCTTAGA TACTACCAGC
    9810          9820          9830          9840          9850          9860          9870

    >> Fnu4HI
    >> DdeI
    >> Fnu4HI   >> BfrI
    >> BbvI   >> AluI   >> BbvI          >> DdeI >> AlwNI
TATCGTGAAG CAGCTTGCTG CCACTTAGCA AAGGCTCTAA ATGACTTTAG CAACTCAGGT GCTGATGTTT
    9880          9890          9900          9910          9920          9930          9940

                                >> SfcI          >> BsmI
                                >> PstI          >> BscCI
TCTACCAACC ACCACAGACA TCAATCACTT CTGCTGTTCT GCAGAGTGGT TTTAGGAAAA TGGCATTCCC
    9950          9960          9970          9980          9990          10000          10010

    >> RsaI
    >> NlaIII
    >> MaeIII
    >> Csp6I          >> Tru9I
    >> AfaI          >> MseI
GTCAGGCAAA GTTGAAGGGT GCATGGTACA AGTAACCTGT GGAAGTACAA CTCTTAATGG ATTGTGGTTG
    10020          10030          10040          10050          10060          10070          10080

                                XhoII >
                                Sau3AI >
                                >> Tru9I   NdeII >
                                >> NspI     MflI >
                                >> NspHI    MboI >
                                >> NlaIII   DpnII >
                                >> MseI     BstYI >
                                >> MboII   BspAI >
                                >> BbsI     BglII >
GATGACACAG TATACTGTCC AAGACATGTC ATTTGCACAG CAGAAGACAT GCTTAATCCT AACTATGAAG
    10090          10100          10110          10120          10130          10140          10150

                                Pali >
                                MscI >
                                HaeIII >
                                EaeI >
                                BsuRI >
                                BshI >
                                Bali >
>> DpnI >> MboII
>> Bsp143I          >> AluI
ATCTGCTCAT TCGCAAATCC AACCATAGCT TTCTTGTTCA GGCTGGCAAT GTTCAACTTC GTGTATTGG
    10160          10170          10180          10190          10200          10210          10220

                                >> DdeI > > Tru9I
                                >> BfrI > > MseI          >> DdeI
CCATTCTATG CAAAATTGTC TGCTTAGGCT TAAAGTTGAT ACTTCTAACC CTAAGACACC CAAGTATAAA
    10230          10240          10250          10260          10270          10280          10290

    >> ScrFI
    >> MvaI
    >> EcoRII
    >> Ecl136I          >> SphI

```

FIGURE 13.23

```

>< DsaV
>< BstOI
>< BstNI
>< BsiLI
>< ApyI
TTGTGTCGTA TCCAACCTGG TCAAACATTT TCAGTTCTAG CATGCTACAA TGGTTCACCA TCTGGTGTTC
10300 10310 10320 10330 10340 10350 10360

>< PaeI
>< NspI
>< NspHI
>< RmaI >< NlaIII
>< MaeI >< HphI

>< Sau3AI
>< NdeII
>< MboI>< NlaIII
>< DpnII
>< Eco3II
>< BsmAI
>< BsaI>< NlaIII
>< Alw26I
ATCAGTGTGC CATGAGACCT AATCATACCA TTAAAGGTTT TTTCCCTAAT GGATCATGTG GTAGTGTGTTG
10370 10380 10390 10400 10410 10420 10430

>< Tru9I
>< MseI
>< Tru9I>< DpnI
>< MseI >< BspI43I
>< BspAI>< AlwI

>< Zsp2I
>< PpuI0I
>< NsiI>< SfaNI
>< NdeI
>< MphI103I
>< EcoT22I
>< AvaIII >< AluI AfaI ><
>< Tru9I
>< MseI
>< Tru9I >< AluI AfaI ><
TTTAAACATT GATTATGATT GCGTGTCTTT CTGCTATATG CATCATATGG AGCTTCCAAC AGGAGTACAC
10440 10450 10460 10470 10480 10490 10500

>< SinI
>< Sau96I
>< NspIV
>< NspHII
>< Eco47I
>< CfrI13I
>< BsiZI
>< BmeI18I >< HindII
>< AvaII >< HincII
>< AsuI>< BsgI >< BbvI >< BspMI AfaI ><
>< RsaI
>< Csp6I>< DdeI
>< AfaI>< BfrI
GCTGGTACTG ACTTAGAAGG TAAATTCTAT GGTCCATTG TTGACAGACA AACTGCACAG GCTGCAGGTA
10510 10520 10530 10540 10550 10560 10570

>< Tru9I
>< MseI
>< BbvI
>< Fnu4HI
>< HphI ><
CAGACACAAC CATAACATTA AATGTTTTGG CATGGCTGTA TGCTGCTGTT ATCAATGGTG ATAGGTGGTT
10580 10590 10600 10610 10620 10630 10640

>< Tru9I
>< TfiI
>< MseI
>< HphI
>< HinfI
>< RsaI
>< Csp6I
>< AfaI
TCTTAATAGA TTCACCACTA CTTTGAATGA CTTTAACCTT GTGGCAATGA AGTACAACCTA TGAACCTTTG
10650 10660 10670 10680 10690 10700 10710

>< SinI
>< Sau96I
>< PssI
>< Psp5II
>< PpuMI
>< NspIV
>< NspHII
>< NlaIV

```

FIGURE 13. 24

```

                >> Eco0109I
                >> Eco47I
    >> Sau3AI      >> DraII
    >> NdeII       >> Cfr13I
    >> MboI        >> BsiZI
    >> DpnII>> NlaIII >> BscBI
    >> DpnI >> HindII >> Bme18I
    >> BspAI >> HincII >> AvaII
    >> Bsp143I >> AsuI >> MnlI
ACACAAGATC ATGTTGACAT ATTGGGACCT CTTTCTGCTC AACAGGAAT TGCCGTCTTA GATATGTGTG
10720      10730      10740      10750      10760      10770      10780

                >> StyI
                >> RsaI
                >> EcoT14I
                >> Eco130I
    >> SfcI
    >> Fnu4HI      >> Fnu4HI
    >> BbvI        >> Fnu4HI
    >> BbvI >> AluI >> PstI
    >> BbvI >> AluI >> PstI
CTGCTTTGAA AGAGCTGCTG CAGAATGGTA TGAATGGTCG TACTATCCTT GGTAGCACTA TTTTACAAGA
10790      10800      10810      10820      10830      10840      10850

                >> StyI
                >> EcoT14I
                >> Eco130I
                >> BssTII
    >> MboII
    >> MaeIII>> BsaJI
TGAGTTTACA CCATTTGATG TTGTTAGACA ATGCTCTGGT GTTACCTTCC AAGGTAAGTT CAAGAAAATT
10860      10870      10880      10890      10900      10910      10920

    >> SfaNI
    >> SduI
    >> NspII
    >> Tru9I>> Bsp1286I
    >> MseI >> BmyI
    >> Tru9I>> Bsp1286I
    >> MseI >> BmyI
GTTAAGGGCA CTCATCATTG GATGCTTTTA ACTTTCTTGA CATCACTATT GATTCTTGTT CAAAGTACAC
10930      10940      10950      10960      10970      10980      10990

                >> XmnI
                >> BsmI
                >> BscCI
    >> MaeIII
    >> Asp700I
    >> BbvI
    >> BbvI
AGTGGTCACT GTTTTTCTTT GTTTACGAGA ATGCTTTCTT GCCATTTACT CTTGGTATTA TGGCAATTGC
11000      11010      11020      11030      11040      11050      11060

    >> NspI
    >> NspHI
    >> NlaIII
    >> BspWI >> Fnu4HI>> BspWI >> BscCI
    >> MaeIII
TGCATGTGCT ATGCTGCTTG TTAAGCATAA GCACGCATTC TTGTGCTTGT TTCTGTTACC TTCTCTTGCA
11070      11080      11090      11100      11110      11120      11130

                >> SfaNI
                >> RmaI
    >> NspI
    >> NlaIII
    >> NheI
    >> MaeI
    >> BsiBI
    >> BsaBI
    >> NlaIII
    >> BspWI >> MseI >> AccI >> NspHI>> AluI >> BsaBI >> NlaIII
ACAGTTGCTT ACTTTAATAT GGTCTACATG CCTGCTAGCT GGGTGATGCG TATCATGACA TGGCTTGAAT
11140      11150      11160      11170      11180      11190      11200

```

FIGURE 13.25

```

                                >< Tru9I
                                >< MseI
    > < RmaI                    > < Esp4I
    > < MaeI                    >< Eco57I
                                >< AluI
                                > < AflIII
                                >< AluI
TGGCTGACAC TAGCTTGTCT GGTATATAGGC TTAAGGATTG TGTTATGTAT GCTTCAGCTT TAGTTTGTCT
11210      11220      11230      11240      11250      11260      11270

                                >< RmaI
                                >< MaeII
                                >< MaeI
    > < NlaIII   >< SfaNI   >< Fnu4HI
    >< BspHI >< AluI   >< BbvI   >< AflIII
TATTCTCATG ACAGCTCGCA CTGTTTATGA TGATGCTGCT AGACGTGTTT GGACACTGAT GAATGTCATT
11280      11290      11300      11310      11320      11330      11340

                                >< Sau96I
                                >< Pali
                                >< NspIV
                                >< NlaIII
                                >< HaeIII
                                >< Sau3AI
                                > < DdeI
                                >< NdeII
                                >< Cfr13I
                                >< MboI
                                >< BsuRI
                                >< DpnII
                                >< BsiZI
                                >< DpnI
                                >< BshI
                                >< Bsp143I
                                > < BfrI
    >< AccI
    >< BspAI>< AluI
    >< AsuI
ACACTTGTTC ACAAAGTCTA CTATGGTAAT GCTTTAGATC AAGCTATTTC CATGTGGGCC TTAGTTATTG
11350      11360      11370      11380      11390      11400      11410

                                >< RmaI
                                >< NlaIII
                                >< MaeI>< SfcI
    >< MaeIII   >< MnlI   >< MaeIII   >< AluI>< AluI
CTGTAACCTC TAACTATTCT GGTGTCGTTA CGACTATCAT GTTTTATAGCT AGAGCTATAG TGTTTGTGTG
11420      11430      11440      11450      11460      11470      11480

                                DdeI >
                                >< BsrI
                                >< NlaIII BfrI >
TGTTGAGTAT TACCCATTGT TATTTATTAC TGGCAACACC TTACAGTGTA TCATGCTTGT TTATTGTTTC
11490      11500      11510      11520      11530      11540      11550

                                >< Pali
                                >< HaeIII
                                >< Fnu4HI >< BsuRI
    >< BbvI   >< Fnu4HI   >< BspWI
    >< BbvI   >< BspWI   >< BshI   >< Eco57I >< MaeIII
TTAGGCTATT GTTGCTGCTG CTACTTGGC CTTTCTGTT TACTCAACCG TTACTTCAGG CTTACTCTTG
11560      11570      11580      11590      11600      11610      11620

                                >< ScrFI
                                >< MvaI
                                >< EcoRII
                                >< Ecl136I
                                >< DsaV
                                >< BstOI
                                >< BstNI
                                >< BsiLI
                                > < BsaJI
                                >< BsaJI
    >< Eco31I
    >< BsmAI
    >< BsaI

```

FIGURE 13. 26

```

    >> DrdI >> Alw26I >> ApyI DdeI >>
GTGTTTATGA CTACTTGGTC TCTACACAAG AATTTAGGTA TATGAACTCC CAGGGGCTTT TGCCTCCTAA
11630 11640 11650 11660 11670 11680 11690

    >> Tru9I
    >> MseI
>> SfaNI >< HindIII> < Tru9I
    >> MnlI >< AluI > < MseI > < MnlI > < NlaIII
GAGTAGTATT GATGCTTTCA AGCTTAACAT TAAGTTGTTG GGTATTGGAG GTAAACCATG TATCAAGGTT
11700 11710 11720 11730 11740 11750 11760

    >> VneI
    >> SnaI
    >> SduI
    >> NspII
    >> HgiAI
    >> Bsp1286I
    >> BmyI >> RsaI
    >> RsaI >> ApaLI >> MboII
    >> Csp6I >> Alw44I >> Csp6I DdeI >
    >> AfaI >> MaeII >> Alw21I >> AfaI BfrI >
GCTACTGTAC AGTCTAAAAT GTCTGACGTA AAGTGACAT CTGTGGTACT GCTCTCGGTT CTCAACAAC
11770 11780 11790 11800 11810 11820 11830

    >> NspII> < RsaI
    >> DraIII
    >> SduI>< Csp6I
    >> Bsp1286I
    >> MboII
    >> HinfI >> PfuI >> BmyI > < AfaI >> MboII
TTAGAGTAGA GTCATCTTCT AAATTGTGGG CACAATGTGT ACAACTCCAC AATGATATTC TTCTTGCAAA
11840 11850 11860 11870 11880 11890 11900

    >> TthHB8I
    >> TaqI SfcI >>
    >> HindIII >> MboII >> NlaIII
    >> AluI >< Eco57I >> BspWI AccI >>
AGACACAACCT GAAGCTTTTCG AGAAGATGGT TTCTCTTTTG TCTGTTTTGC TATCCATGCA GGGTGCTGTA
11910 11920 11930 11940 11950 11960 11970

    >> VspI
    >> Tru9I >< Ksp632I
    >> MseI >> TthHB8I >< EarI
    >> AsnI >> TaqI >> MboII >< Eam1104I
    >> AseI>< MnlI >< BcgI/a >< Eco57I >< Eco57I >< BcgI
GACATTAATA GGTGTGCGA GGAAATGCTC GATAACCGTG CTACTCTTCA GGCTATTGCT TCAGAATTTA
11980 11990 12000 12010 12020 12030 12040

    >> StuI
    >> ScrFI
    >> PstI
    >> MvaI>< HaeIII
    >> EcoRII>< Eco147I
    >> Ecl136I
    >> DsaV >> BsuRI
    >> BstOI
    >> BstNI
    >> BspWI
    >> BsiLI
    >> Fnu4HI >> BsaJI >> BshI TfiI >>
    >> NdeI >> BspWI>< MnlI >> BglI >> SfcI HinfI >>
    >> AclI >> ApyI>< AatI >> < AluI

```

FIGURE 13. 27

```

GTTCTTTACC ATCATATGCC GCTTATGCCA CTGCCCAGGA GGCCTATGAG CAGGCTGTAG CTAATGGTGA
12050      12060      12070      12080      12090      12100      12110

      >< XmnI      >< Tru9I      >< SfaNI
      >< HphI      >< MseI      >< DdeI
      >< Asp700I   >< Eco57I   >< BbvI Fnu4HI ><
TTCTGAAGTC GTTCTCAAAA AGTTAAAGAA ATCTTTGAAT GTGGCTAAAT CTGAGTTTGA CCGTGATGCT
12120      12130      12140      12150      12160      12170      12180

                                XhoII ><
                                Sau3AI ><
                                NdeII ><
                                MnlI >
                                >< MnlI
                                >< MflI
                                >< MboI
      > < Sau3AI
      > < NdeII      DpnII ><
      > < MboI      DpnI ><
      > < DpnII      DdeI ><
      >< DpnI      BstYI ><
      >< BspWI      >< RsaIBspAI ><
      > < BspAI      >< Csp6IBsp143I ><
      >< Bsp143I      >< AfaIBglII ><
      >< NlaIII
GCCATGCAAC GCAAGTTGGA AAAGATGGCA GATCAGGCTA TGACCCAAAT GTACAAACAG GCAAGATCTG
12190      12200      12210      12220      12230      12240      12250

                                >< SpeI
                                >< RmaI
      >< MaeIII      >< MboII      >< Eam1104I >< BspWI
      >< MaeI      >< BspWI      >< EarI>< BfrI >< AluI
AGGACAAGAG GGCAAAAGTA ACTAGTGCTA TGCAAACAAT GCTCTTCACT ATGCTTAGGA AGCTTGATAA
12260      12270      12280      12290      12300      12310      12320

                                >< ThaI
                                >< MvnI
      >< HinP1I
      >< Hin6I
      >< HhaI
      >< CfoI
      >< BstUI
      >< Tru9I      >< Bsp50I
      >< MseI      >< AccII      SfcI ><
TGATGCACTT AACAACATTA TCAACAATGC GCGTGATGGT TGTGTTCCAC TCAACATCAT ACCATTGACT
12330      12340      12350      12360      12370      12380      12390

                                >< RsaI
                                >< NlaIV
                                >< Eco64I
                                >< Csp6I
      >< BslI
      >< BsiYI>< KpnI
      >< BscBI
      >< Bani
      >< Asp718
      >< NlaIII      >< AfaI
      >< BstXI      >< AccB1I      >< MaeIII
      >< Fnu4HI >< BbvI      >< Acc65I      BsgI ><
ACAGCAGCCA AACTCATGGT TGTGTCCCT GATTATGGTA CCTACAAGAA CACTTGATGAT -GGTAACACCT
12400      12410      12420      12430      12440      12450      12460

      >< Zsp2I
      >< Ppu10I

```

FIGURE 13.28

```

    >< NsiI
    >< Mph1103I
    >< NdeI>< EcoT22I
    >< AvaIII >< SfaNI
    >< SfaNI
    >< AciI
    DdeI ><
    BfrI ><
    TTACATATGC ATCTGCACTC TGGGAAATCC AGCAAGTTGT TGATGCGGAT AGCAAGATTG TTCAACTTAG
    12470      12480      12490      12500      12510      12520      12530

    >< Pali
    >< HaeIII >< MnlI >< OdeI>< OdeI ><
    >< BsuRI >< MaeIII >< BspWI
    >< Tru9I>< NlaIII
    >< MseI>< HphI
    >< XcmI>< BshI
    >< AluI BspWI ><
    TGAAATTAAC ATGGACAATT CACCAAATTT GGCTTGGCCT CTTATTGTTA CAGCTCTAAG AGCCAACTCA
    12540      12550      12560      12570      12580      12590      12600

    RsaI ><
    NlaIV ><
    KpnI ><
    >< Fnu4HI
    Eco64I ><
    Csp6I ><
    BscBI ><
    Asp718 ><
    AfaI ><
    >< AciI>< BanI
    AccB1I ><
    >< AluI >< SfcI
    >< DdeI>< BsrI
    >< PshAI Acc65I ><
    GCTGTAAAC TACAGAATAA TGAAGTGTAGT CCAGTAGCAC TACGACAGAT GTCCTGTGCG GCTGGTACCA
    12610      12620      12630      12640      12650      12660      12670

    >< TthHB8I
    >< TaqI
    >< SfuI
    >< NspV
    >< MnlI
    >< LspI
    >< Csp45I
    >< BstBI
    >< Bsp119I
    >< BsiCI
    >< Bpu14I
    >< AsuII
    >< RsaI
    >< Csp6I
    >< AluI
    >< AfaI
    CACAAACAGC TTGTACTGAT GACAATGCAC TTGCCTACTA TAACAATTTCG AAGGGAGGTA GGTGGTGTCT
    12680      12690      12700      12710      12720      12730      12740

    >< XhoII
    >< Sau3AI
    >< NdeII
    >< MflI
    >< MboI
    >< DpnII
    >< DpnI
    >< BstYI
    >< BspAI
    >< Bsp143I
    >< BglII
    >< TfiI
    >< RmaI
    >< HinfI
    >< MaeI
    >< DdeI
    >< RsaI
    >< Csp6I
    >< Csp6I>< RsaI
    >< AfaI>< AfaI
    GGCATTACTA TCAGACCACC AAGATCTCAA ATGGGCTAGA TTCCCTAAGA GTGATGGTAC AGGTACAATT
    12750      12760      12770      12780      12790      12800      12810

    >< Sau96I
    >< PssI
    >< Pali
    >< NspIV

```

FIGURE 13.29

```

                                >< HaeIII
                                >< EcoO109I
                                >< DraII
                                >< Cfr13I
                                >< BsuRI
                                >< BsiZI      RsaI >
                                >< BshI      Csp6I ><
                                >< AsuI      AfaI >
TACACAGAAC TGGAAACCACC TTGTAGGTTT GTTACAGACA CACCAAAAAGG GCCTAAAGTG AAATACTTGT
12820      12830      12840      12850      12860      12870      12880

                                >< SfcI
                                > < MboII
                                MaeII ><
                                >< Fnu4HI >< RsaI
                                >< Eco57I >< Csp6I
                                > < BbsI
                                >< Tru9I
                                >< MseI >< MnlI      >< BbvI      >< AluI      >< AfaI
ACTTCATCAA AGGCTTAAAC AACCTAAATA GAGGTATGGT GCTGGGCAGT TTAGCTGCTA CAGTACGTCT
12890      12900      12910      12920      12930      12940      12950

                                >< RsaI
                                >< SfcI >< Csp6I
                                >< BspWI      >< AfaI      >< BspMI      AccI ><
TCAGGCTGGA AATGCTACAG AAGTACCTGC CAATTCAACT GTGCTTTCCT TCTGTGCTTT TGCAGTAGAC
12960      12970      12980      12990      13000      13010      13020

                                >< RmaI
                                >< MnlI
                                >< MaeI      >< HphI
CCTGCTAAAG CATATAAGGA TTACCTAGCA AGTGGAGGAC AACCAATCAC CAACTGTGTG AAGATGTTGT
13030      13040      13050      13060      13070      13080      13090

                                >< SinI
                                >< Sau96I
                                >< NspIV
                                >< NspHII
                                >< NlaIII
                                >< Eco47I
                                >< Eam1105I
                                >< Cfr13I
                                >< BsiZI
                                >< Bme18I >< XcmI
                                >< AvaII   PleI ><
                                >< AfaI      >< AfaI      >< MaeIII      >< AluI      >< AsuI > < HinfI
GTACACACAC TGGTACAGGA CAGGCAATTA CTGTAACACC AGAAGCTAAC ATGGACCAAG AGTCCTTTGG
13100      13110      13120      13130      13140      13150      13160

                                >< TfiI
                                >< SfaNI
                                >< NlaIII      >< FokI      >< HinfI
TGGTGCTTCA TGTGTCTGT ATTGTAGATG CCACATTGAC CATCCAAATC CTAAAGGATT CTGTGACTTG
13170      13180      13190      13200      13210      13220      13230

                                > < RsaI
                                >< MaeII
                                >< Csp6I
                                > < AfaI
                                >< BsrI      >< DdeI
                                >< BfrI
AAAGGTAAGT ACGTCCAAAT ACCTACCACT TGTGCTAATG ACCCAGTGGG TTTTACACTT AGAAACACAG
13240      13250      13260      13270      13280      13290      13300

                                >< Thai

```

FIGURE 13.30


```

>< SfaNI
>< MvnI
>< BstUI
>< Bsp50I
>< RsaI
>< Csp6I
>< AfaI >< AciI
>< SfcI >< MaeIII
>< AccIISfaNI ><
TCTGTACCGT CTGCGGAATG TGGAAAGGTT ATGGCTGTAG TTGTGACCAA CTCCGCGAAC CCTTGATGCA
13310 13320 13330 13340 13350 13360 13370

>< Zsp2I
>< SfaNI
>< Mph1103I>< Tru9I
>< Ppu10I>< MaeII
>< NsiI> < FokI
>< EcoT22I >< MseI
Fnu4HI ><
BsgI ><
>< BbvI
>< AciI>> AvaIII >< DraI >< AciI >< Fnu4HI AciI ><
GTCTGCGGAT GCATCAACGT TTTTAAACGG GTTTGCGGTG TAAGTGCAGC CCGTCTTACA CCGTGGCGCA
13380 13390 13400 13410 13420 13430 13440

>< SpeI
>< ScaI
>< RsaI
>< RmaI
>< MaeI
>< Csp6I >< SfcI
>< BspWI >< AfaI >< AccI >< BcgI/a >< BspWI
CAGGCACTAG TACTGATGTC GTCTACAGGG CTTTGTATAT TTACAACGAA AAAGTTGCTG GTTTTGCAAA
13450 13460 13470 13480 13490 13500 13510

>< ScrFI
>< MvaI
>< MnlI
>< EcoRII
>< Ecl136I
>< BstOI
>< BstNI
>< BslI
>< DsaV >< BsiYI
>< BsiLI
>< ApyI >< PfuI >< HinfI
GTCCTAAAA ACTAATTGCT GTCGCTTCCA GGAGAAGGAT GAGGAAGGCA ATTTATTAGA CTCTTACTTT
13520 13530 13540 13550 13560 13570 13580

>< NlaIII
>< Ksp632I
>< EarI
>< Eam1104I
>< BsmAI
>< Tru9I
>< MnlI >< Alw26I >< MboII >< MseI
GTAGTTAAGA GGCATACTAT GTCTAACTAC CAACATGAAG AGACTATTTA TAACTTGGTT AAAGATTGTC
13590 13600 13610 13620 13630 13640 13650

>< RsaI
>< NlaIV
>< NlaIII
>< KpnI
>< HphI
>< Eco64I
>< Csp6I
>< BscBI
>< BanI
>< Asp718

```

FIGURE 13.31

```

>< NspBII
>< Acil
CAGCGGTGC TGTCCATGAC TTTTCAAGT TTAGAGTAGA TGGTGACATG GTACCACATA TATCAGCTCA
13660 13670 13680 13690 13700 13710 13720

>< MaeIII >< AfaI
> < AccBII MaeII ><
> < Acc65I > < HgaI

>< MnlI
>< MaeII
GCGTCTAACT AAATACACAA TGGCTGATTT AGTCTATGCT CTACGTCATT TTGATGAGGG TAATTGTGAT
13730 13740 13750 13760 13770 13780 13790

>< Tru9I
>< MseI >< MaeIII >< MunI
ACATTAAGAG AAATACTCGT CACATACAAT TGCTGTGATG ATGATTATTT CAATAAGAAG GATTGGTATG
13800 13810 13820 13830 13840 13850 13860

>< ThaI
>< MvnI
>< MluI
>< BstUI
>< Bsp50I
>< RsaI
>< HphI
>< TfiI >< AflIII >< DdeI >< Csp6I Tru9I ><
>< HinfI >< AccII >< BfrI >< AfaI MseI ><
ACTTCGTAGA GAATCCTGAC ATCTTACGCG TATATGCTAA CTTAGGTGAG CGTGTACGCC AATCATTATT
13870 13880 13890 13900 13910 13920 13930

XhoII >
Sau3AI >
NdeII >
MflI >
MboI >
DpnII >
BstYI >
BspAI >
> < SfaNI
>< RsaI
>< Csp6I
>< AfaI >< SfaNI
AAAGACTGTA CAATTCTGCG ATGCTATGCG TGATGCAGGC ATTGTAGGCG TACTGACATT AGATAATCAG
13940 13950 13960 13970 13980 13990 14000

> < ScrFI
> < MvaI
>< Fnu4HI
>< EcoRII
> < Ecl136I
> < BstOI
> < BstNI
>< Tru9I
>< MseI >< RsaI
>< DpnI >< Csp6I
>< Bsp143I >< BsrI
>< AlwI >< AfaI
GATCTTAATG GGAAGTGGTA CGATTTCGGT GATTTCGTAC AAGTAGCACC AGGCTGCGGA GTTCCTATTG
14010 14020 14030 14040 14050 14060 14070

>< RmaI
> < HinfI
>< Fnu4HPIeI ><
>< TfiI >< SfaNI
>< HinfI >< FokI
>< MamI >< BsiBI
>< BsaBI
>< MnlI >< MaeI
>< BbvI >< BspWI NdeI ><
TGGATTCATA TTACTCATTG CTGATGCCCA TCCTCACTTT GACTAGGGCA TTGGCTGCTG AGTCCCATAT
14080 14090 14100 14110 14120 14130 14140

>< Sau3AI
>< NdeII

```

FIGURE 13.32

```

>< MboI
>< MamI
>< DpnII
>< DpnI
>< BspWI
>< BspAI
>< BspI43I
>< BsiBI
>< BsaBI >< FokI
GGATGCTGAT CTCGCAAAAC CACTTATTAA GTGGGATTG CTGAAATATG ATTTTACGGA AGAGAGACTT
14150      14160      14170      14180      14190      14200      14210

>< SinI
>< Sau96I
>< NspIV
>< NspHII
>< NlaIV
>< TthHB8I
>< TaqI
>< McrI
>< Ksp632I
>< EarI
>< Eam1104I
>< BsmAI
>< MboII
>< Alw26I
TGTCTCTCG ACCGTTATTT TAAATATTGG GACCAGACAT ACCATCCCAA TTGTATTAACT TGTTCGGATG
14220      14230      14240      14250      14260      14270      14280

>< SinI ><
>< Sau96I ><
>< NspIV ><
>< NspHII >
>< Eco47I ><
>< Cfr13I ><
>< BsiZI ><
>< Bme18I ><
>< AvaII ><
>< AsuI ><
>< FokI
>< Tru9I
>< MseI
ATAGGTGTAT CCTTCATTGT GCAAACTTTA ATGTGTTATT TTCTACTGTG TTTCCACCTA CAAGTTTGG
14290      14300      14310      14320      14330      14340      14350

>< SpeI
>< RmaI
>< MaeI
>< SspI
>< BsrI
ACCACTAGTA AGAAAAATAT TTGTAGATGG TGTTCCTTTT GTTGTTCCTA CTGGATACCA TTTTCGTGAG
14360      14370      14380      14390      14400      14410      14420

>< ThaI>< Esp3I
>< DdeI
>< BstUI
>< Bsp50I >< BsmBI
>< MvnI>< BsmAI
>< HgaI>< AluI >< Alw26I
>< AfaI
>< FokI >< AccII >< BbvI
TTAGGAGTCG TACATAATCA GGATGTAAAC TTACATAGCT CGCGTCTCAG TTTCAAGGAA CTTTGTAGTGT
14430      14440      14450      14460      14470      14480      14490

>< Zsp2I
>< SphI
>< Ppu10I
>< PaeI
>< NspI

```

FIGURE 13.33

```

>< Sau3AI      >< NspHI
>< NdeII       >< NsiI
>< MboI        >< NlaIII
>< DpnII       >< Mph1103I
>< DpnI        >< Fnu4HI
>< Fnu4HI>< BspWI >< EcoT22I      NspHI ><
>< BspAI       >< BspWI      NlaIII ><
>< Bsp143I> < AvaIII > < AlwNI    >< RmaI    >< BspWI
>< AlwI        >< AluI       >< AluI    >< BbvI    >< MaeI    >< BbvI
ATGCTGCTGA TCCAGCTATG CATGCAGCTT CTGGCAATTT ATTGCTAGAT AAACGCACTA CATGCTTTTC
14500      14510      14520      14530      14540      14550      14560

>< ScrFI
>< NciI
>< MspI
>< HpaII
>< HapII
>< Fnu4HI
>< AlwNI
>< AluI
AGTAGCTGCA CTAACAAACA ATGTTGCTTT TCAAAGTGTG AAACCCGGTA ATTTTAATAA AGACTTTTAT
14570      14580      14590      14600      14610      14620      14630

>< Tru9I
>< MseI
>< MboII
DdeI ><
BbvI ><
GACTTTGCTG TGTCTAAAGG TTTCTTTAAG GAAGGAAGTT CTGTTGAACT AAAACACTTC TTCTTTGCTC
14640      14650      14660      14670      14680      14690      14700

>< FokI
>< Fnu4HI
EcoRV ><
Eco32I ><
AGGATGGCAA CGCTGCTATC AGTGATTATG ACTATTATCG TTATAATCTG CCAACAATGT GTGATATCAG
14710      14720      14730      14740      14750      14760      14770

>< VspI
>< Tru9I
>< MseI
>< AsnI
>< AseI
>< MaeIII
AACACTCCTA TTCGTAGTTG AAGTTGTTGA TAAATACTTT GATTGTTACG ATGGTGGCTG TATTAATGCC
14780      14790      14800      14810      14820      14830      14840

>< Tru9I
>< MseI
>< PvuII
>< HpaI
>< Psp5I
>< XcmI
>< HindII
>< NspBII
>< Tru9I
RmaI ><
>< HincII
>< AluI
>< MseI
MaeI ><
AACCAAGTAA TCGTTAACAA TCTGGATAAA TCAGCTGGTT TCCCATTTAA TAAATGGGGT AAGGCTAGAC
14850      14860      14870      14880      14890      14900      14910

>< SfaNI
>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< DpnI
>< Bsp143I
>< PfuI
>< HinfI>< MnlI
>< BspAI >< AlwI
>< AccI
TTTATTATGA CTCAATGAGT TATGAGGATC AAGATGCACT TTTCGCGTAT ACTAAGCGTA ATGTCATCCC
14920      14930      14940      14950      14960      14970      14980

>< SstI
>< SduI
>< SacI

```

FIGURE 13.34

```

>> NspII
>> HgiAI
>> Eco24I
>< Tru9I
>< TfiI
>< MseI
>< HinfI
>< Esp4I
>< AflII
>< BspWI
>< AluI
>< AluI
TACTATAACT CAAATGAATC TTAAGTATGC CATTAGTGCA AAGAATAGAG CTCGCACCGT AGCTGGTGTC
14990 15000 15010 15020 15030 15040 15050

>> ScaI
>< SfcI>< RsaI
>< BsmAI >< Csp6I
>< Alw26I >< AfaI
TCTATCTGTA GTACTATGAC AAATAGACAG TTTCATCAGA AATTATTGAA GTCAATAGCC GCCACTAGAG
15060 15070 15080 15090 15100 15110 15120

>> AluI
GAGCTACTGT GGTAATTGGA ACAAGCAAGT TTTACGGTGG CTGGCATAAT ATGTTAAAAA CTGTTTACAG
15130 15140 15150 15160 15170 15180 15190

>> Tru9I
>< MseI
>< NspI ><
>< NspHI ><
>< NlaIII ><
>< NlaIII
>< DdeI ><
>< BspWI ><
>< MaeIII
>< BfrI ><
TGATGTAGAA ACTCCACACC TTATGGGTTG GGATTATCCA AAATGTGACA GAGCCATGCC TAACATGCTT
15200 15210 15220 15230 15240 15250 15260

>< Pali
>< HaeIII
>< BsuRI
>< BshI
>< MnlI
>< MaeIII
>< SfcI ><
AGGATAATGG CCTCTCTTGT TCTTGCTCGC AAACATAACA CTTGCTGTAA CTTATCACAC CGTTTCTACA
15270 15280 15290 15300 15310 15320 15330

>> Tru9I ><
>< ScrFI >
>< MvaI >
>< MseI
>< MstI
>< HinfII
>< HinfI
>< HhaI
>< FspI
>< FdiII
>< NlaIII
>< NlaIII
>< Fnu4HI
>< AciI
>< ApyI >
>< AluI
>< AviII >< MseI
>< BspWI
>< Tru9I
>< MaeIII ><
>< MseI
>< AluI ><
GGTTAGCTAA CGAGTGTGCG CAAGTATTAA GTGAGATGGT CATGTGTGGC GGCTCACTAT ATGTTAAACC
15340 15350 15360 15370 15380 15390 15400

>< SfaNI
>< MspI
>< HpaII
>< HapII
>< HphI
>< BspWI
>< Tru9I
>< MaeIII ><
>< MseI
>< AluI ><

```

FIGURE 13.35

```

AGGTGGAACA TCATCCGGTG ATGCTACAAC TGCTTATGCT AATAGTGTCT TTAACATTTG TCAAGCTGTT
15410      15420      15430      15440      15450      15460      15470

>< BspWI                                     >< AluI                                     >< DrdI
ACAGCCAATG TAAATGCACT TCTTTCAACT GATGGTAATA AGATAGCTGA CAAGTATGTC CGCAATCTAC
15480      15490      15500      15510      15520      15530      15540

                                     >< Sau3AI
                                     >< NdeII
                                     >< MboI
> < MamI
   >< FbaI
   >< DpnII
   >< DpnI
   >< BspHI
   >< BspAI
   >< Bsp143I
   >< BsiQI
                                     >< SfcI                                     > < BsiBI>< NlaIII
                                     >< BsmAI                                     > < BsaBI>< FokI
                                     >< Alw26I                                     >< BclI>< EcoRI                                     FokI ><
AACACAGGCT CTATGAGTGT CTCTATAGAA ATAGGGATGT TGATCATGAA TTCGTGGATG AGTTTTACGC
15550      15560      15570      15580      15590      15600      15610

                                     >< TfiI
                                     >< SfaNI
                                     >< NlaIII
>< BspMI                                     >< HinfI                                     >< MaeIII
TTACCTGCGT AAACATTCTT CCATGATGAT TCTTTCTGAT GATGCCGTTG TGTGCTATAA CAGTAACTAT
15620      15630      15640      15650      15660      15670      15680

                                     > < RmaI
                                     >< NheI >< Tru9I
>< Fnu4HI                                     > < MaeI                                     >< Tru9I
>< AciI                                     >< AluI >< MseI                                     >< MseI                                     MnlI ><
GCGGCTCAAG GTTTAGTAGC TAGCATTAAAG AACTTTAAGG CAGTCTTTA TTATCAAAAT AATGTGTTC
15690      15700      15710      15720      15730      15740      15750

                                     >< SniI
                                     >< Sau96I
                                     >< PssI
                                     >< Psp5II
                                     >< PpuMI
                                     >< NspIV
                                     >< NspHII
                                     >< EcoO109I
                                     >< Eco47I
                                     >< DraII
                                     >< Cfr13I
                                     >< BsiZI
                                     >< Bme18I
                                     >< DdeI
>< NlaIII                                     >< BsmAI
>< DdeI                                     >< Alw26I
TGCTGAGGC AAAATGTTGG ACTGAGACTG ACCTTACTAA AGGACCTCAC GAATTTTGCT CACAGCATAC
15760      15770      15780      15790      15800      15810      15820

                                     >< XhoII
                                     >< Sau3AI
                                     >< NdeII
                                     >< MflI
                                     >< MboI

```

FIGURE 13. 36

```

                >< RsaI          >< DpnII
                >< MaeII        >< DpnI
                >< Csp6I        >< BstYI    > < SspI
                >< RmaI          >< BsaAI      >< BspMI    HinPII ><
                >< MaeI          >< AflIII     >< BspAI    Hin6I ><
                >< BspWI>< MseI      >< AfaI      >< AlwI>< Bsp143I  HhaI ><
AATGCTAGTT AAACAAGGAG ATGATTACGT GTACCTGCCT TACCCAGATC CATCAAGAAT ATTAGGCGCA
15830      15840      15850      15860      15870      15880      15890

                >< RsaI          >< SfaNI
                >< TthHB8I        >< Csp6I        >< MaeIII
                >< TaqI          >< AfaI          BsrI ><
GGCTGTTTTG TCGATGATAT TGTCAAAACA GATGGTACAC TTATGATTGA AAGGTTCTGT TCACTGGCTA
15900      15910      15920      15930      15940      15950      15960

> < FokI
>< BspWI
TTGATGCTTA CCCACTTACA AAACATCCTA ATCAGGAGTA TGCTGATGTC TTCACTTGT ATTTACAATA
15970      15980      15990      16000      16010      16020      16030

                >< Van91I
                >< PflMI
                >< NspI
                > < Pali>< NspHI
                > < MscI>< NlaIII
                > < HaeIII
                > < BsuRI
                >< BsrI
                >< EaeI >< BslI >< NspI
                > < BshI>< BsiYI >< NspHI
                >< NlaIII      >< AflIII >< AflIII
                >< MaeIII      >< AluI > < BalI>< AccB7I >< NlaIII
CATTAGAAAG TTACATGATG AGCTTACTGG CCACATGTTG GACATGTATT CCGTAATGCT AACTAATGAT
16040      16050      16060      16070      16080      16090      16100

                >< RsaI> < NlaIV
                >< MnlI
                >< Csp6I      >< DdeI          >< RsaI
                >> BsrI >< MnlI      >< Csp6I
                >< AfaI> < BscBI      >< AfaI          SfcI ><
AACACCTCAC GGTACTGGGA ACCTGAGTTT TATGAGGCTA TGTACACACC ACATACAGTC TTGCAGGCTG
16110      16120      16130      16140      16150      16160      16170

                >< NlaIV
                >< EcoNI
                >< Eco31I
                >< Eco64I>< BsmAI
                >< BscBI >< BslI
                >< BanI >< BsiYI
                >< AclI >< BsaI
                >< BspWI      >< AccB1I>< Alw26I BbvI ><
TAGGTGCTTG TGTATTGTGC AATTCACAGA CTTCACTTCG TTGCGGTGCC TGTATTAGGA GACCATTTCCT
16180      16190      16200      16210      16220      16230      16240

                >< Tth111I
                >< Fnu4HI      >< NlaIII
                >< BspWI >< AspI          > < Tru9I
                >> BspWI >< MseI          > < MseI
ATGTTGCAAG TGCTGCTATG ACCATGTCAT TTCAACATCA CACAAATTAG TGTGTCTGT TAATCCCTAT
16250      16260      16270      16280      16290      16300      16310

                >< ScrFI
                >< MvaI

```

FIGURE 13.37

```

>< EcoRII
>< Ecl136I
>< DsaV
>< BstOI
>< BstNI
>< BsiI
>< BsaJI
>< ApyI
>< MaeIII >< MaeIII
>< MaeI
>< AluI
GTTTGCAATG CCCAGGTTG TGATGTCAC TATGTGACAC AACTGTATCT AGGAGGTATG AGCTATTATT
16320 16330 16340 16350 16360 16370 16380

>< MaeIII >< MnlI
GCAAGTCACA TAAGCCTCCC ATTAGTTTTTC CATTATGTGC TAATGGTCAG GTTTTGGTT TATACAAAAA
16390 16400 16410 16420 16430 16440 16450

>< NspI
>< NspHI >< Tth111I
>< NlaIII>< MaeIII>< MaeIII
>< AflIII >< AfpI
CACATGTGTA GGCAGTGACA ATGTCAC TCAATGCG ATAGCAACAT GTGATTGGAC TAATGCTGGC
16460 16470 16480 16490 16500 16510 16520

>< RsaI
>< P1eI
>< DdeI
>< Csp6I
>< BsmAI >< HinfI
>< Alw26I >< HindIII
>< AfaI >< AluI >< Fnu4HI >< BbvI
GATTACATAC TTGCCAACAC TTGTACTGAG AGACTCAAGC TTTTCGCAGC AGAAACGCTC AAAGCCACTG
16530 16540 16550 16560 16570 16580 16590

>< ThaI
>< ScaI
>< RsaI >< RsaI
>< MvnI
>< Csp6I >< Csp6I
>< BstUI
>< Bsp50I
>< Tru9I
>< MseI >< NdeI
>< AluI
AGGAAACATT TAAGCTGTCA TATGGTATTG CCACTGTACG CGAAGTACTC TCTGACAGAG AATTGCATCT
16600 16610 16620 16630 16640 16650 16660

MaeIII ><
>< MaeIII
>< EcoO65I
>< Eco91I
>< BstPI
>< BstEII
>< BsrI
>< SfaNI
>< NlaIII
>< RmaI
>< MaeI
TTCATGGGAG GTTGGAAAAC CTAGACCACC ATTGAACAGA AACTATGTCT TTAAGGGTTA CCGTGTAACT
16670 16680 16690 16700 16710 16720 16730

RsaI ><
>< MnlI
>< HphI
>< RsaI
>< Csp6I
>< Csp6I
>< SfaNI
>< MaeIII
>< HphI AfaI ><
AAAAATAGTA AAGTACAGAT TGGAGAGTAC ACCTTTGAAA AAGGTGACTA TGGTGATGCT GTTGTGTACA
16740 16750 16760 16770 16780 16790 16800

```

FIGURE 13.38


```

>< RsaI
>< Csp6I
>< AfaI
GAGGTACTAC GACATACAAG TTGAATGTTG GTGATTACTT TGTGTTGACA TCTCAGCTG TAATGCCACT
16810      16820      16830      16840      16850      16860      16870

>< HphI
>< HindII
>< HincII
DdeI ><
BfrI ><

>< VneI
>< SnuI
>< SduI
>< NspII
>< HgiAI
>< DraIII
>< Bsp1286I
>< BmyI
>< ApaLI
>< Alw44I
>< Alw21I
TAGTGACCT ACTCTAGTGC CACAAGAGCA CTATGTGAGA ATTACTGGCT TGTACCCAAC ACTCAACATC
16880      16890      16900      16910      16920      16930      16940

>< SphI
>< PaeI
>< NlaIII
>< NspI
>< NspHI
>< BspWI
>< DraIII
>< RsaI
>< Csp6I
>< BsrI
>< AfaI
DdeI ><
StyI ><
SinI >
Sau96I >
NspIV >
EcoT14I ><
Eco47I >
Eco130I ><
>< ScaI Cfr13I >
BssT1I ><
>< SphI >< RsaI Bsi2I >
>< PaeI BsaJI ><
>< NlaIII Bme18I >
>< NspI>< Csp6I AvaII >
>< NspHI>< AfaI AsuI >
TCAGATGAGT TTTCTAGCAA TGTGCAAAT TATCAAAGG TCGGCATGCA AAAGTACTCT AACTCCAAG
16950      16960      16970      16980      16990      17000      17010

>< ScrFI
>< RsaI
>< MvaI
>< EcoRII
>< Ecl136I
>< Csp6I
>< BstOI
>< BstNI
>< XcmI >< BslI
>< NspHII >< BsiYI
>< BsilI
>< ApyI >< BsrI
>< DsaV>< AfaI >< HinfI>< PleI
GACCACCTGG TACTGGTAAG AGTCATTTTG CCATCGGACT TGCTCTCTAT TACCCATCTG CTCGCATAGT
17020      17030      17040      17050      17060      17070      17080

>< SfaNI
>< SphI >< PvuII
>< PaeI >< Psp5I
>< NspI >< NspBII
>< NspHI >< Fnu4HI
>< Bst1107I >< NlaIII>< BspWI >< Tru9I
>< AccI >< NlaIII >< AluI >< BbvI >< SspI
>< MseI
GTATACGGCA TGCTCTCATG CAGCTGTTGA TGCCCTATGT GAAAAGGCAT TAAATATTT GCCCATAGAT
17090      17100      17110      17120      17130      17140      17150

```

FIGURE 13.39

```

> < ThaI
>< ThaI
> < MvnI
>< MvnI >< ThaI
> < HinP1I
>< HinP1I
>< HinP1I >< MvnI
> < Hin6I
>< Hin6I
> < HhaI
>< HhaI >< HhaI
> < CfoI
>< CfoI >< CfoI
> < BstUI
>< BstUI >< BstUI
>< BssHII
>< BspMI
> < Bsp50I
>< Bsp50I>< Bsp50I
>< TfiI >< Hin6I> < AccII RmaI >
>< HinfI >< AccII >< AccII MaeI >
> < EcoRI
AAATGTAGTA GAATCATACC TGC GCGT GCG CGCGTAGAGT GTTTGTGATAA ATTCAAAGTG AATTCAACAC
17160 17170 17180 17190 17200 17210 17220

>< Zsp2I
>< Ppu10I
>< NsiI
>< Mph1103I
>< EcoT22I
>< BsgI > < AvaIII >< DrdI
TAGAACAGTA TGT TTTCTGC ACTGTAAATG CATTGCCAGA AACAACTGCT GACATTGTAG TCTTTGATGA
17230 17240 17250 17260 17270 17280 17290

>< RmaI
>< MaeI >< MaeII
AATCTCTATG GCTACTAATT ATGACTTGAG TGTTGTCAAT GCTAGACTTC GTGCAAAACA CTACGTCTAT
17300 17310 17320 17330 17340 17350 17360

>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< DpnI
>< BspAI >< RmaI
>< AlwI>< Bsp143I > < AciI >< MaeI SspI ><
ATTGGCGATC CTGCTCAATT ACCAGCCCC CGCACATTGC TGAATAAGG CACACTAGAA CCAGATATT
17370 17380 17390 17400 17410 17420 17430

>< SniI
>< Sau96I
>< NspIV >< StyI
>< NspHII >< NspI
>< Eco47I >< NspHI
>< Cfr13I >< NlaIII
>< Bsi2I >< EcoT14I
>< BsgI >< Eco130I
>< Bme18I >< BssT1I
>< AvaII >< BsaJI
>< AsuI> < AflIII
>< Tru9I
>< MseI
TTAATTCAGT GTGCAGACTT ATGAAAACAA TAGGTCCAGA CATGTTCCCTT GGAACCTGTC GCCGTTGTCC
17440 17450 17460 17470 17480 17490 17500

```

FIGURE 13. 40

```

>< HindII
>< HincII
>< AluI
TGCTGAAATT GTTGACACTG TGAGTGCTTT AGTTTATGAC AATAAGCTAA AAGCACACAA GGATAAGTTA
17510 17520 17530 17540 17550 17560 17570

>< AluI
GCTCAATGCT TCAAAATGTT CTACAAAGGT GTTATTACAC ATGATGTTTC ATCTGCAATC AACAGACCTC
17580 17590 17600 17610 17620 17630 17640

>< MnlI
>< EcoNI
>< BslI
>< BsiYI
>< HphI
>< AluI
AAATAGGCGT TGTAAGAGAA TTTCTTACAC GCAATCCTGC TTGGAGAAAA GCTGTTTTTA TCTCACTTA
17650 17660 17670 17680 17690 17700 17710

>< SfcI
>< DdeI
>< TfiI
>< AluI
>< BfrI
>< HinfI
TAATTCACAG AACGCTGTAG CTTCAAAAAT CTTAGGATTG CCTACGCAGA CTGTTGATTC ATCAGAGGGT
17720 17730 17740 17750 17760 17770 17780

>< Tth111I
>< AspI
>< HindII
>< HincII
>< AclI
TCTGAATATG ACTATGTCAT ATTCACACAA ACTACTGAAA CAGCACACTC TTGTAATGTC AACCGTTTCA
17790 17800 17810 17820 17830 17840 17850

>< XhoII
>< Sau3AI
>< NdeII
>< MflI
>< MboI
>< MamI
>< DpnII
>< DpnI
>< BstYI
>< BspAI
>< Bspl43I
>< BsiBI
>< BsaBI
>< BglII
>< BspWI
ATGTGGCTAT CACAAGGGCA AAAATTGGCA TTTTGTGCAT AATGTCTGAT AGAGATCTTT ATGACAACT
17860 17870 17880 17890 17900 17910 17920

>< XbaI
>< RmaI
>< MaeI
>< MaeII
>< MaeIII
>< BsrI
><
GCAATTTACA AGTCTAGAAA TACCACGTCG CAATGTGGCT ACATTACAAG CAGAAAATGT AACTGGACTT
17930 17940 17950 17960 17970 17980 17990

>< Sau3AI
>< NdeII
>< MboII
>< MboI
>< FokI
>< DpnII
>< DpnI
>< BspAI
>< Bspl43I
>< BanI
>< BscBI
>< Tru9I
>< MseI>< SfcI
>< BbsI>< BsrI
>< NlaIV
>< Eco64I
>< AccBI
>< MnlI><
>< DdeI

```

FIGURE 13. 41

```

TTTAAGGACT GTAGTAAGAT CATTACTGGT CTTTCATCCTA CACAGGCAC TACACACCTC AGCGTTGATA
18000      18010      18020      18030      18040      18050      18060

                                >< ScrFI
                                >< MvaI
                                >< EcoRII
                                >< Eco57I
                                >< Ecl136I
                                >< DsaV
                                >< BstOI
                                >< BstNI
                                >< PleI
                                >< NlaIII
                                >< HinfI ><
                                >< AccI ><
                                >< HindII>< BsiII
                                >< HincII>< ApyI
TAAAGTTCAA GACTGAAGGA TTATGTGTG ACATACCAGG CATACCAAAG GACATGACCT ACCGTAGACT
18070      18080      18090      18100      18110      18120      18130

                                >< MaeIII
                                >< Eco65I
                                >< Eco91I
                                >< BstXI
                                >< BstPI
                                >< BstEII
                                >< HphI
                                >< AccII ><
                                >< ThaI ><
                                >< MvnI ><
                                >< BstUI ><
                                >< Bsp50I ><
                                >< AciI
CATCTCTATG ATGGGTTTCA AAATGAATTA CCAAGTCAAT GGTTACCCTA ATATGTTTAT CACCCGCGAA
18140      18150      18160      18170      18180      18190      18200

                                >< XmnI
                                >< MboII
                                >< MaeIII
                                >< Asp700I
                                >< SfaNI
                                >< RmaI
                                >< NlaIII
                                >< MaeI
>< AluI >< MaeII >< MnlI
GAAGCTATTC GTCACGTTTC TGCCTGGATT GGCTTTGATG TAGAGGGCTG TCATGCAACT AGAGATGCTG
18210      18220      18230      18240      18250      18260      18270

                                >< Tru9I
                                >< MseI
                                >< HpaI
                                >< HindII
                                >< RsaI
                                >< HincII
                                >< Csp6I
                                >< DdeI >< AluI
                                >< BsrI ><
                                >< AfaI
                                >< BfrI
                                >< AfaI
TGGGTACTAA CCTACCTCTC CAGCTAGGAT TTTCTACAGG TGTTAACTTA GTAGCTGTAC CGACTGGTTA
18280      18290      18300      18310      18320      18330      18340

                                >< ScrFI
                                >< MvaI
                                >< MnlI
                                >< MaeIII
                                >< EcoRII
                                >< Eco65I
                                >< EcoNI
                                >< Eco91I
                                >< Ecl136I
                                >< DsaV
                                >< Tru9I ><
                                >< DraIII
                                >< BstPI
                                >< BstOI
                                >< BstNI
                                >< PmeI ><
                                >< BstEII
                                >< BsiI
                                >< MseI ><
                                >< BsiII
                                >< HphI ><
                                >< BsiI
                                >< DraI ><
                                >< ApyI >< BsrI
>< HindII >< HphI >< Tru9I
>< HincII >< EcoRI >< MseI

```

FIGURE 13.42

```

TGTGACACT GAAAATAACA CAGAATTCAC CAGAGTTAAT GCAAAACCTC CACCAGGTGA CCAGTTTAAA
18350      18360      18370      18380      18390      18400      18410

                >< ScrFI
                >< MvaI
                >< EcoRII
                >< Ecl136I
                >< DsaV
                >< BstOI
                >< BstNI
                >< BsiLI
                >< BsaJI
                >< NlaIII
                >< ApyI
                >< RsaI
                >< DdeI ><
                >< Tru9I>< Csp6I
                >< MseI >< AfaI
CATCTTATAC CACTCATGTA TAAAGGCTTG CCCTGGAATG TAGTGCGTAT TAAGATAGTA CAAATGCTCA
18420      18430      18440      18450      18460      18470      18480

                >< NlaIII
                >< HinPII
                >< Tth111I
                >< Hin6I
                >< HhaI
                >< AspI
                >< PleI
                >< CfoI
                >< AluI
GTGATACACT GAAAGGATTG TCAGACAGAG TCGTGTTCGT CCTTTGGGCG CATGGCTTTG AGCTTACATC
18490      18500      18510      18520      18530      18540      18550

                >< SinI
                >< Sau96I
                >< NspIV
                >< NspHII
                >< Eco47I
                >< Cfr13I
                >< ScaI
                >< BsiZI
                >< RsaI
                >< Bme18I
                >< Csp6I
                >< AvaII
                >< MaeII
                >< AfaI
                >< AsuI
                >< AflIII
                >< MaeIII>< MaeII
AATGAAGTAC TTTGTCAAGA TTGGACCTGA AAGAACGTGT TGTCTGTGTG ACAAACGTGC AACTTGCTTT
18560      18570      18580      18590      18600      18610      18620

                >< TfiI
                >< Tth111I
                >< HinfI
                >< AspI
TCTACTTCAT CAGATACTTA TGCCTGCTGG AATCATTCTG TGGGTTTGA CTATGTCTAT AACCCATTTA
18630      18640      18650      18660      18670      18680      18690

                >< ScrFI
                >< RsaI ><
                >< MvaI
                >< EcoRII
                >< Ecl136I ><
                >< DsaV
                >< Csp6I ><
                >< BstXI ><
                >< MaeIII
                >< EcoO65I
                >< Eco91I
                >< BstPI
                >< Eco57I>
                >< BstEII
                >< MaeIII >< NlaIII
                >< AfaI ><
TGATTGATGT TCAGCAGTGG GGCTTTACGG GTAACCTTCA GAGTAACCAT GACCAACATT GCCAGGTACA
18700      18710      18720      18730      18740      18750      18760

                >< SfaNI
                >< RmaI
                >< NspI
                >< NspHI

```

FIGURE 13.43

```

                >< NlaIII
                >< MaeI
    >< NlaIII    >< BspWI
    > < AflIII
TGGAAATGCA CATGTGGCTA GTTGTGATGC TATCATGACT AGATGTTTAG CAGTCCATGA GTGCTTTGTT
    18770      18780      18790      18800      18810      18820      18830

                >< RmaI
                >< NlaIII
                >< MaeI
                Tru9I ><
                >< NlaIII
                MseI ><
                >< BspHI
                >< EcoNI> < MnlI
                >< BslI
                >< BsiYI
                >< DdeI >< MseI
                >< Tru9I
                >< AccII
AAGCGCGTTG ATTGGTCTGT TGAATACCCCT ATTATAGGAG ATGAAGTCTAG GGTAAATCTCT GCTTGCAGAA
    18840      18850      18860      18870      18880      18890      18900

                >< RsaI
                >< Csp6I
                >< AfaI
                >< NlaIII
                >< BspWI
                >< BsrI
                >< BspHI
                >< MboII
                > < NlaIII
AAGTACAACA CATGGTTGTG AAGTCTGCAT TGCTTGCTGA TAAGTTTCCA GTTCTTCATG ACATTGGAAA
    18910      18920      18930      18940      18950      18960      18970

                >< SauI
                >< MstII
                >< Eco81I
                >< DdeI
                >< CvnI
                >< Bsu36I
                >< Bse21I
                >< AxyI
                >< AocI
                >< MnlI
                >< SfaNI
                >< NlaIII ><
                >< EspI
                >< Eco57I
                >< MaeIII ><
                >< DdeI
                >< CeliI
                >< Bpu1102I
TCCAAAGGCT ATCAAGTGTG TGCCTCAGGC TGAAGTAGAA TGAAGTTCT ACGATGCTCA GCCATGTAGT
    18980      18990      19000      19010      19020      19030      19040

                >< MnlI
                >< Ksp632I
                >< HindIII
                >< EarI
                >< AluI
                >< MboII
                >< Eam1104I
GACAAAGCTT ACAAATAGA GGAAGTCTTC TATTCTTATG CTACACATCA CGATAAATTC ACTGATGGTG
    19050      19060      19070      19080      19090      19100      19110

                >< Sau3AI
                >< NdeII
                >< MboI
                >< MaeII> < MaeIII
                >< DpnII
                >< DpnI
                >< BspAI
                >< MaeIII >< Bsp143I
                >< MunI
                >< HinfI >
                >< DrdI ><
TTGTTTGTGTT TTGGAATTGT AACGTGATC GTTACCCAGC CAATGCAATT GTGTGTAGGT TTGACACAAG
    19120      19130      19140      19150      19160      19170      19180

                >< ScrFI
                >< MvaI
                >< EcoRII
                Zsp2I ><
                >< SphI
                > < Ppu10I
                >< PaeI
                >< NspI
                >< NspHI
                >< NlaIII
                Mph1103I ><

```

FIGURE 1344

```

                >> Ecl136I
                >> DsaV
                >> BstOI
                >> BstNI
                >> BsiLI
                >> ApyI
    >> PleI
    AGTCTTGTCA AACTTGAAGT TACCAGGCTG TGATGGTGGT AGTTTGATG TGAATAAGCA TGCATTCCAC
    19190      19200      19210      19220      19230      19240      19250

                >> Tru9I
                > < MunI
                >> TthHB8I
                >> MseI
    >> BcgI/a >> TaqI
                >> DraI
                >> BcgI
    ACTCCAGCTT TCGATAAAAG TGCATTACT AATTAAAGC AATTGCCTTT CTTTACTAT TCTGATAGTC
    19260      19270      19280      19290      19300      19310      19320

                >> PleI
                >> NlaIII
                >> BsmAI
                >> HinfI>> Alw26I
    CTTGTGAGTC TCAATGCAAA CAAGTAGTGT CGGATATTGA TTATGTTCCA CTCAAATCTG CTACGTGTAT
    19330      19340      19350      19360      19370      19380      19390

                Zsp2I >
                >> ScaI
                Ppu10I ><
                >> RsaINsiI >
                Mph1103I >
                >> SfaNIEcoT22I >
                > < RsaI >> Csp6I
                >> Csp6I
                >> NlaIII> < AfaI >> AfaI
    TACACGATGC AATTTAGGTG GTGCTGTTTG CAGACACCAT GCAAATGAGT ACCGACAGTA CTTGGATGCA
    19400      19410      19420      19430      19440      19450      19460

    >> FokI
    TATAATATGA TGATTCTGC TGGATTAGC CTATGGATT ACAAACAATT TGATACTTAT AACCTGTGGA
    19470      19480      19490      19500      19510      19520      19530

                >> ScrFI
                >> MvaI
                >> MaeIII
    >> EcoRII
    >> Ecl136I
    >> DsaV
    >> BstOI
    >> BstNI
    >> BsiLI
    >> ApyI
    ATACATTAC CAGGTACAG AGTTTAGAAA ATGTGGCTTA TAATGTTGTT AATAAAGGAC ACTTTGATGG
    19540      19550      19560      19570      19580      19590      19600

                >> SgrAI
                >> NaeI
    >> MspI
    >> HpaII
    >> HapII
    >> Cfr10I
    >> BspWI
    ACACGCCGGC GAAGCACCTG TTTCCATCAT TAATAATGCT GTTACACAA AGGTAGATGG TATTGATGG
    19610      19620      19630      19640      19650      19660      19670

                > < VspI
                > < Tru9I
                > < MseI
                > < AsnI
                > < AseI

```

FIGURE 13. 45

```

>< XhoII
>< Sau3AI
>< NdeII
>< MflI
>< MboI
>< DpnII
  >< DpnI
>< BstYI
>< BspAI
  >< Bsp143I
>< BglII
GAGATCTTTG AAAATAAGAC AACACTTCCT GTTAATGTTG CATTGAGCT TTGGGCTAAG CGTAACATTA
19680      19690      19700      19710      19720      19730      19740

                                >< MaeIII
                                >< EspI
                                >< DdeITru9I ><
                                >< CeliIMseI ><
                                >< Bpu1102I
                                >< Tru9I
                                >< MseI
                                >< AluI
                                >< Fnu4HI
                                >< EcoRV
                                >< Eco32I
  >< BsrI      >< MseI      >< BbvI      >< EcoRV
AACCAGTGCC AGAGATTAAG ATACTCAATA ATTTGGGTGT TGATATCGCT GCTAATACTG TAATCTGGGA
19750      19760      19770      19780      19790      19800      19810

                                >< NspI
                                >< NspHI
                                >< NlaIII
                                >< BsqI
                                >< AflIII
CTACAAAAGA GAAGCCCCAG CACATGTATC TACAATAGGT GTCTGCACAA TGA CTGACAT TGCCAAGAAA
19820      19830      19840      19850      19860      19870      19880

  >< DdeI>< MboII
CCTACTGAGA GTGCTTGTTC TTCCTTACT GTCTTGTTC ATGGTAGAGT GGAAGGACAG GTAGACCTTT
19890      19900      19910      19920      19930      19940      19950

                                SinI ><
                                Sau96I ><
                                NspIV ><
                                NspHII ><
                                NlaIV ><
                                Eco47I ><
                                Cfr13I ><
                                >< BslI
                                BsiZI ><
                                >< BsiYI
                                BscBI ><
                                Bme18I ><
                                AvaII ><
                                AsuI ><
                                >< Tru9I
                                >< MseI
TTAGAAACGC CCGTAATGGT GTTTTAATAA CAGAAGGTTT AGTCAAAGGT CTAACACCTT CAAAGGGACC
19960      19970      19980      19990      20000      20010      20020

                                >< VspI
                                >< Tru9I
                                >< PleI
                                >< MseI
  >< RmaI
  >< NheI      >< MaeIII
  >< MaeI
  >< HgaI>< AluI      >< HinfI>< AseI      >< HinfI
AGCACAAGCT AGCGTCAATG GAGTCACATT AATTGGAGAA TCAGTAAAAA CACAGTTTAA CTACTTTAAG
20030      20040      20050      20060      20070      20080      20090

                                >< DdeI      >< MnlI      Tru9I ><
                                >< BsmAI      >< DdeI

```

FIGURE 1346


```

>< AccI
AAAGTAGACG GCATTATTCA ACAGTTGCCT GAAACCTACT TTACTCAGAG CAGAGACTTA GAGGATTTTA
20100 20110 20120 20130 20140 20150 20160

>< TthHB8I
>< TaqI
>< SstI
>< SduI
>< SacI
> < Paer7I
> < NspIII
>< NspII
>< HgiAI
> < Eco88I
> < XhoI> Eco24I
>< XcmI
>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< DpnI
>< BspAI
>< Bsp143I
AGCCCAGATC ACAAATGGAA ACTGACTTTC TCGAGCTCGC TATGGATGAA TTCATACAGC GATATAAGCT
20170 20180 20190 20200 20210 20220 20230

>< TthHB8I
>< TaqI
>< SfuI
>< NspV
>< LspI
>< Csp45I
>< BstBI
>< Bsp119I
>< BsiCI
>< Bpu14I
>< AsuII >< BcgI
CGAGGGCTAT GCCTTCGAAC ACATCGTTTA TGGAGATTTC AGTCATGGAC AACTTGGCGG TCTTCATTTA
20240 20250 20260 20270 20280 20290 20300

>< HphI
>< HinPII
>< Hin6I
>< EspI > < HhaI >< TfiI
>< DdeI >< HaeII
>< CelII >< Eco47III >< Tru9I
>< Bpu1102I > < CfoI >< HinfI >< MseI
>< BfrI >< Bsp143II >< MnlI
ATGATAGGCT TAGCCAAGCG CTCACAAGAT TCACCACTTA AATTAGAGGA TTTTATCCCT ATGGACAGCA
20310 20320 20330 20340 20350 20360 20370

>< MstI
>< HinPII
>< Hin6I
>< HhaI
>< FspI
>< FdiII
>< CfoI
>< SfaNI >< AviII
CAGTGAAAAA TTACTTCATA ACAGATGCGC AAACAGGTTC ATCAAAATGT GTGTGTTCTG TGATTGATCT
20380 20390 20400 20410 20420 20430 20440

>< TthHB8I

```

FIGURE 13.47

```

>> Tth111I
>> TaqI
>> AspI > < MaeIII MaeIII ><
TTTACTTGAT GACTTTGTCTG AGATAATAAA GTCACAAGAT TTGTCAGTGA TTTCAAAAGT GGTCAAGGTT
20450 20460 20470 20480 20490 20500 20510

>< NspI
>< NspHI
>< NlaIII
>< FokI

>< MunI > < NlaIII >< AflIII
ACAATTGACT ATGCTGAAAT TTCATTTCATG CTTTGGTGTA AGGATGGACA TGTGAAACC TTCTACCCAA
20520 20530 20540 20550 20560 20570 20580

>< SfaNI
>< ScrFI
>< MvaI
>< EcoRII
>< Ecl136I
>< DsaV
>< BstOI >< SfaNI
>< BstNI >< RsaI BspWI ><
>< BsiLI > < Csp6I BsmI >
>< BspWI >< ApyI >< AfaI BscCI ><
AACTACAAGC AAGTCAAGCG TGGCAACCAG GTGTTGCGAT GCCTAACTTG TACAAGATGC AAAGAATGCT
20590 20600 20610 20620 20630 20640 20650

>< Eco57I >< MaeIII >< HphI
TCTTGAAAAG TGTGACCTTC AGAATTATGG TGAAAATGCT GTTATACCAA AAGGAATAAT GATGAATGTC
20660 20670 20680 20690 20700 20710 20720

> < RsaI
>< Csp6I
>< Bst1107I >< Tru9I >< AluI
>< AccI >< MseI > < AfaINlaIII ><
GCAAAGTATA CTCAACTGTG TCAATACTTA AATACACTTA CTTTAGCTGT ACCCTACAAC ATGAGAGTTA
20730 20740 20750 20760 20770 20780 20790

>< ScrFI
>< RsaI
>< MvaI
>< EcoRII >< NspBII
>< Ecl136I >< SduI
> < Csp6I >< NspII
>< BstOI >< PvuII>< HqiAI
>< BstNI >< DdeI
>< BsiLI >< Psp5I>< Bsp1286I
>< ApyI >< AluI >< BmyI
>< DsaV>< AfaI >< Alw21I
TTCACCTTGG TGCTGGCTCT GATAAAGGAG TTGCACCAGG TACAGCTGTG CTCAGACAAT GGTGCGCAAC
20800 20810 20820 20830 20840 20850 20860

>< XhoII
>< Tru9I
>< Sau3AI
>< NdeII
>< TthHB8I >< MseI
>< MflI
>< MboI
>< MmiI
>< DpnII
>< TfiI >< DpnI

```

FIGURE 13. 48

```

                >< BstYI                > < TfiI
                >< BspAI                > < HinfI
                >< HinfI>< Bsp143I        >< Esp3I        >< Tru9I
                >< BsiBI        >< Tth111I    >< BsmBI        >< MseI
                >< BsaBI        >< BsmAI        >< BsmAI
    >< BsrI        >< TaqI >< BglII    >< AspI        >< Alw26I    >< HgaI> < Alw26I
TGGCACACTA CTTGTCGATT CAGATCTTAA TGACTTCGTC TCCGACGCAG ATTCTACTTT AATTGGAGAC
    20870        20880        20890        20900        20910        20920        20930

                >< StyI
                >< SinI
                >< Sau96I
                >< SinI
                >< Sau96I
                >< PssI
                >< Psp5II
                >< PpuMI
                >< NspIV
                >< NspHII
                >< NlaIV
                >< EcoO109I
                >< Eco47I
                >< DraII
                >< Cfr13I
                >< Bsi2I
                >< BscBI
                >< RsaI
                >< Csp6I
                >< AfaI
                >< Bme18I
                >< AvaII
                >< AsuI
                >< AflIII ><
TGTGCAACAG TACATACGGC TAATAAATGG GACCTTATTA TTAGCGATAT GTATGACCCT AGGACCAAAC
    20940        20950        20960        20970        20980        20990        21000

    >< NspI
    >< NspHI
    >< NlaIII >< PleI
    >< MaeIII    >< HinfI
ATGTGACAAA AGAGAATGAC TCTAAAGAAG GGTTTTTCAC TTATCTGTGT GGATTATATA AGCAAAACT
    21010        21020        21030        21040        21050        21060        21070

    >< ScrFI
    >< MvaI
    >< EcoRII
    >< Ecl136I
    >< DsaV
    >< BstOI
    >< BstNI
    >< BsiLI
    >< BsaJI
    >< BsaJI    >< SfcI
    >< ApyI        >< AluI        >< BscCI        >< BscCIHindIII ><> AluI
AGCCCTGGGT GGTTCATAG CTGTAAAGAT AACAGAGCAT TCTTGAATG CTGACCTTTA CAAGCTTATG
    21080        21090        21100        21110        21120        21130        21140

                >< Zsp2I
                >< Ppu10I
    >< Pali
    >< HaeIII
    >< BsuRI
    >< BshI
    >< NlaIII>< AluI    >< BcgI    >< AvaIII >< SfaNIBcgI/a ><
GGCCATTTCT CATGGTGGAC AGCTTTTGTG ACAAATGTAA ATGCATCATC ATCGGAAGCA TTTTAAATTG
    21150        21160        21170        21180        21190        21200        21210

```

FIGURE 13.49

```

                                >< Zsp2I
                                >< SphI
                                >< Ppu10I
                                    >< PaeI
                                    >< NspI
                                    >< NspHI
                                    >< NsiI
                                    >< NlaIII
                                > < NlaIII
                                    >< Mph1103I
                                    >< EcoT22I
                                > < AvaIII >< MnlI
GGGCTAACTA TCTTGGCAAG CCGAAGGAAC AAATTGATGG CTATACCATG CATGCTAACT ACATTTTCTG
21220      21230      21240      21250      21260      21270      21280

                                Tru9I ><
                                >< Tru9I
                                >< GsuI      MseI ><
                                >< BsrI      >< MseI
                                >< BpmI      MnlI ><
                                >< BbsI      >< NlaIII >< MnlI
GAGGAACACA AATCCTATCC AGTTGTCTTC CTATTCACCTC TTTGACATGA GCAAATTTCC TCTTAAATTA
21290      21300      21310      21320      21330      21340      21350

                                >< Tru9I
                                >< MseI
                                >< Esp4I> < TfiI
                                >< BsmAI
                                >< Alw26I      Ksp632I ><
                                >< AflII> < HinfI      >< MboII >< EarI
                                >< AflII> < HinfI      Eam1104I ><
AGAGGAACTG CTGTAATGTC TCTTAAGGAG AATCAAATCA ATGATATGAT TTATTCTCTT CTGGAAAAAG
21360      21370      21380      21390      21400      21410      21420

                                >< Tru9I
                                >< MseI
                                >< HindII
                                >< HincII
                                >< HpaI AflIII >
GTAGGCTTAT CATTAGAGAA AACAAACAGAG TTGTGGTTTC AAGTGATATT CTTGTTAACA ACTAAACGAA
21430      21440      21450      21460      21470      21480      21490

                                >< VneI
                                >< SnoI
                                    >< SduI
                                    >< NspII
                                >< HpaII
                                    >< HgiAI
                                >< HapII
                                >< Cfr10I
                                    >< Bsp1286I
                                >< MspI>< BmyI
                                >< NspI      >< SpeI      >< ApaLI
                                >< NspHI      >< RmaI      >< Alw44I
                                >< NlaIII      >< MaeI >< MaeIII >< AgeI >< Alw21I
CATGTTTATT TTCTTATTAT TTCTTACTCT CACTAGTGGT AGTGACCTTG ACCGGTGCAC CACTTTTGAT
21500      21510      21520      21530      21540      21550      21560

                                > < AluI      >< MnlI
GATGTTCAAG CTCCTAATTA CACTCAACAT ACTTCATCTA TGAGGGGGGT TTACTATCCT GATGAAATTT
21570      21580      21590      21600      21610      21620      21630

                                >< Sau3AI

```

FIGURE 13. 50

```

>< NdeII
>< MboI
>< DpnII
  >< DpnI          >< Tru9I
>< BspAI          >< MseI > < MboII
  >< Bsp143I       >< DdeI          >< MaeIII
TTAGATCAGA CACTCTTTAT TTAACCTCAGG ATTTATTTCT TCCATTTTAT TCTAATGTGA CAGGGTTTCA
21640      21650      21660      21670      21680      21690      21700

>< VspI
>< Tru9I
>< MseI
>< AsnI          >< Tru9I          >< FokI
>< AseI >< MaeII >< MseI >< BbvI    > < Fnu4HI
TACTATTAAT CATACGTTTG GCAACCCTGT CATACCTTTT AAGGATGGTA TTTATTTTGC TGCCACAGAG
21710      21720      21730      21740      21750      21760      21770

          >< BslI
          >< DsaI>< BsiYI          >< NlaIII
          >< BsaJI          > < MaeIII
AAATCAAATG TTGTCCGTGG TTGGGTTTTT GGTCTACCA TGAACAACAA GTCACAGTGG GTGATTATTA
21780      21790      21800      21810      21820      21830      21840

          >< NspI
>< Tru9I          >< NspHI
>< MseI          >< NlaIII
>< HphI          >< MaeIII          >< MaeIII
TTAACAATTC TACTAATGTT GTTATACGAG CATGTAACCT TGAATTGTGT GACAACCCTT TCTTTGCTGT
21850      21860      21870      21880      21890      21900      21910

          >< StyI          >< Zsp2I
          >< NlaIII          >< Tru9I
>< NcoI >< RsaI          >< Ppu10I TthHB8I ><
>< EcoT14I          >< NsiI          >< TaqI
>< Eco130I          >< MseI          SfaNI ><
>< DsaI>< Csp6I          >< Mph1103I RsaI ><
>< BssT1I          >< TthHB8I >< EcoT22I Csp6I ><
>< BsaJI>< AfaI          >< TaqI >< AvaIII AfaI ><
TTCTAAACCC ATGGGTACAC AGACACATAC TATGATATTC GATAATGCAT TTAATTGCAC TTTTCGAGTAC
21920      21930      21940      21950      21960      21970      21980

          >< Tru9I
          >< MseI
          >< DraI
ATATCTGATG CCTTTTCGCT TGATGTTTCA GAAAAGTCAG GTAATTTTAA ACACCTTACGA GAGTTTGTGT
21990      22000      22010      22020      22030      22040      22050

          >< Sau3AI
          >< NdeII
          >< MboI
          >< DpnII
>< Tru9I          >< DpnI
>< MseI          >< BspAI
>< DraI          >< SfcI Bsp143I ><
TTAAAAATAA AGATGGGTTT CTCTATGTTT ATAAGGGCTA TCAACCTATA GATGTAGTTC GTGATCTACC
22060      22070      22080      22090      22100      22110      22120

          >< Tru9I
          > < Tru9I          >< MseI
          >< MseI          > < MseI          >< MnlI
TTCTGGTTTT AACACTTTGA AACCTATTTT TAAGTTGCCT CTTGGTATTA ACATTACAAA TTTTAGAGCC
22130      22140      22150      22160      22170      22180      22190

```

FIGURE 13.51

```

> < SduI>< SfcI
    >< PvuII
    >< Psp5I
> < NspII
    >< NspBII
> < MaeII > < Fnu4HI
> < Bsp1286I >< PstI
    >< BspMI > < BmyI>< Fnu4HI
    >< HphI >< BbvI >< AluI >< BbvI
ATTCTTACAG CCTTTTCACC TGCTCAAGAC ATTTGGGGCA CGTCAGCTGC AGCCTATTTT GTTGGCTATT
22200 22210 22220 22230 22240 22250 22260

    >< SfaNI
    >< RsaI
> < Csp6I
    >< AfaI >< AlwNI
TAAAGCCAAC TACATTTATG CTCAAGTATG ATGAAAATGG TACAATCACA GATGCTGTTG ATTGTTCTCA
22270 22280 22290 22300 22310 22320 22330

    > < Tru9I
    > < MseI
    >< AluI
AAATCCACTT GCTGAAGTCA AATGCTCTGT TAAGAGCTTT GAGATTGACA AAGGAATTTA CCAGACCTCT
22340 22350 22360 22370 22380 22390 22400

    >< SauI
    >< MstII
    >< Eco81I
    >< DdeI
    >< CvnI
    >< Bsu36I
    >< Bse21I
    >< AxyI >< TfiI
> < MnlI >< AocI >< MnlI >< HinfI >< SspI >< MnlI
AATTCAGGG TTGTTCCCTC AGGAGATGTT GTGAGATTCC CTAATATTAC AAACCTGTGT CCTTTTGAG
22410 22420 22430 22440 22450 22460 22470

    >< Zsp2I
    >< Ppu10I
    >< NsiI
    > < NlaIII
    >< Mph1103I
    >< EcoT22I
    >< Tru9I >< AvaIII
    >< MseI
AGGTTTTTAA TGCTACTAAA TTCCCTTCTG TCTATGCATG GGAGAGAAAA AAAATTTCTA ATTGTGTTGC
22480 22490 22500 22510 22520 22530 22540

    >< SduI
    >< NspII
    >< HgiAI
    >< Bsp1286I
    >< BmyI >< Tru9I
    >< Alw21I >< MseI DdeI ><
TGATTACTCT GTGCTCTACA ACTCAACATT TTTTCAACC TTAAAGTGCT ATGGCGTTTC TGCCACTAAG
22550 22560 22570 22580 22590 22600 22610

    >< Sau3AI
    >< NdeII
    >< MboI
    >< DpnII
    >< DpnI

```

FIGURE 1352

```

    >< BspAI
    >< Bsp143I
    TTGAATGATC TTGCTTCTC CAATGTCTAT GCAGATTCTT TTGTAGTCAA GGGAGATGAT GTAAGACAAA
    22620      22630      22640      22650      22660      22670      22680

    >< ScrFI
    >< MvaI
    >< HinfII
    >< HinfI
    >< HhaI
    >< HaeII
    >< EcoRII
    >< Ecl136I
    >< DsaV
    >< CfoI
    >< BstOI
    >< BstNI
    >< Bsp143II
    >< BsiLI
    >< ApyI      >< BsrI
    TAGCGCCAGG ACAAACGGT GTTATTGCTG ATTATAATTA TAAATTGCCA GATGATTCCA TGGGTTGTGT
    22690      22700      22710      22720      22730      22740      22750

    >< SfaNI
    >< RmaI
    >< MaeI
    >< BsrI
    >< NlaIII
    CCTTGCTTGG AATACTAGGA ACATTGATGC TACTTCAACT GGTAATTATA ATTATAAATA TAGGTATCTT
    22760      22770      22780      22790      22800      22810      22820

    >< Sau96I
    >< Pali
    >< NspIV
    >< HindIII
    >< HaeIII
    >< EcoO109I
    >< DraII
    >< DdeI
    >< Cfr13I
    >< BsuRI
    >< BsiZI
    >< BshI
    >< BfrI >< PssI
    >< NlaIII >< AsuI>< BsmAI
    >< AluI      >< Alw26I
    AGACATGGCA AGCTTAGGCC CTTTGAGAGA GACATATCTA ATGTGCCTTT CTCCCCTGAT GGCAAACCTT
    22830      22840      22850      22860      22870      22880      22890

    >< Tru9I
    >< Pali
    >< MscI
    >< HaeIII
    >< EaeI>< MseI
    >< Tru9I      >< BsuRI
    >< MseI      >< BshI
    >< BspMI      >< BalI
    GCACCCACCC TGCTCTTAAT TGTTATTGGC CATTAAATGA TTATGGTTTT TACACCACTA CTGGCATTGG
    22900      22910      22920      22930      22940      22950      22960

    Sau96I ><
    >< PalINspIV ><
    >< MspI NspHII ><
    >< HaeIII

```

FIGURE 13.53

```

                                > < HpaII Eco47I ><
                                >< DsaI
                                > < HapII Cfr13I ><
                                >< BsuRISinI ><
                                >< GdiII BsiZI ><
                                >< BsaJI
                                >< ScaI
                                >< RsaI
                                >< Csp6I
                                >< AfaI
                                >< Tru9I
                                >< EaeI Bmel8I ><
                                >< MseI >< Cfr10I AvaII ><
                                >< DraI
                                >< BshI AsuI ><
CTACCAACCT TACAGAGTTG TAGTACTTTC TTTTGAACCT TTAATGTCAC CGGCCACGGT TGTGGACCA
22970 22980 22990 23000 23010 23020 23030

                                >< Tru9I
                                >< RsaI
                                >< Tru9I
                                >< Csp6I
                                >< PleI
                                >< BsrI ><
                                >< MseI
                                >< BsrI
                                > < Tru9I
                                > < MseI>< BsrI
                                >< MseI
                                >< HinfI
                                >< AfaI
AAATTATCCA CTGACCTTAT TAAGAACCAG TGTGTCAATT TTAATTTTAA TGGACTCACT GGTACTGGTG
23040 23050 23060 23070 23080 23090 23100

                                >< Tru9I
                                >< Pali
                                >< MseI
                                >< HaeIII
                                >< MboII
                                >< GdiII
                                >< HpaI
                                >< EaeI
                                >< HindII
                                >< BsuRI
                                >< BshI
                                >< HincII
                                >< TfiI ><
                                >< HinfI ><
TGTTAACTCC TTCTTCAAAG AGATTTC AAC CATTTC AACA ATTTGGCCGT GATGTTTCTG ATTTCACTGA
23110 23120 23130 23140 23150 23160 23170

                                > < XhoII
                                >< TthHB8I
                                >< TaqI
                                > < Sau3AI
                                > < NdeII
                                > < MflI
                                > < MboI
                                > < DpnII
                                >< DpnI
                                > < BstYI
                                > < BspAI
                                > < SspI
                                >< AlwI >< Bsp143I
                                >< HphI
TTCCGTTTCA GATCCTAATA CATCTGAAAT ATTAGACATT TCACCTTGCT CTTTGGGGG TGTAAGTGTA
23180 23190 23200 23210 23220 23230 23240

                                >< ScrFI
                                >< MvaI
                                >< EcoRII
                                >< Ecl136I
                                >< DsaV
                                >< BstOI
                                >< BstNI
                                >< BsiLI
                                >< ApyI
                                >< Tru9I
                                >< MseI
                                >< HpaI
                                >< HindII
                                >< Eco57I
                                >< BsgI
                                >< HincII
ATTACACCTG GAACAAATGC TTCATCTGAA GTTGCTGTTC TATATCAAGA TGTTAACTGC ACTGATGTTT
23250 23260 23270 23280 23290 23300 23310

                                >< Sau3AI
                                >< NlaIII
                                >< NdeII
                                >< MboI
                                >< DpnII
                                >< DpnI
                                >< HinfII

```

FIGURE 13. 54


```

                >< BspWI                >< Hin6I
                >< BspAI                > < HhaI                PleI ><
                >< SfcI                >< Bsp143I                >< AluI> < CfoI                >< BsrI
CTACAGCAAT TCATGCAGAT CAACTCACAC CAGCTTGGCG CATATATTCT ACTGGAAACA ATGTATTCCA
    23320      23330      23340      23350      23360      23370      23380

                >< TthHB8I
                >< TaqI
                >< SalI
                >< RtrI
                >< NspI
                >< EspI >< NspHI
                >< DdeI >< NlaIII
                >< CelII >< HindII
                >< Bpu102I>< HincII
>< HinfI                >< AluI                >< AccI
GACTCAAGCA GGCTGTCTTA TAGGAGCTGA GCATGTCGAC ACTTCTTATG AGTGGGACAT TCCTATTGGA
    23390      23400      23410      23420      23430      23440      23450

                > < SnaBI
                >< ScaI
                >< RsaI
                >< RmaI
                >< MaeII >< MaeI
                > < Eco105I
                >< Csp6I
                > < BsaAI
                >< AfaI
>< AluI                >< MaeI
GCTGGCATTG GTGCTAGTTA CCATACAGTT TCTTTATTAC GTAGTACTAG CCAAAAATCT ATTGTGGCTT
    23460      23470      23480      23490      23500      23510      23520

                >< MniI
ATACTATGTC TTTAGGTGCT GATAGTTCAA TTGCTTACTC TAATAACACC ATTGCTATAC CTAATAACTT
    23530      23540      23550      23560      23570      23580      23590

                RsaI ><
                >< MnlI
                Csp6I ><
                AfaI ><
                >< SfcI
TTCAATTAGC ATTACTACAG AAGTAATGCC TGTTCCTATG GCTAAAACCT CCGTAGATTG TAATATGTAC
    23600      23610      23620      23630      23640      23650      23660

                > < TfiI
                > < HinfI
                >< AciI                > < AluI
ATCTGCGGAG ATTCTACTGA ATGTGCTAAT TTGCTTCTCC AATATGGTAG CTTTTCACACA CAACTAAATC
    23670      23680      23690      23700      23710      23720      23730

>< VneI
>< SduI
>< NspII
>< HgiAI
>< SnaI>< DdeI                >< Sau3AI                >< PmlI
>< Bsp1286I                >< NdeII                >< MaeII
>< BmyI                >< MboI                >< Eco72I
>< BbvI                >< DpnI                >< BsaAI
>< ApaLI                >< Bsp143I                >< BbrPI
>< Alw44I                >< DpnII >< AlwI
>< Alw21I                >< Fnu4HI >< BspAI                >< AflIII
GTGCACTCTC AGGTATTGCT GCTGAACAGG ATCGCAACAC ACGTGAAGTG TTCGCTCAAG TCAAACAAAT
    23740      23750      23760      23770      23780      23790      23800

```

FIGURE 13.55

```

>< RsaI
>< Csp6I
>< AfaI
GTACAAAACC CCAACTTTGA AATATTTTGG TGGTTTAAAT TTTTCACAAA TATTACCTGA CCCTCTAAAG
23810 23820 23830 23840 23850 23860 23870

>< MnlI
>< MnlI
>< DdeI >< MnlI
CCAATAAGA GGTCTTTTAT TGAGGACTTG CTCTTTAATA AGGTGACACT CGCTGATGCT GGCTTCATGA
23880 23890 23900 23910 23920 23930 23940

>< XhoII
>< Sau3AI
>< StyI
>< RmaI
>< MaeI
>< EcoT14I
>< Eco130I
>< BssTII
>< BsmI
>< BscCI
>< BsaJI
>< BlnI
>< AvrII
>< RmaI
>< NdeII
>< MflI
>< MboI
>< MaeI
>< VspI
>< Tru9I
>< MseI
>< AsnI
>< AseI
>< RmaI
>< NdeII
>< MflI
>< MboI
>< MaeI
>< VspI
>< Tru9I
>< MseI
>< AsnI
>< AseI
>< XhoII
>< Sau3AI
>< StyI
>< RmaI
>< MaeI
>< EcoT14I
>< Eco130I
>< BssTII
>< BsmI
>< BscCI
>< BsaJI
>< BlnI
>< AvrII
>< MstI
>< HinPII
>< Hin6I
>< HhaI
>< FspI
>< FdiII
>< CfoI
>< AviII
AGCAATATGG CGAATGCCTA GGTGATATTA ATGCTAGAGA TCTCATTTGT GCGCAGAAGT TCAATGGACT
23950 23960 23970 23980 23990 24000 24010

>< RmaIRsaI ><
>< MnlI >< Fnu4HI >< Fnu4HI Csp6I ><
>< BspWI >< BbvI >< BbvI >< BspWI >< MaeIAfaI ><
TACAGTGTG CCACCTCTGC TCACTGATGA TATGATTGCT GCCTACACTG CTGCTCTAGT TAGTGGTACT
24020 24030 24040 24050 24060 24070 24080

>< MboII
>< HinPII
>< Hin6I
>< HhaI
>< HaeII
>< Fnu4HI >< Ksp632I
>< CfoI >< EarI
>< FokI >< BspWI >< Eam1104I
>< BbvI >< Bsp143II
GCCACTGCTG GATGGACATT TGGTGCTGGC GCTGCTCTTC AAATACCTTT TGCTATGCAA ATGGCATATA
24090 24100 24110 24120 24130 24140 24150

>< MaeIII
>< MaeIII
GGTTCAATGG CATTGGAGTT ACCCAAAATG TTCTCTATGA GAACCAAAAA CAAATCGCCA ACCAATTTAA
24160 24170 24180 24190 24200 24210 24220

>< TfiI
>< HinfI
>< Fnu4HI
>< BbvI
>< AluI
CAAGGCGATT AGTCAAATTC AAGAATCACT TACAACAACA TCAACTGCAT TGGGCAAGCT GCAAGACGTT
24230 24240 24250 24260 24270 24280 24290

>< Tru9I
>< MseI
>< HpaI
>< HindII
>< HincII>< BscCI
>< DdeI
>< Tru9I >< BfrI
>< MseI >< AluI

```

FIGURE 13. 56

```

GTTAACCAGA ATGCTCAAGC ATTAACACACA CTTGTTAAAC AACTTAGCTC TAATTTTGGT GCAATTTCAA
24300      24310      24320      24330      24340      24350      24360

>< ThaI
>< SpoI
>< NruI
>< MvnI
>< BstUI >< TthHB8I
>< Bsp68I >< TaqI >< RsaI
>< EcoRV >< Bsp50I >< MnlI >< Csp6I >< Tru9I
>< Eco32I >< AccII >< MnlI >< AciI>< AfaI >< MseI
GTGTGCTAAA TGATATCCTT TCGCGACTTG ATAAAGTCGA GCGGAGGTA CAAATTGACA GTTAATTAC
24370      24380      24390      24400      24410      24420      24430

>< MaeIII >< BbvI >< Fnu4HI BbvI ><
AGGCAGACTT CAAAGCCTTC AAACCTATGT AACACAACAA CTAATCAGGG CTGCTGAAAT CAGGCTTCT
24440      24450      24460      24470      24480      24490      24500

>< Fnu4HI >< HindII
>< BspWI >< DdeI >< HincII
GCTAATCTTG CTGCTACTAA AATGTCTGAG TGTGTTCTTG GACAATCAAA AAGAGTTGAC TTTCTGGAA
24510      24520      24530      24540      24550      24560      24570

>< NspI
>< NspHI
>< NlaIII
>< MaeIII
>< NlaIII >< MaeII
>< MboII >< FokI
>< Fnu4HI >< BbsI BsaAI ><
>< AciI>< BbvI >< AflIII
AGGGCTACCA CCTTATGTCC TTCCACAAG CAGCCCCGCA TGGTGTGTC TTCCTACATG TCACGTATGT
24580      24590      24600      24610      24620      24630      24640

>< ScrFI
>< MvaI
>< EcoRII
>< Ecl136I
>< BstOI
>< BstNI >< HinPII
>< MnlI >< BslI >< Hin6I
>< DsaV>< BsiYI >< HhaI
>< BsiLI >< HaeII
>< BsaJI>< HphI >< CfoI >< NlaIII
>< ApyI >< Bsp143II >< BspHI EcoNI ><
GCCATCCCAG GAGAGGAACT TCACCACAGC GCCAGCAATT TGTCATGAAG GCAAAGCATA CTTCCCTCGT
24650      24660      24670      24680      24690      24700      24710

>< MnlI
>< BslI >< Tru9I
>< BsiYI >< MseI >< MnlI
GAAGGTGTTT TTGTGTTTAA TGGCACTTCT TGGTTTATTA CACAGAGGAA CTTCTTTTCT CCACAAATAA
24720      24730      24740      24750      24760      24770      24780

>< DdeI >< Tru9I
>< BsmAI >< SfaNI
>< SfcI >< Alw26I >< MseIAlwI ><
T TACTACAGA CAATACATTT GTCTCAGGAA ATTGTGATGT CGTTATTGGC ATCATTAACA ACACAGTTTA
24790      24800      24810      24820      24830      24840      24850

>< Sau3AI
>< NdeII

```

FIGURE 13.57

```

>< MboI          >< P1eI          > < ScaI
>< DpnII         >< MnlI          > < Ksp632I      > < RsaI
  >< DpnI         >< DdeI >< HinfI      >< MboII
>< BspAI         >< BspWI         > < Eam1104I      >< Csp6I
  >< Bsp143I      >< AluI          > < EarI    > < AluI    > < AfaI    > < HphI
TGATCCTCTG CAACCTGAGC TTGACTCATT CAAAGAAGAG CTGGACAAGT ACTTCAAAAA TCATACATCA
  24860      24870      24880      24890      24900      24910      24920

  >< Sau3AI
  >< NdeII
  >< MboI
>< MmI
  >< DpnII
  >< DpnI
  >< BspAI
  >< Bsp143I
  >< BsiBI          >< Tru9I          >< HindII
  >< BsaBI          >< MseI          >< HincII      AciI ><
CCAGATGTTG ATCTTGGCGA CATTTCAGGC ATTAACGCTT CTGTCGTCAA CATTCAAAAA GAAATTGACC
  24930      24940      24950      24960      24970      24980      24990

  >< Tru9I
  > < TfiI
  >< MnlI          >< SwaI
  >< EcoNI         >< MseI
  >< BslI          > < HinfI
>< MnlI>< BsiYI      >< DraI
GCCTCAATGA GGTGCTAAA AATTTAAATG AATCACTCAT TGACCTTCAA GAATTGGGAA AATATGAGCA
  25000      25010      25020      25030      25040      25050      25060

  >< StyI
  >< Pali
  >< HaeIII
  >< EcoT14I
  >< Eco130I
  >< BsuRI
  >< BssTII
  >< Tru9I>< BshI          NlaIII ><
  >< MseI >< BsaJI          MaeIII ><
  >> BstXI
ATATATTAAA TGGCCTTGGT ATGTTTGGCT CGGCTTCATT GCTGGACTAA TTGCCATCGT CATGGTTACA
  25070      25080      25090      25100      25110      25120      25130

  > < SphI
  > < PaeI
  >< SpeI          > < NspI
  > < RmaI          > < NspHI
  >< NlaIII        > < NlaIII
  > < MaeI          >< MnlI>< BbvI Fnu4HI ><
ATCTTGCTTT GTTGCAAGAC TAGTTGTTGC AGTTGCCTCA AGGGTGCATG CTCTTGTGGT TCTTGCTGCA
  25140      25150      25160      25170      25180      25190      25200

  >< FokI
  >< DdeI
>< MnlI >< P1eI>< HinfI >< BsrI
AGTTTGATGA GGATGACTCT GAGCCAGTTC TCAAGGGTGT CAAATTACAT TACACATAAA CGAACTTATG
  25210      25220      25230      25240      25250      25260      25270

  >< Sau3AI
  >< NdeII
  >< MboI
  >< DpnII
  > < DpnI

```

FIGURE 13.58

```

                >< BspAI
                > < Bsp143I
                >< BsgI      >< AlwI      >< BsrI      BspWI >
GATTTGTTTA TGAGATTTT TACTCTTGGA TCAATTACTG CACAGCCAGT AAAAATTGAC AATGCTTCTC
25280      25290      25300      25310      25320      25330      25340

                >< ScaI
                >< RsaI
                >< Csp6I      >< SfcI
                >< AfaI      >< NlaIII      >< AciI      >< MnlI      FokI >
CTGCAAGTAC TGTTCATGCT ACAGCAACGA TACCGCTACA AGCCTCACTC CCTTTCGGAT GGCTTGTTAT
25350      25360      25370      25380      25390      25400      25410

                > < HinPII
                > < Hin6I
                >< HhaI
                >< HaeII      >< HinPII      RmaI ><
                >< Eco47III      >< Hin6I      NheI ><
                >< CfoI      >< HhaI      MaeI ><
                >< BspWI      >< Bsp143II      Fnu4HI ><
                >< CfoI      AluI ><
TGGCGTTGCA TTTCTTGCTG TTTTTCAGAG CGCTACCAAA ATAATTGCGC TCAATAAAAG ATGGCAGCTA
25420      25430      25440      25450      25460      25470      25480

                >< EcoNI
                >< BslI
                >< BsiYI
                >< BbvI      >< BsrI      >< BbvI      > < Fnu4HI      BbvI ><
GCCCTTTATA AGGGCTTCCA GTTCATTTGC AATTTACTGC TGCTATTTGT TACCATCTAT TCACATCTTT
25490      25500      25510      25520      25530      25540      25550

                >< SfcI      >< HinPII
                >< PstI      >< Hin6I      >< RsaI      NsiI ><
                > < Fnu4HI      >< HhaI      >< Csp6I      Mph1103I ><
                >< BspMI      >< MnlI      >< CfoI      >< AfaI      >< MnlI      EcoT22I ><
                >< BspMI      >< MnlI      >< CfoI      >< AfaI      >< MnlI      AvaIII ><
TGCTTGTCGC TGCAGGTATG GAGGCGCAAT TTTTGTACCT CTATGCCTTG ATATATTTTC TACAATGCAT
25560      25570      25580      25590      25600      25610      25620

                >< SfaNI
                >< NspI
                >< NspHI
                >< NlaIII
                >< SfaNI
CAACGCATGT AGAATTATTA TGAGATGTTG GCTTTGTTGG AAGTGCAAAT CCAAGAACCC ATTACTTTAT
25630      25640      25650      25660      25670      25680      25690

                >< Bst1107I
                >< AccI      MaeIII ><
GATGCCAACT ACTTTGTTTG CTGGCACACA CATAACTATG ACTACTGTAT ACCATATAAC AGTGTACACG
25700      25710      25720      25730      25740      25750      25760

                >< MboII
                >< HphI      BstXI ><
                >< MunI >< MaeIII >< MaeIII >< Eco57I >< BbsI MnlI >
ATACAATTGT CGTTACTGAA GGTGACGGCA TTTCAACACC AAAACTCAAA GAAGACTACC AAATTGGTGG
25770      25780      25790      25800      25810      25820      25830

                >< RsaI
                > < NlaIII
                >< HphI
                >< Tru9I >< Tth111I >< Csp6I
                >< DdeI      >< DdeI >< MseI >< AspI >< AfaI

```

FIGURE 13.59

```

TTATTCTGAG GATAGGCACT CAGGTGTTAA AGACTATGTC GTTGATCATG GCTATTTTAC CGAAGTTTAC
25840      25850      25860      25870      25880      25890      25900

      > < HinfI>< PleI      >< BsrI      Tru9I ><
      >< AluI >< AccI      >< SfcI >< AlwNI      MseI ><
TACCAGCTTG AGTCTACACA AATTACTACA GACACTGGTA TTGAAAATGC TACATTCTTC ATCTTTAACA
25910      25920      25930      25940      25950      25960      25970

      > < TthHB8I
      >< Tru9I      > < TaqI      >< Ksp632I
      >< MseI      > < MboII      >< EarI BspWI ><
      >< AluI      >< Eco57I      >< Eam1104I AlwI ><
AGCTTGTTAA AGACCCACCG AATGTGCAAA TACACACAAT CGACGGCTCT TCAGGAGTTG CTAATCCAGC
25980      25990      26000      26010      26020      26030      26040

      >< XhoII
      >< Sau3AI
      >< NlaIV
      >< NdeII
      >< MflI
      >< MboI
      >< DpnII
      >< DpnI
      >< BstYI
      >< BstI
      >< BspAI
      >< Bsp143I
      >< BscBI
      >< BamHI >< AlwI      >< RmaI      RsaI ><
      >< BamHI >< AlwI      >< MaeI      Csp6I ><
AATGGATCCA ATTTATGATG AGCCGACGAC GACTACTAGC GTGCCTTTGT AAGCACAAGA AAGTGAGTAC
26050      26060      26070      26080      26090      26100      26110

      > < Tru9I
      >< RsaI
      > < MseI
      >< MboII
      > < RsaI      >< MaeII      >< RsaI
      >< Csp6I      >< Csp6I      >< Tru9I >< Csp6I
      > < AfaI      >< AfaI      >< MseI >< AfaI
GAACTTATGT ACTCATTCGT TTCGGAAGAA ACAGGTACGT TAATAGTTAA TAGCGTACTT CTTTTCTTG
26120      26130      26140      26150      26160      26170      26180

      >< TthHB8I
      >< TaqI
      >< RmaI      >< HinPII      > < RsaI
      > < MaeIII      >< Hin6I      Fnu4HI ><
      >< MaeI >< RmaI      >< HhaI      >< Csp6I
      >< FokI >< MaeI      >< CfoI >< BbvI > < AfaI
CTTTCGTGGT ATTCTTGCTA GTCACACTAG CCATCCTTAC TGCGCTTCGA TTGTGTGCGT ACTGCTGCAA
26190      26200      26210      26220      26230      26240      26250

      >< Tru9I
      >< MseI
      >< SspI >< MaeII
      >< HpaI
      >< HindII
      >< HincII
      >< Tru9I
      >< ThaI
      >< MvnI
      >< MseI
      >< BstUI      Ksp632I >
      >< MaeII >< Bsp50I >< MboII EarI >
      >< AccI >< AccII      Eam1104I >
TATTGTTAAC GTGAGTTTAG TAAAACCAAC GGTTCACGTC TACTCGCGTG TTAATAATCT GAACCTCTCT
26260      26270      26280      26290      26300      26310      26320

```

FIGURE 13.60

```

>< Sau3AI
>< NdeII
>< MboI
>< DpnII
>< MboII>< DpnI
>< XmnI >< BspAI> < Eco57I
>< Asp700I>< Bsp143I
GAAGGAGTTC CTGATCTTCT GGTCTAAACG AACTAACTAT TATTATTATT CTGTTTGGAA CTTTAACATT
26330      26340      26350      26360      26370      26380      26390

>< ScrFI
>< MvaI
>< EcoRII
>< Ecl136I
>< DsaV NlaIV ><
>< RsaI
>< MnlI
>< Tru9I
>< BstOI
>< BstNI RmaI ><
>< Csp6I
>< MseI
>< BsiLI MaeI ><
> < NlaIII >< AfaI > < AluI >< ApyIBscBI ><
GCTTATCATG GCAGACAACG GTACTATTAC CGTTGAGGAG CTTAAACAAC TCCTGGAACA ATGGAACCTA
26400      26410      26420      26430      26440      26450      26460

>< ScrFI
>< RmaI
>< MvaI
>< MaeI
>< EcoRII
>< Ecl136I
>< DsaV
>< BstOI
>< BstNI
>< BsiLI
>< ApyI >< MaeIII
GTAATAGGTT TCCTATTCCT AGCCTGGATT ATGTTACTAC AATTGCGCTA TTCTAATCGG AACAGGTTTT
26470      26480      26490      26500      26510      26520      26530

>< Pali
>< MscI
>< MnlI >< MaeIII
>< HaeIII
>< EaeI
>< BsuRI
>< BsrI
>< RsaI
>< Csp6I >< HindIII
>< AfaI >< AluI
>< BspWI
>< BshI
>< BalI
>< BbvI Fnu4HI ><
TGTACATAAT AAAGCTTGTT TTCCTCTGGC TCTTGTGGCC AGTAACACTT GCTTGTTTTG TGCTTGCTGC
26540      26550      26560      26570      26580      26590      26600

>< VspI
>< Tru9I
>< MseI
>< SfcI >< AsnI >< BsrI
>< AccI >< AseI>< MaeIII>< AciI
TGTTCTACAGA ATTAATTGGG TGACTGGCGG GATTGCGATT GCAATGGCTT GTATTGTAGG CTTGATGTGG
26610      26620      26630      26640      26650      26660      26670

>< EspI
>< Eco57I
>< DdeI
>< CelII
>< Bpu102I
>< RsaI
>< Csp6I

```

FIGURE 13.61

```

>< BfrI
>< AluI
CTTAGCTACT TCGTTGCTTC CTTCAGGCTG TTTGCTCGTA CCCGCTCAAT GTGGTCATTC AACCCAGAAA
26680      26690      26700      26710      26720      26730      26740

>< AfaI
>< AciI
MboII >
>< ScrFI
>< NciI
>< MspI
>< HpaII
>< HapII
>< DsaV>< MnlI
>< BslI
>< BsiYI
>< BsaJI >< MniI      > < XcmI
>< BcnI      >< MaeIII >< AciI >< NlaIII
CAAACATTCT TCTCAATGTG CCTCTCCGGG GGACAATTGT GACCAGACCG CTCATGGAAA GTGAAGTTGT
26750      26760      26770      26780      26790      26800      26810

Tru9I ><
SinI >
Sau96I >
PpuMI >
NspIV >
MseI ><
>< MaeIII
>< Sau3AI
>< NdeII
>< MboI
>< FbaI
>< DpnII
>< DpnI
>< BspAI
>< Bsp143I
>< BsiQI
>< BclI
>< PstI >< BspMI
>< Psp5II
>< NspHII
GACCTGCCAA AAGAGATCAC TGTGGCTACA TCACGAACGC TTTCTTATTA CAAATTAGGA GCGTCGCAGC
26890      26900      26910      26920      26930      26940      26950

>< TfiI
>< HinfI
>< BbvI
>< BbvI
>< Fnu4HI >< AciI
GTGTAGGCAC TGATTGAGGT TTTGCTGCAT ACAACCGCTA CCGTATTGGA AACTATAAAT TAAATACAGA
26960      26970      26980      26990      27000      27010      27020

>< MspI
>< HpaII
>< HapII
>< Cfr10I
>< BcgI/a
>< RsaI
>< RmaI
>< Csp6I
>< MaeI>< BcgI
>< AfaI >< MaeIII
HindII ><
HincII ><

```

FIGURE 13.62


```

CCACGCCGGT AGCAACGACA ATATTGCTTT GCTAGTACAG TAAGTGACAA CAGATGTTTC ATCTTGTTGA
 27030      27040      27050      27060      27070      27080      27090

>< ScrFI
>< MvaI
  >< MaeIII
>< EcoRII
  >< Ecl136I
>< DsaV
  >< BstOI
  >< BstNI
  >< BsiLI
  >< ApyI
                                >< MnlI
                                HinfI ><
CTTCCAGGTT ACAATAGCAG AGATATTGAT TATCATTATG AGGACTTTCA GGATTGCTAT TTGGAATCTT
 27100      27110      27120      27130      27140      27150      27160

                                >< BsmAI
                                >< Tru9I
                                > < MnlI
>< MaeII
  >< Alw26I
  >< MseI
  >< DdeI
                                >< MboII
GACGTTATAA TAAGTTCAAT AGTGAGACAA TTATTTAAGC CTCTAACTAA GAAGAATTAT TCGGAGTTAG
 27170      27180      27190      27200      27210      27220      27230

                                >< Ksp632I
                                >< MboII
                                >< EarI
  >< MboII
                                >< NlaIII Eam1104I ><
ATGATGAAGA ACCTATGGAG TTAGATTATC CATAAAACGA ACATGAAAAT TATTCTCTTC CTGACATTGA
 27240      27250      27260      27270      27280      27290      27300

                                > < RsaI >< RsaI
                                >< Csp6I >< Csp6I
  > < AluI
                                >< MnlI
                                > < AfaI >< AfaI
TTGTATTTAC ATCTTGCGAG CTATATCACT ATCAGGAGTG TGTAGAGGT ACGACTGTAC TACTAAAAGA
 27310      27320      27330      27340      27350      27360      27370

                                >< MnlI
                                >< HphI
                                >< HphI
                                >< MnlI
ACCTTGCCCA TCAGGAACAT ACGAGGGCAA TTCACCATT T CAGCCTCTTG CTGACAATAA ATTGCACTA
 27380      27390      27400      27410      27420      27430      27440

                                Sau3AI >
                                > < PvuII
                                > < Psp5I
                                > < NspBII
                                >< TthHB8I
                                >< TaqI
                                >< RsaI
                                >< Csp6I
                                >< BbvI
                                >< AfaI
                                >< Fnu4HI
                                >< DpnII
                                >< BspAI
                                >< AluI
>< RmaI
>< MaeI
ACTTGCACTA GCACACACTT TGCTTTTGCT TGTGCTGACG GTACTCGACA TACCTATCAG CTGCGTGCAA
 27450      27460      27470      27480      27490      27500      27510

                                >< SstI
                                >< SduI
                                >< SacI
                                >< NspII
                                >< HgiAI
                                >< Eco24I
                                > < Ecl136II
                                >< BspWI
                                >< Bsp1286I
                                >< BmyI
                                >< BanII
                                >< Alw21I
>< HphI
>< DpnI
                                >< MnlI

```

FIGURE 13. 63

```

>< Bsp143I          >< MnlI          > < AluI    BbvI ><
GATCAGTTTC ACCAAACTT TTCATCAGAC AAGAGGAGGT TCAACAAGAG CTCTACTCGC CACTTTTCTT
27520      27530      27540      27550      27560      27570      27580

SstI ><
SduI ><
SacI ><
NspII ><
HgiAI ><
Eco24I ><
Ecl136II ><
Bsp1286I ><
BmyI ><
BanII ><
Alw21I ><
AluI ><
>< RmaI    >< Tru9I
>< MaeI    >< MseI          >< Tru9I
>< Fnu4HI   >< HphI          >< MseI          AluI ><
CATTGTTGCT GCTCTAGTAT TTTTAATACT TTGCTTCACC ATTAAGAGAA AGACAGAATG AATGAGCTCA
27590      27600      27610      27620      27630      27640      27650

>< Tru9I          >< Tru9I
>< MseI          >< MseI
CTTTAATTGA CTTCTATTTG TGCTTTTTAG CCTTCTGCT ATTCCTTGTT TTAATAATGC TTATTATATT
27660      27670      27680      27690      27700      27710      27720

>< XhoII
>< XbaI
> < ScrFI
>< Sau3AI
>< RmaI
>< NdeII
> < MvaI
>< MflI
>< MboI
>< EcoRI>< MaeI
> < Ecl136I
>< DpnII
>< DpnI
>< BstYI
> < BstOI
> < BstNI
>< TthHB8I >< BspAI          > < RsaI
>< DsaV>< Bsp143I          >< MboII
> < BsiLI          >< Csp6I
>< TaqI > < ApyI > < AlwI > < AfaI          >< NlaIII
TTGGTTTTCa CTCGAAATCC AGGATCTAGA AGAACCTTGT ACCAAAGTCT AAACGAACAT GAAACTTCTC
27730      27740      27750      27760      27770      27780      27790

>< HinP1I
>< Hin6I
>< HhaI
>< RsaI >< HaeII
>< SfcI          >< Eco47III
>< Csp6I>< CfoI SfaNI ><
>< AfaI >< Bsp143II
ATTGTTTTCa CTTGTATTTT TCTATGCAGT TGCATATGCA CTGTAGTACA GCGCTGTGCA TCTAATAAAC
27800      27810      27820      27830      27840      27850      27860

>< XhoII
>< Sau3AI
>< NdeII
> < MnlI
>< MflI

```

FIGURE 13.64

```

    >< MboI
    >< DpnII
    >< DpnI    >< RsaI
    >< BstYI    >< MboII
    >< NlaIII>< BspAI    >< Csp6I >< RmaI
    >< AlwI >< Bsp143I    >< AfaI >< MaeI
CTCATGTGCT TGAAGATCCT TGTAAAGGTAC AACACTAGGG GTAATACTTA TAGCACTGCT TGGCTTTGTG
27870      27880      27890      27900      27910      27920      27930

>< SduI
>< RmaI
>< NspII
>< MaeI
>< HgiAI
>< Bsp1286I
>< BmyI
>< Alw21I
CTCTAGGAAA GGTTTTACCT TTTCATAGAT GGCACACTAT GGTTCAAACA TGCACACCTA ATGTTACTAT
27940      27950      27960      27970      27980      27990      28000

    > < XhoII
    > < Sau3AI > < Van91I
    >< PvuII
    >< Psp5I
    > < NdeII > < PflMI
    > < MflI>< NspBII
    > < DpnII    >< HinPII
    >< Bsp143I    >< Hin6I
    > < BstYI > < BslI >< HhaI >< RmaI
    > < BspAI > < BsiYI>< CfoI >< MaeI
    > < MboI>< AluI>< BspWI >< BspWI
    >< AlwI >< DpnI > < AccB7I >< AluI
CAACTGTCAA GATCCAGCTG GTGGTGGCTG TATAGCTAGG TGTGGTACC TTCATGAAGG TCACCAAACT
28010      28020      28030      28040      28050      28060      28070

    >< RsaI
    >< NlaIV
    >< KpnI >< NlaIII
    >< Eco64I    >< MaeIII
    >< Csp6I>< HphI
    >< BscBI    >< EcoO65I
    >< BanI >< BspHI
    >< Asp718    >< Eco91I
    >< AfaI    >< BstPI
    >< AccB1I    >< BstEII
    >< Acc65I    >< BbvI

    >< SinI
    >< Sau96I
    >< NspIV
    NspHII ><
    NlaIV ><
    >< Eco47I
    >< Cfr13I
    >< BsiZI
    BscBI ><
    >< Bme18I
    >< AvaII
    >< AsuI

    >< RsaI
    >< MaeII
    >< Fnu4HI
    >< Esp3I    >< Csp6I    >< Tru9I
    >< BsmAI    >< BsmBI    >< MseI
    >< Alw26I    >< AfaI    >< DraI
    >< MseI
GCTGCATTTA GAGACGTACT TGTGTTTTTA AATAAACGAA CAAATTAAAA TGTCTGATAA TGGACCCCAA
28080      28090      28100      28110      28120      28130      28140

    >< SinI
    >< Sau96I
    >< NspIV
    >< NspHII
    >< NlaIV
    >< Eco47I
    >< Cfr13I
    >< BsiZI
    >< BscBI
    >< Bme18I
    >< AvaII
    >< AsuI
    >< TfiI
    >< HinfI
    >< MnlI

    >< SduI
    >< NspII
    >< Bsp1286I
    >< BmyI
    >< MaeII
    >< AciI

```

FIGURE 13. 65

```

TCAAACCAAC GTAGTGCCCC CCGCATTACA TTTGGTGGAC CCACAGGATG AACTGCACAAT AACCGAATG
28150      28160      28170      28180      28190      28200      28210

      << HinPII >< StyI
      << HaeII
      > < Pali >< Hin6I >< EcoT14I
      > < HaeIII >< HhaI>< Ecol30I
      >< BspWI >< BssTII
      > < BsuRI >< Bsp143II
      >< HgaI> < BshI >< CfoI>< BsaJI >< HgaI
GAGGACGCAA TGGGGCAAGG CCAAAACAGC GCCGACCCCA AGGTTTACCC AATAATACTG CGTCTTGGTT
28220      28230      28240      28250      28260      28270      28280

      << TthHB8I
      > < ScrFI
      >< Pali
      >< PaeR7I
      >< NspIII
      > < MvaI
      >< HaeIII
      >< EcoRII
      >< Eco88I
      >< XhoI > < Ecl136I
      >< DsaV
      >< BsuRI
      >< SlaI > < BstOI
      >< MnlI>< TaqI> < BstNI
      >< CcrI > < BsiLI
      >< HinfI >< BshI
      >< TfiI>< BcoI>< BsaJI
      >< MnlI >< DdeI >< Aval > < ApyI
      >< AluI >< DdeI > < NlaIII >< BfrI >< Ama87I >< MnlI
CACAGCTCTC ACTCAGCATG GCAAGGAGGA ACTTAGATTC CCTCGAGGCC AGGGCGTTCC AATCAACACC
28290      28300      28310      28320      28330      28340      28350

      >< SinI
      >< Sau96I
      >< NspIV
      >< NspHII
      >< Eco47I
      >< Cfr13I
      >< Bsi2I
      >< Bme18I >< Ksp632I
      >< AvaII >< Eam1104I
      >< AsuI >< EarI > < AluI>< MboII >< MaeIII
AATAGTGGTC CAGATGACCA AATTGGCTAC TACCGAAGAG CTACCCGACG AGTTTCGTGGT GGTGACGGCA
28360      28370      28380      28390      28400      28410      28420

      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< BmyI
      >< BanII
      >< SstI
      >< SduI
      >< SacI
      >< NspII
      >< HgiAI
      >< EspI
      >< Eco24I
      >< Ecl136II
      >< DdeI
      >< CelII
      >< Bsp1286I
      >< Bpu1102I
      >< B
```

FIGURE 13.66

```

    >> Alw21I    >> Csp6I    >> BlnI    >> BshI>> HindIII
>> HphI >> AluI    >> AfaI    >> AvrII    >> AsuI    >> AluI
AAATGAAAGA GCTCAGCCCC AGATGGTACT TCTATTACCT AGGAACTGGC CCAGAAGCTT CACTTCCCTA
    28430      28440      28450      28460      28470      28480      28490

>> HinPII
>> Hin6I
>> HhaI
>> HaeII
>> CfoI
>> Bsp143II    >> MnlI    >> NlaIV
>> SfaNI >> DdeI >> BscBI
CGGCGCTAAC AAAGAAGGCA TCGTATGGGT TGCAACTGAG GGAGCCTTGA ATACACCCAA AGACCACATT
    28500      28510      28520      28530      28540      28550      28560

>> NlaIV
>> Eco64I
>> BscBI
>> Bani
>> AciI
>> AccBII >> BbvI    >> Fnu4HI    >> MnlI
GGCACCCGCA ATCCTAATAA CAATGCTGCC ACCGTGCTAC AACTTCCTCA AGGAACAACA TTGCCAAAAG
    28570      28580      28590      28600      28610      28620      28630

>> Thal
>> MnlI
>> MaeII >> MvnI
>> MnlI    BstUI >>
>> Fnu4HI    >> Ksp632I    Bsp50I >>
>> BspWI    >> EarI    >> BsaAI>> AciI
>> MnlI >> MnlI    >> AciI>> MboII    >> Eam1104I    AccII >>
GCTTCTACGC AGAGGGAAGC AGAGGCGGCA GTCAAGCCTC TTCTCGCTCC TCATCACGTA GTCGCGGTAA
    28640      28650      28660      28670      28680      28690      28700

>> ScrFI
>> MvaI
>> EcoRII
>> Ecl136I
>> DsaV>> Fnu4HI
>> BstOI
>> BstNI
>> BsiLI
>> ApyI
>> BbvI    >> TaqI    >> AciI
TTCAAGAAAT TCAACTCCTG GCAGCAGTAG GGGAAATTCT CCTGCTCGAA TGGCTAGCGG AGGTGGTGAA
    28710      28720      28730      28740      28750      28760      28770

>> Thal
>> MvnI
>> HphI >> MnlI
>> HinPII
>> Hin6I
>> HhaI
>> BstUI    >> RmaI
>> Bsp50I    >> MaeI
>> BbvI >> CfoI>> Fnu4HI
>> AccII>> BspWI    >> AluI
ACTGCCCTCG CGCTATTGCT GCTAGACAGA TTGAACCAGC TTGAGAGCAA AGTTTCTGGT AAAGGCCAAC
    28780      28790      28800      28810      28820      28830      28840

>> PalI>> MaeIII
>> HaeIII
>> BsuRI    >> DdeI
>> Fnu4HI
>> DdeI
>> RsaI >>
>> MnlI
>> MaeII >>
>> Csp6I >>

```

FIGURE 13.67

```

> < BshI > < BbvI >< MnlI >< BspWI >< SfaNI > AfaI ><
AACAAACAAGG CCAAACTGTC ACTAAGAAAT CTGCTGCTGA GGCATCTAAA AAGCCTCGCC AAAAACGTAC
28850 28860 28870 28880 28890 28900 28910

>< Tth111I
>< SinI
>< Sau96I
>< NspIV
>< NspHII
> < MaeII
>< Eco47I
>< Cfr13I
>< BsmBI
>< RsaI >< BsiZI >< StyI
>< MaeIII >< Bme18I >< EcoT14I
>< MaeII >< Esp3I >< AvaII >< Eco130I
>< Csp6I >< BsmAI >< AsuI >< BssT1I
>< AfaI >< Alw26I> < AspI >< BsaJI
TGCCACAAAA CAGTACAACG TCACTCAAGC ATTGGGGAGA CGTGGTCCAG AACAAACCCA AGGAAATTTC
28920 28930 28940 28950 28960 28970 28980

>< SinI
>< Sau96I
>< NspIV
>< NspHII
>< NlaIV
>< Eco47I
>< Cfr13I
>< BsiZI
>< BscBI
>< Bme18I
>< AvaII
>< AsuI
>< Pali
>< HaeIII
>< GdiII
>< Fnu4HI
>< EaeI
>< BsuRI
>< BshI
>< AciI
BspWI >
GGGGACCAAG ACCTAATCAG ACAAGGAAC TATTACAAAC ATTGGCCGCA AATTGCACAA TTTGCTCCAA
28990 29000 29010 29020 29030 29040 29050

>< BsmI
>< BscCI >< MnlI >< MaeIII >< MaeIII >< NlaIII
GTGCCTCTGC ATTCTTTGGA ATGTCACGCA TTGGCATGGA AGTCACACCT TCGGGAACAT GGCTGACTTA
29060 29070 29080 29090 29100 29110 29120

>< XhoII
>< Sau3AI
>< NdeII
>< MflI
>< MboI
>< FokI
>< Tru9I
>< NlaIV
>< NlaIII
>< MseI
>< BscBI >< BstXI>< AlwI> < Bsp143I >< AspI >< BspWI ><
TCATGGAGCC ATTAAATTGG ATGACAAAGA TCCACAATTC AAAGACAACG TCATACTGCT GAACAAGCAC
29130 29140 29150 29160 29170 29180 29190

>< HgaI
ATTGACGCAT ACAAACATT CCCACCAACA GAGCCTAAAA AGGACAAAAA GAAAAAGACT GATGAAGCTC
29200 29210 29220 29230 29240 29250 29260

EspI ><
DdeI ><
CeiII ><
Bpu1102I ><
AluI ><

```

FIGURE 13.68

```

                >< P1eI
                >< MboII
    >< Fnu4HI
    >< BspWI
    >< BsmAI
    >< Alw26I
    >< AciI
    AGCCTTTGCC GCAGAGACAA AAGAAGCAGC CCACTGTGAC TCTTCTTCCT GCGGCTGACA TGGATGATTT
    29270      29280      29290      29300      29310      29320      29330

                >< NlaIII
                >< HinfI
                NlaIII ><
    >< FokI
    CTCCAGACAA CTTCAAATTT CCATGAGTGG AGCTTCTGCT GATTCAACTC AGGCATAAAC ACTCATGATG
    29340      29350      29360      29370      29380      29390      29400

                >< MaeII
                >< AccI
    ACCACACAAG GCAGATGGGC TATGTAAACG TTTTCGCAAT TCCGTTTACG ATACATAGTC TACTCTTGTG
    29410      29420      29430      29440      29450      29460      29470

                >< Tru9I
                >< Tru9I
                >< MseI
                >< MseI
    >< XmnI
    >< EcoRI>< MaeIII
    >< Asp700I >< BsgI
    CAGAATGAAT TCTCGTAACT AAACAGCACA AGTAGGTTTA GTTAACTTTA ATCTCACATA GCAATCTTTA
    29480      29490      29500      29510      29520      29530      29540

                XorII >
                TthHB8I >
                TaqI >
                Sau3AI ><
                RsaI ><
                >< ThaIPvuI >
                NdeII ><
                >< MnlI
                >< MvnIMcrI >
                MboI ><
                DpnII ><
                DpnI ><
                Csp6I ><
                >< BstUI
                >< HaeIII BspCI >
                BspAI ><
                >< TthHB8I >< Bsp50I
                >< Pali Bsp143I ><
                >< BsuRI BsiEI >
                >< BshIAfaI ><

                >< MnlI
                >< MaeIII
    ATCAATGTGT AACATTAGGG AGGACTTGAA AGAGCCACCA CATTTCATC GAGGCCACGC GGAGTACCAT
    29550      29560      29570      29580      29590      29600      29610

                >< SduI
                >< NspII
                >< MboII >< VspI
                >< Ksp632I >< Eco24I >< Tru9I
                >< RsaI >< RmaI >< Fnu4HI >< Bsp1286I >< MseI
                >< Csp6I >< MaeI >< EarI >< BmyI >< AsnI
                >< AfaI >< BbvI >< AluI>< Eam1104I >< BanII >< AseI

```

FIGURE 13.69

```
CGAGGGTACA GTGAATAATG CTAGGGAGAG CTGCCTATAT GGAAGAGCCC TAATGTGTAA AATTAATTTT
29620      29630      29640      29650      29660      29670      29680

                >< Tru9I   >< DdeI
                >< MseI   >< BfrI
                >< NlaIII  > < AluI
AGTAGTGCTA TCCCCATGTG ATTTTAATAG CTTCTTAGGA GAATGACAAA AAAAAAAAAA AAAAAA
29690      29700      29710      29720      29730      29740
```

FIGURE 13. 70

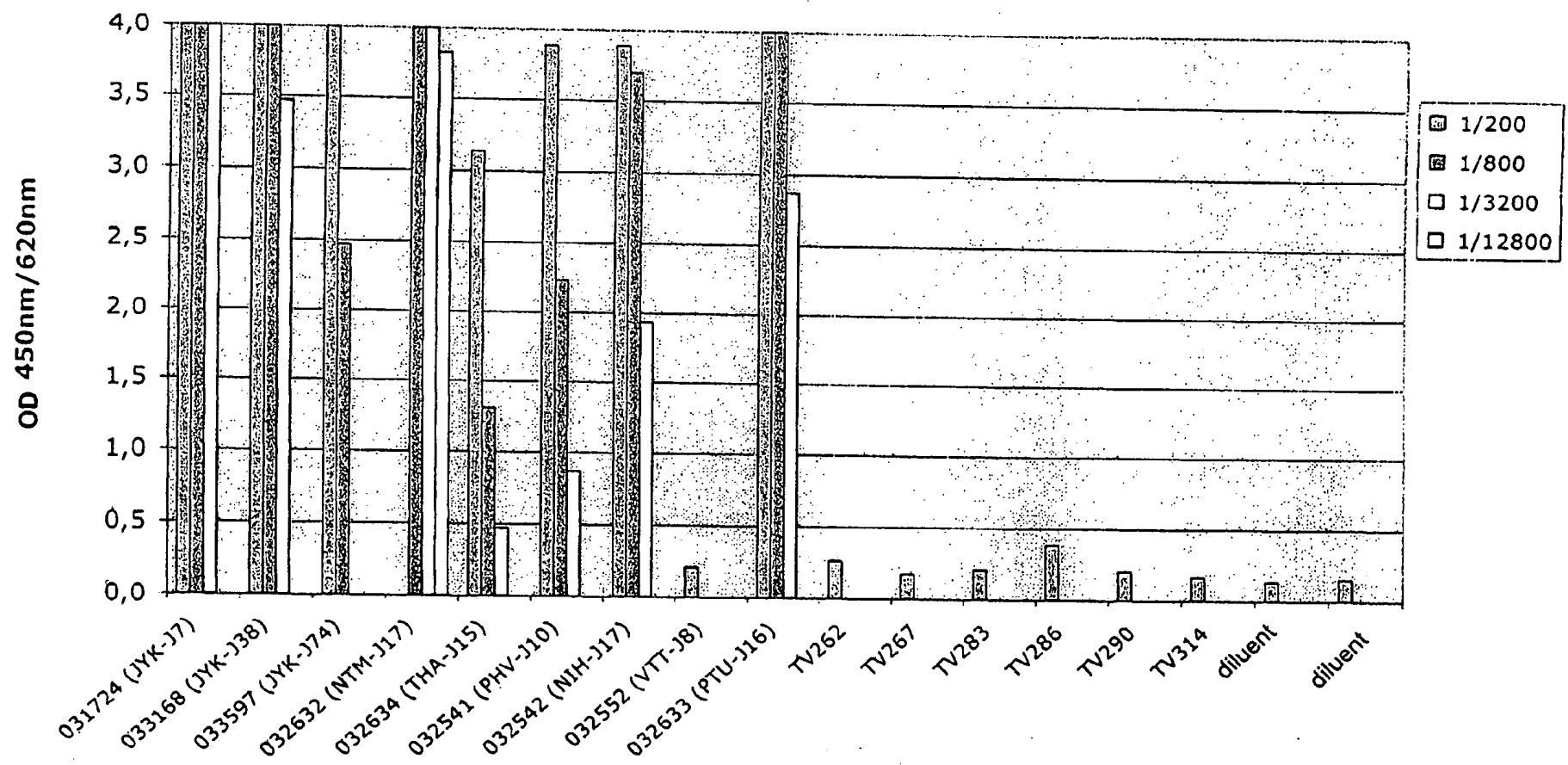


FIGURE 14

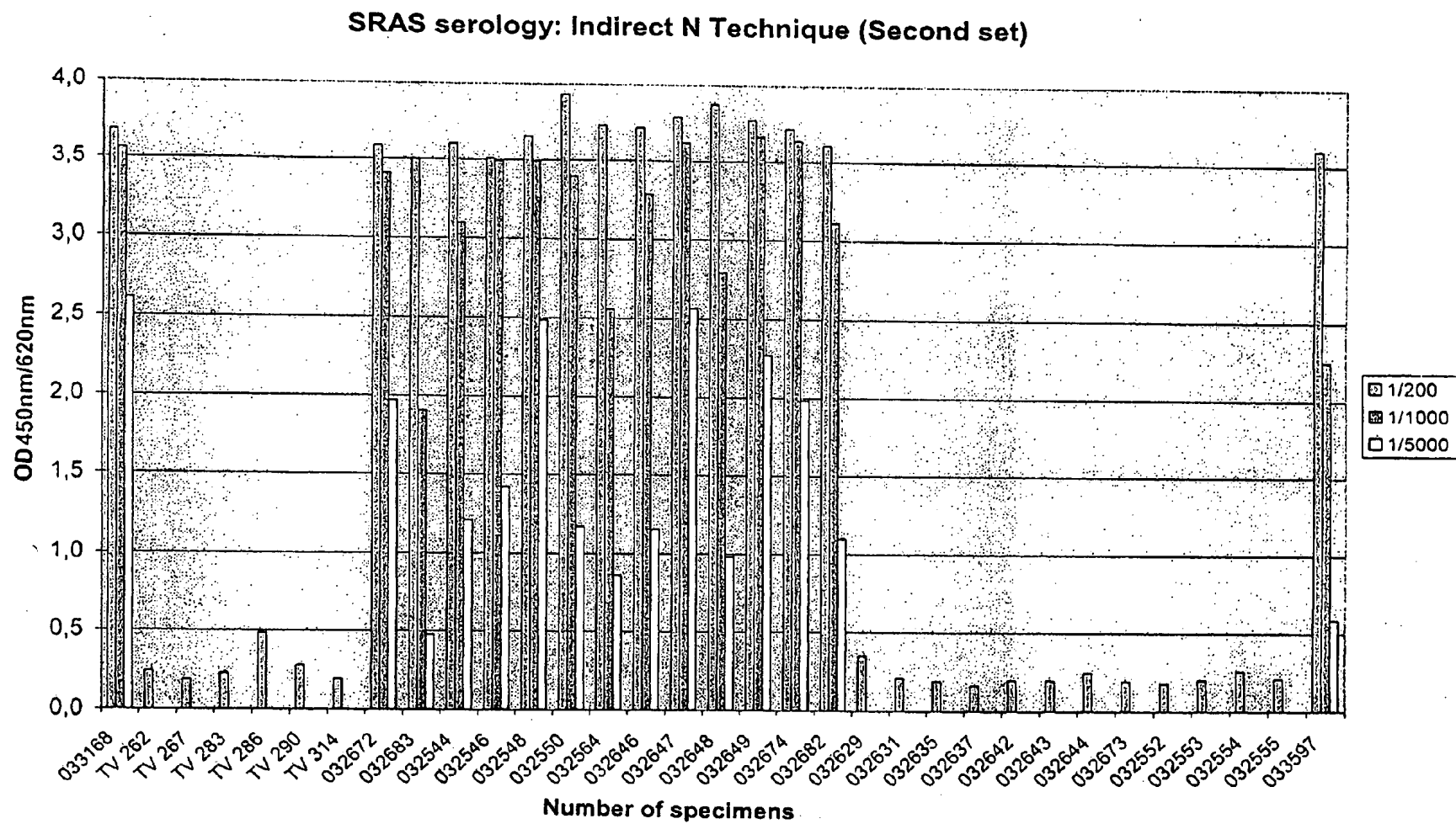


FIGURE 15

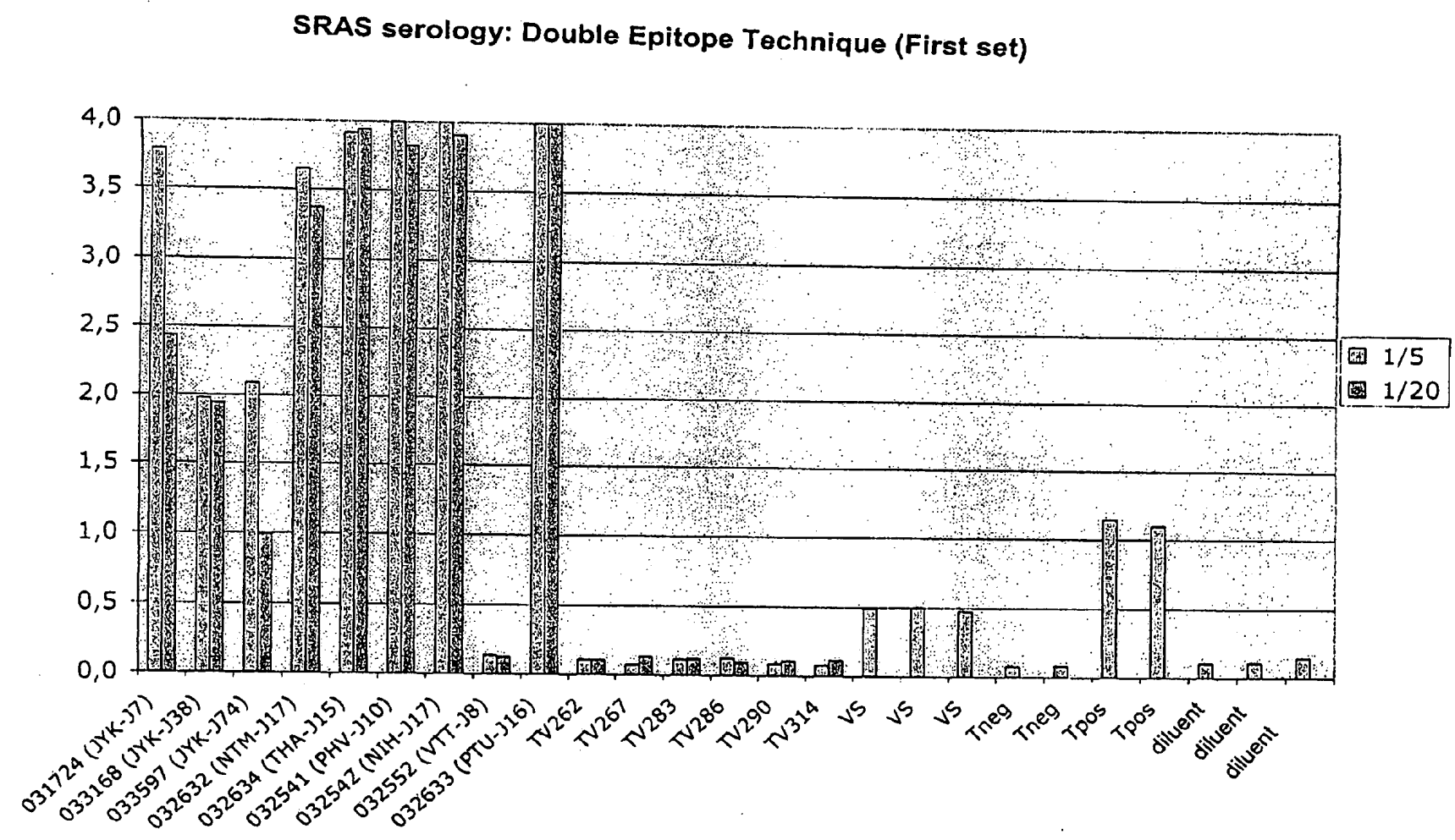


FIGURE 16

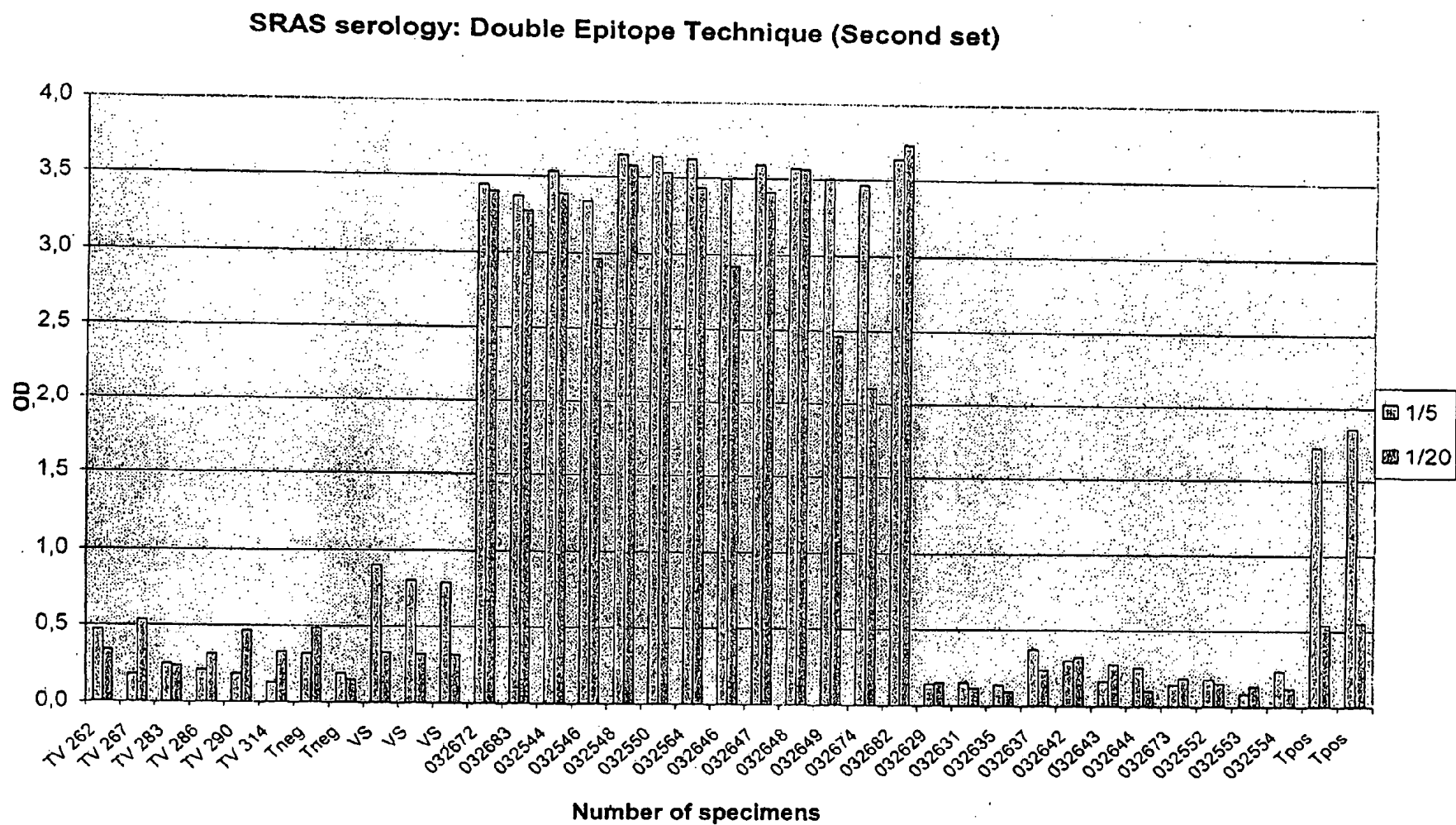


FIGURE 17

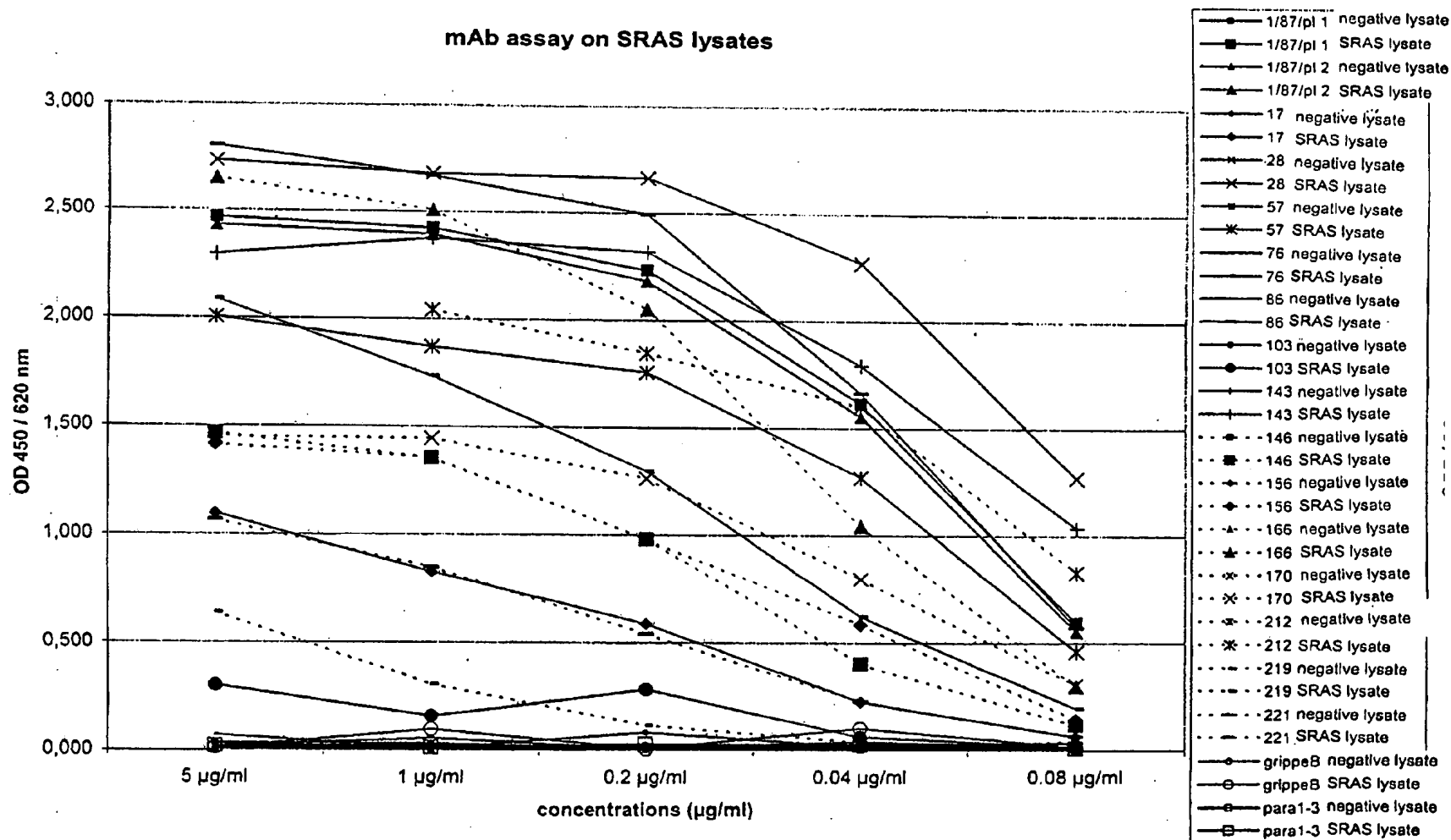
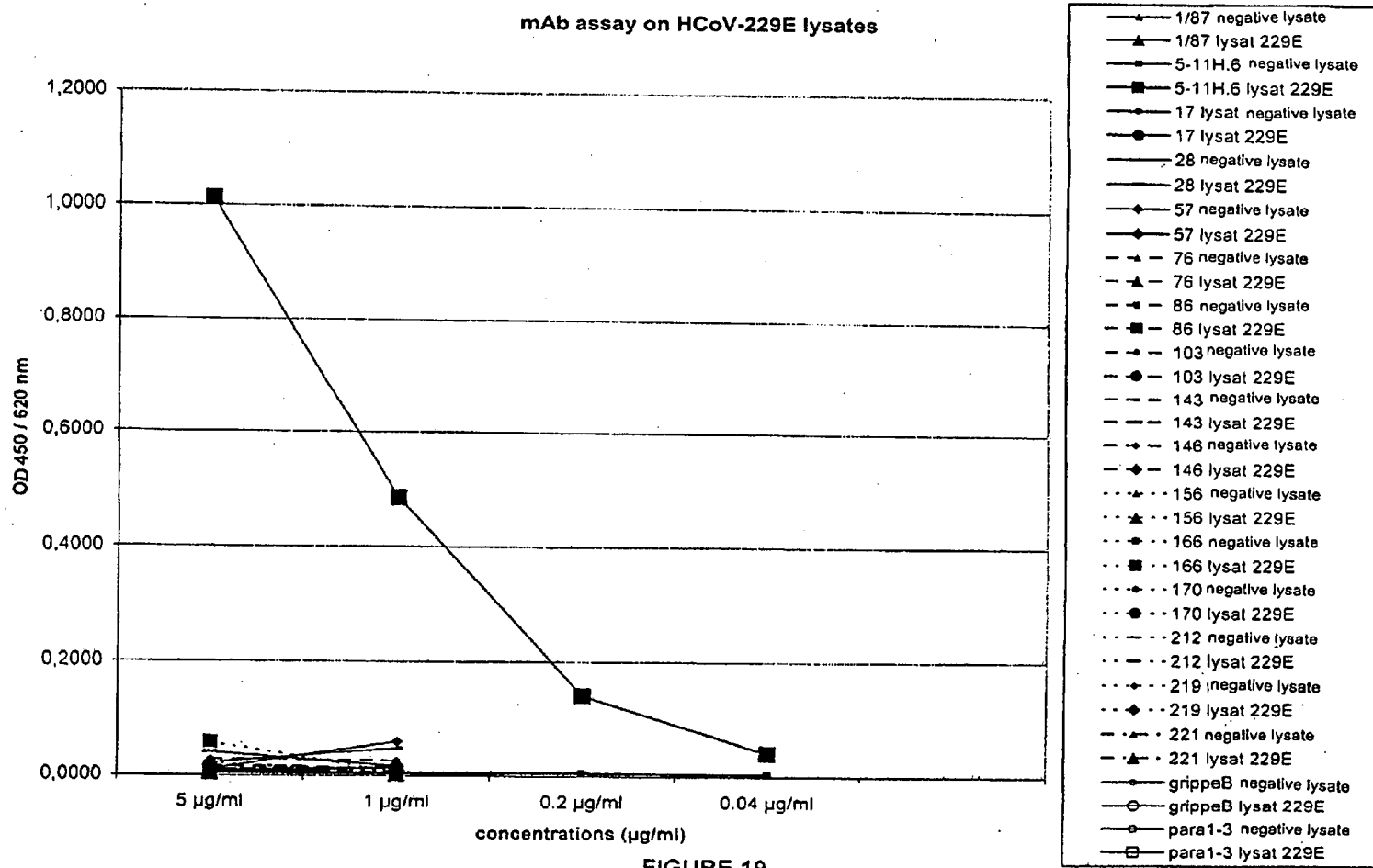


FIGURE 18



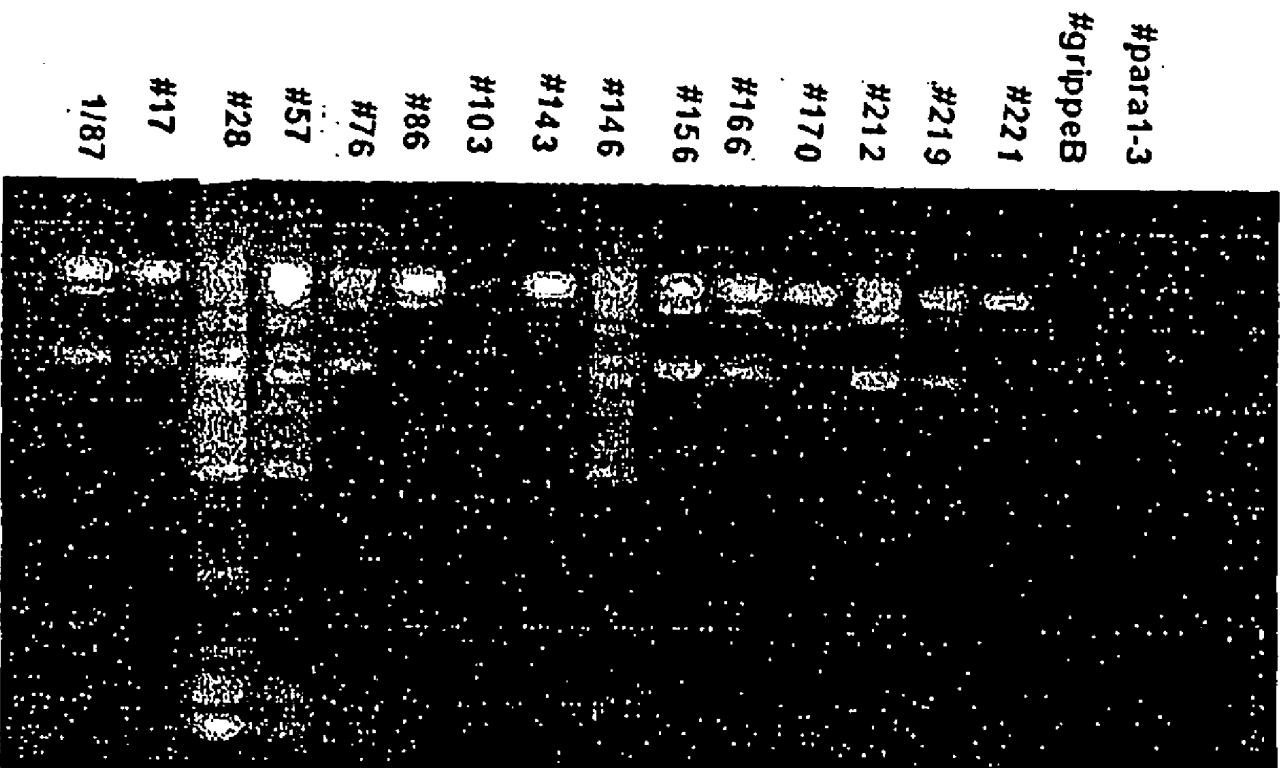


FIGURE 20

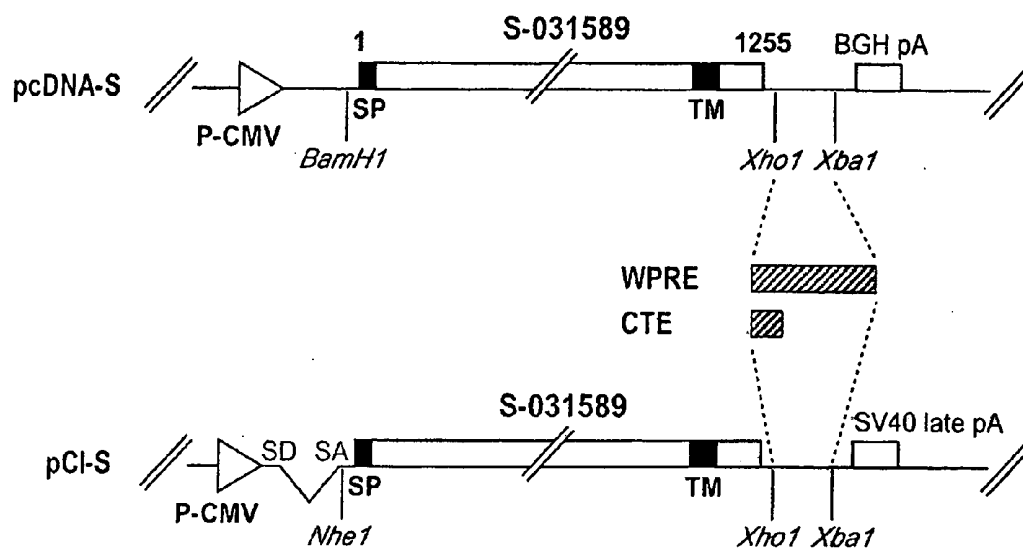


FIGURE 21

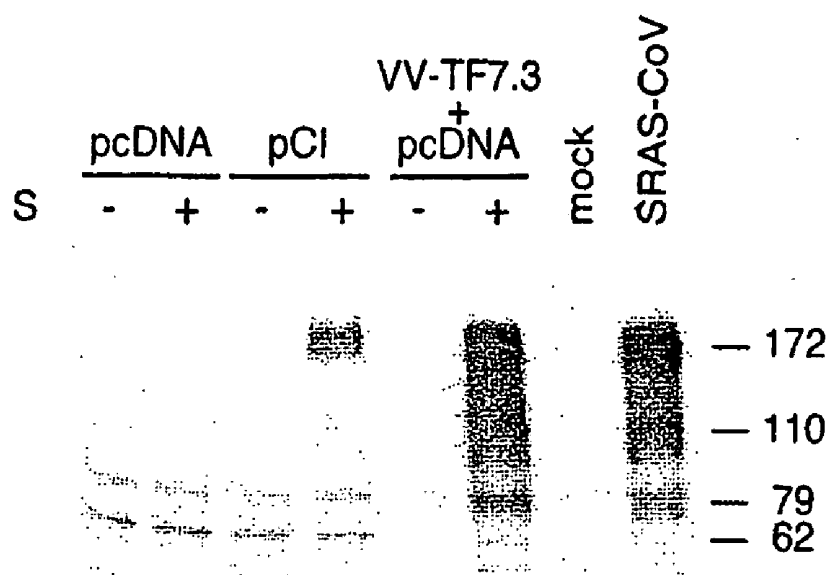
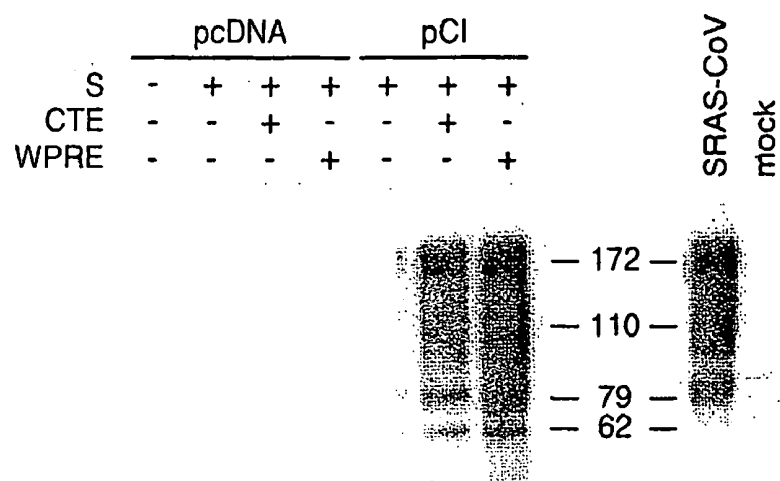


FIGURE 22

A.



B.

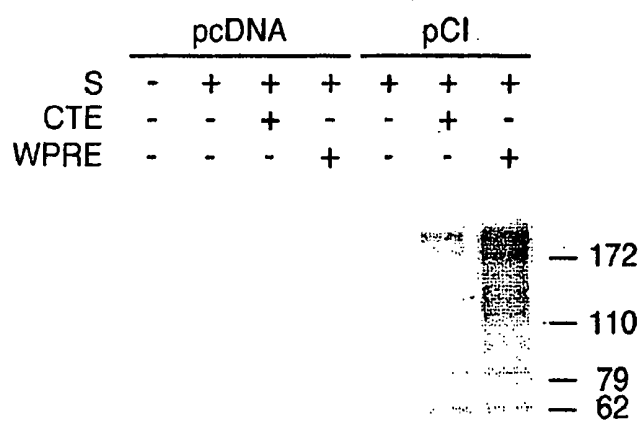


FIGURE 23

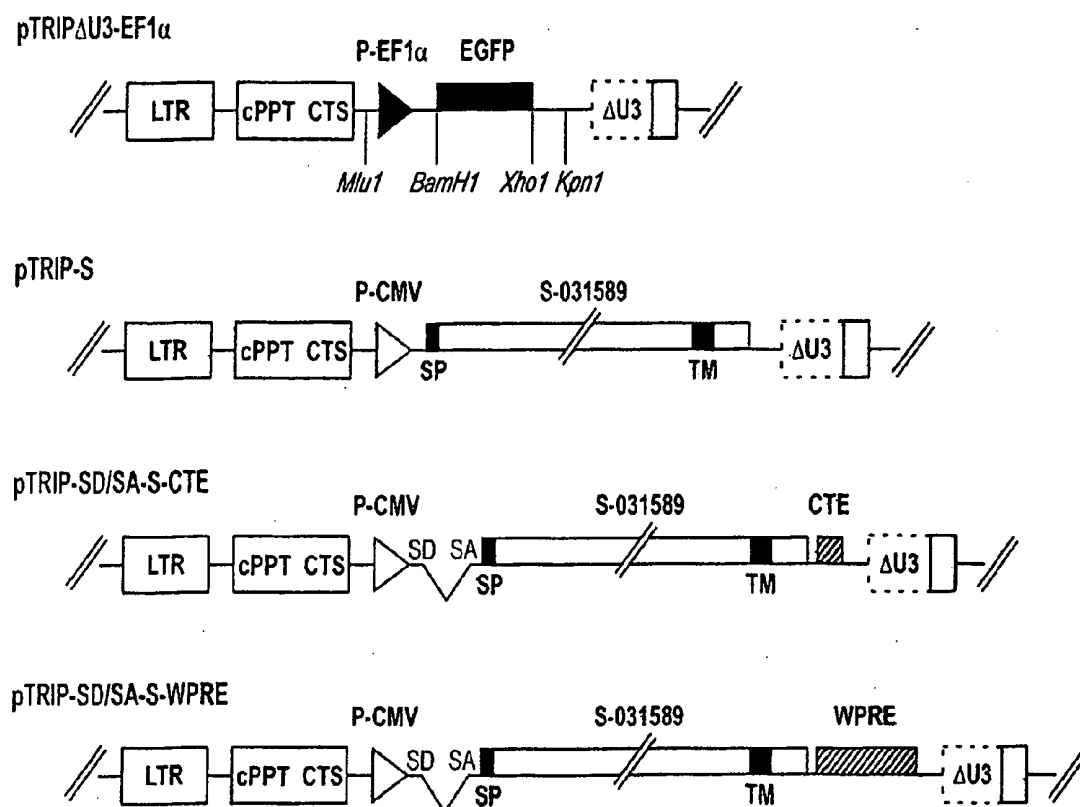


FIGURE 24

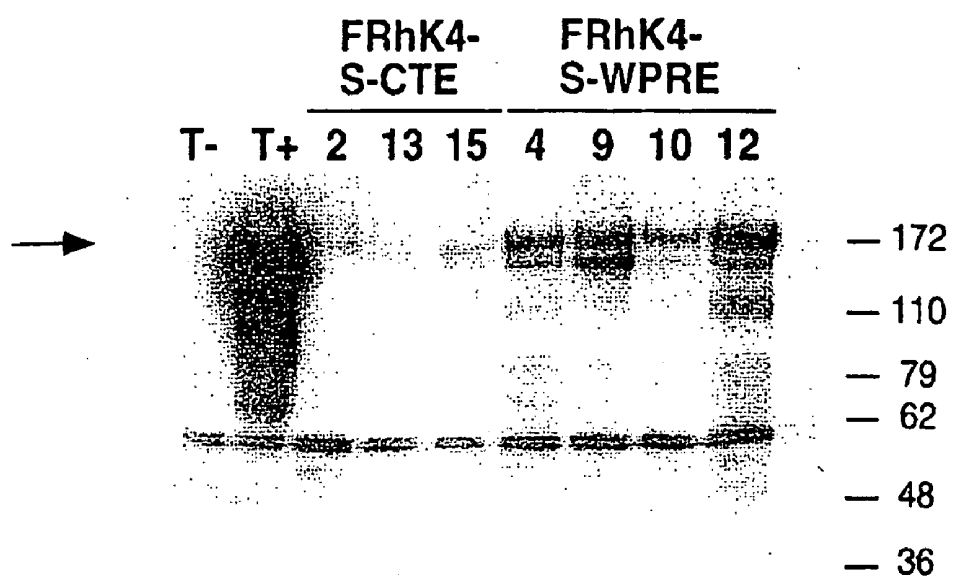


FIGURE 25

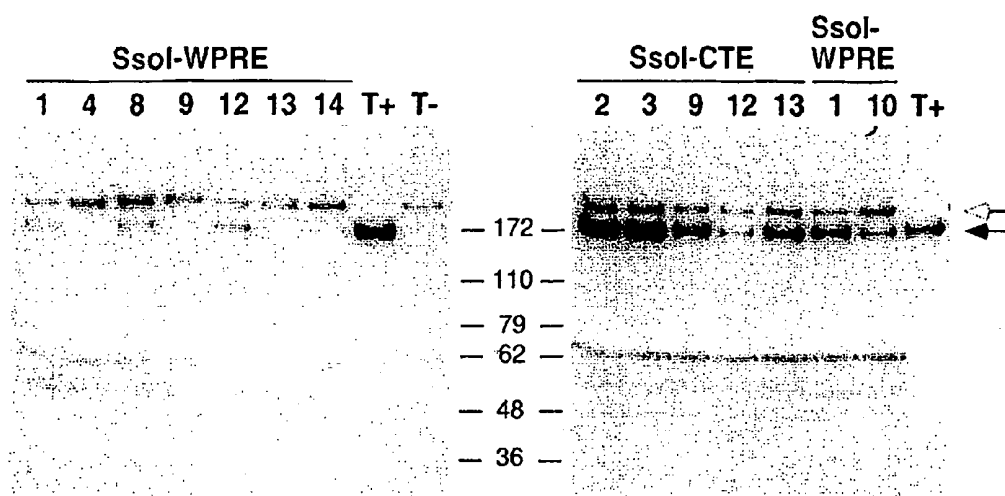
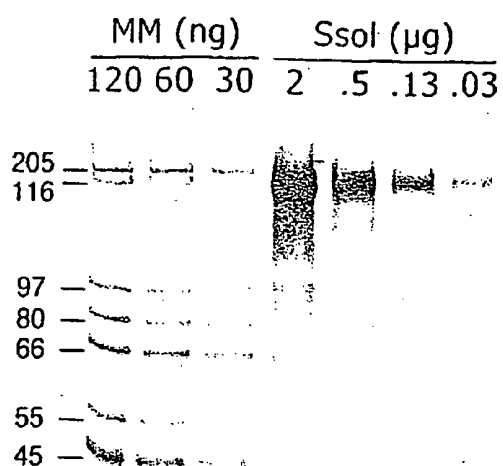
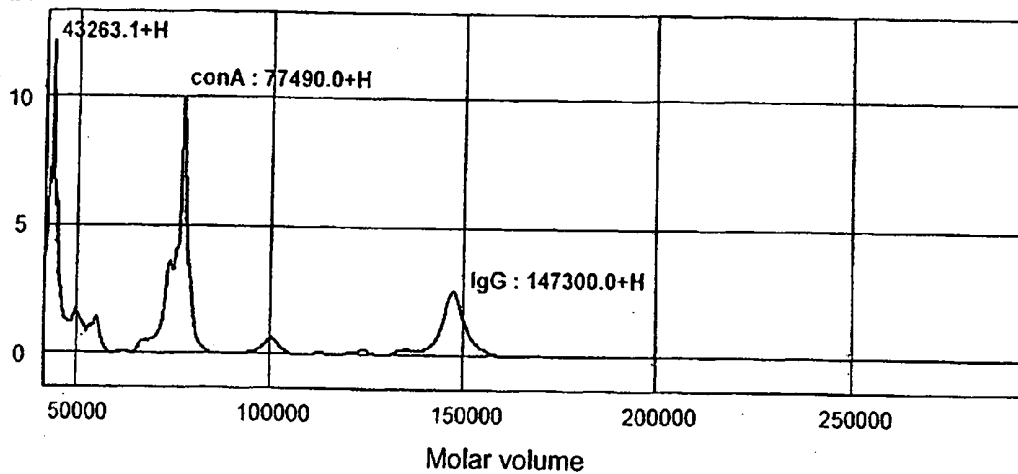


FIGURE 26

A.



B.



C.

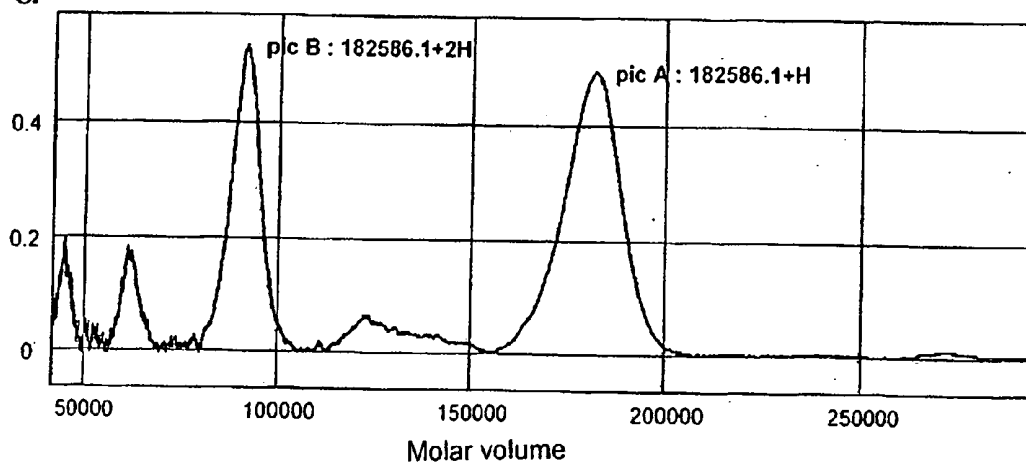


FIGURE 27 A-C

D.

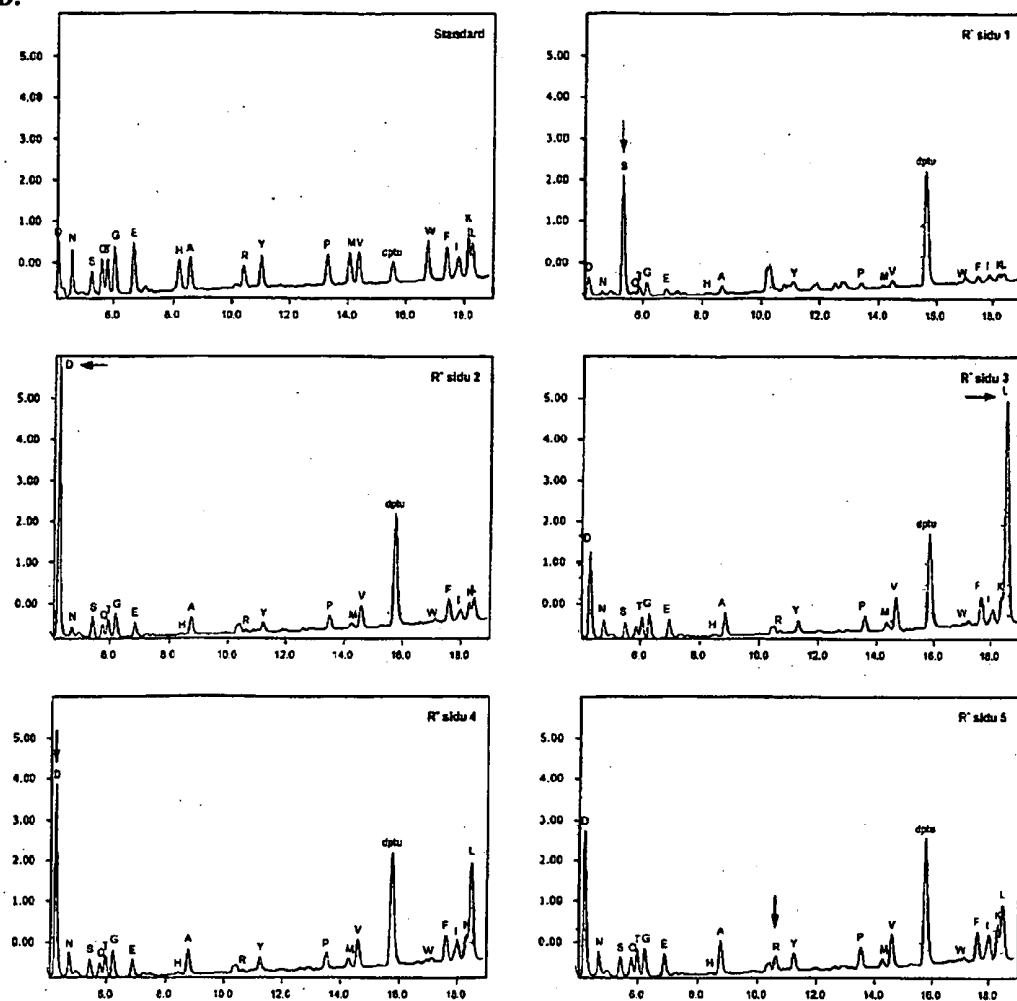
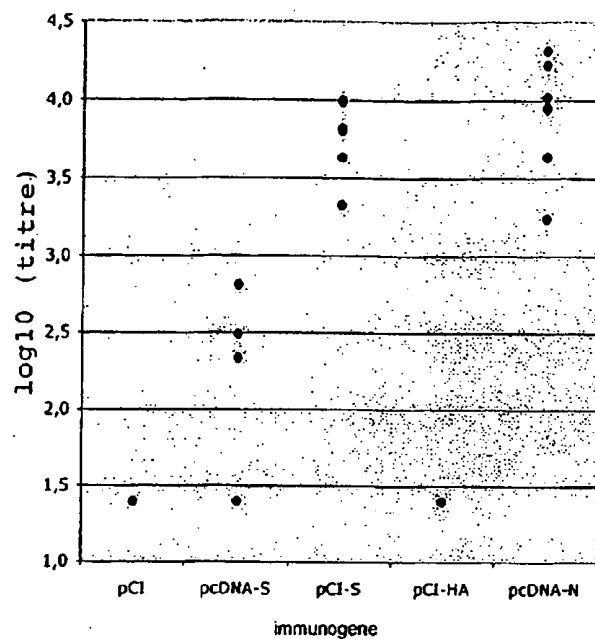


FIGURE 27 D

A.



B.

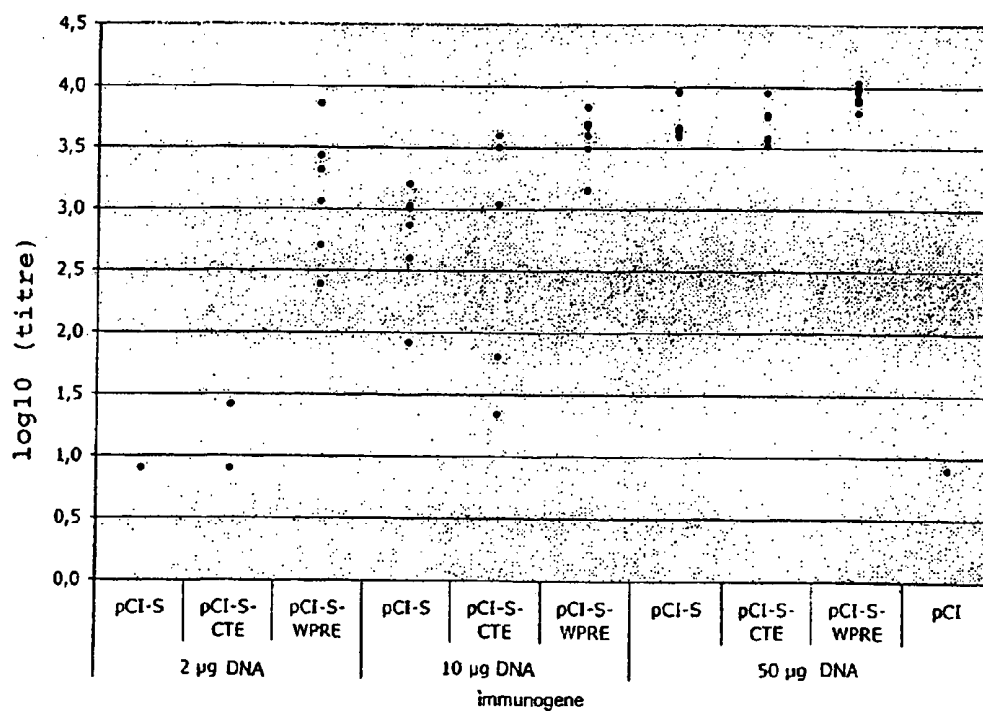


FIGURE 28

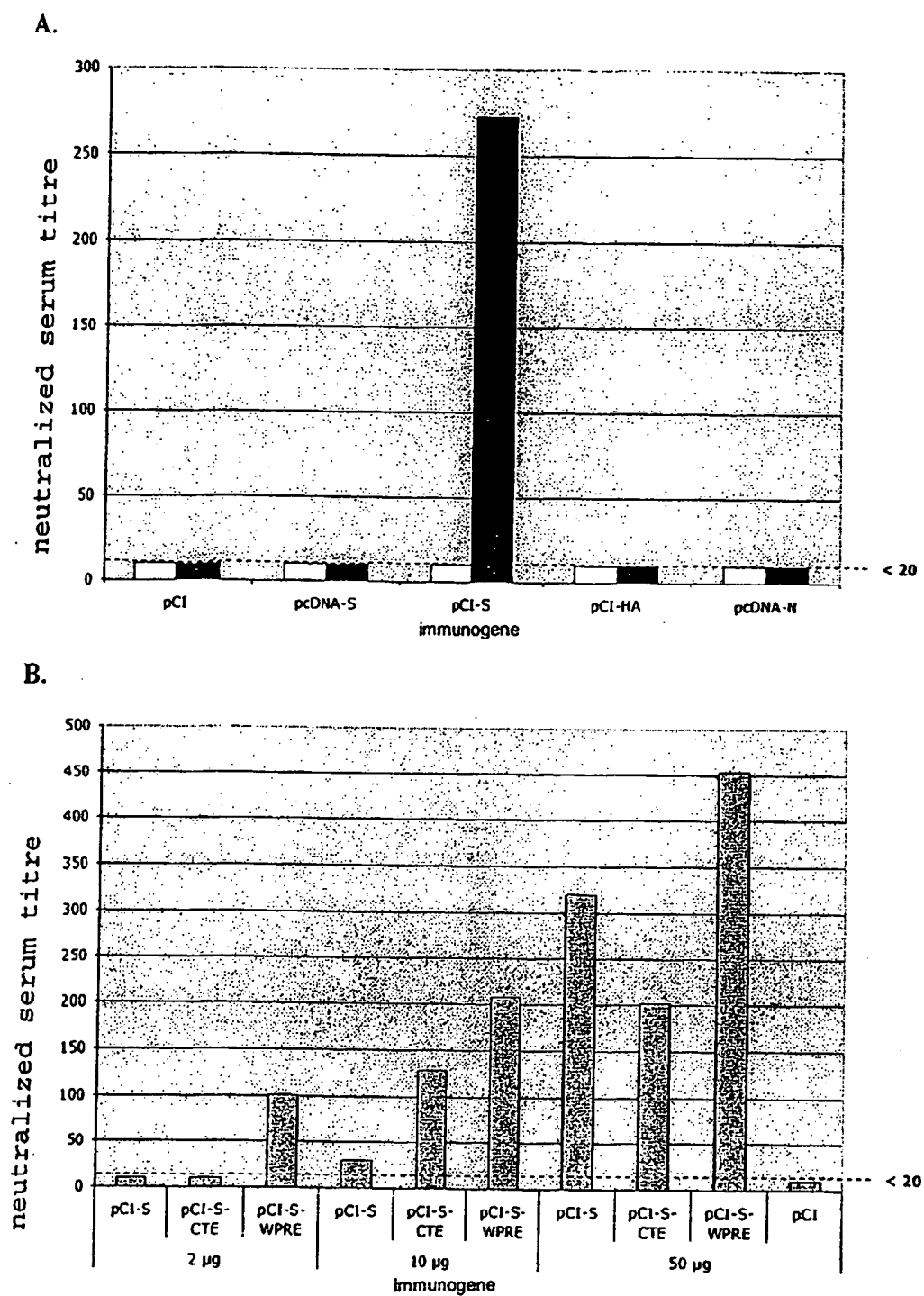


FIGURE 29

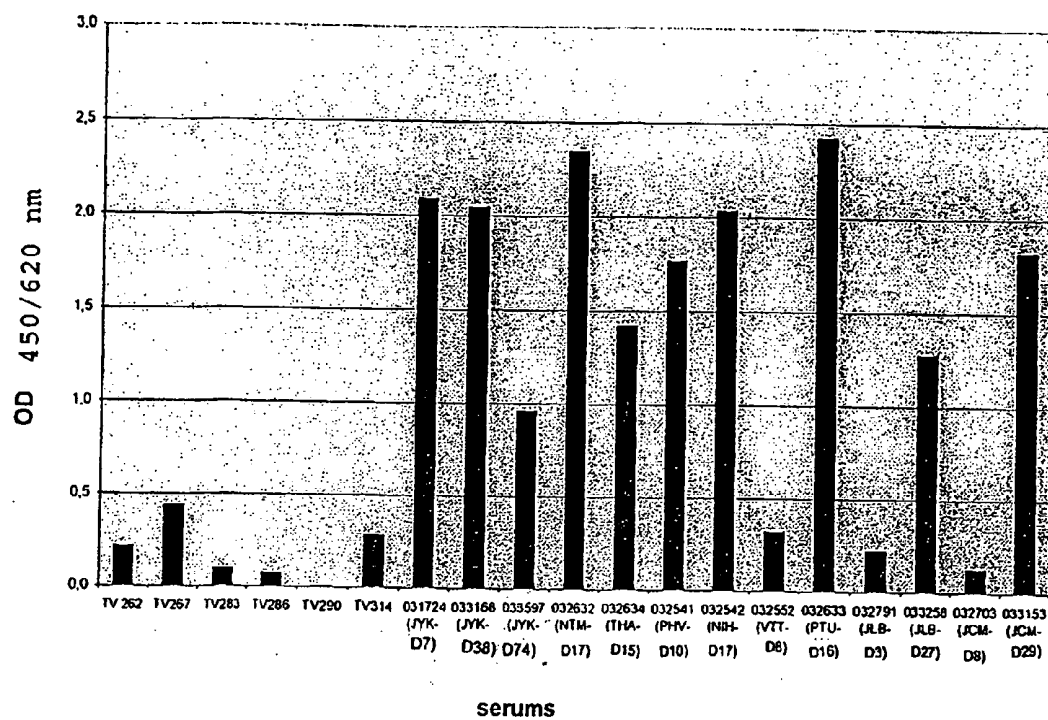


FIGURE 30

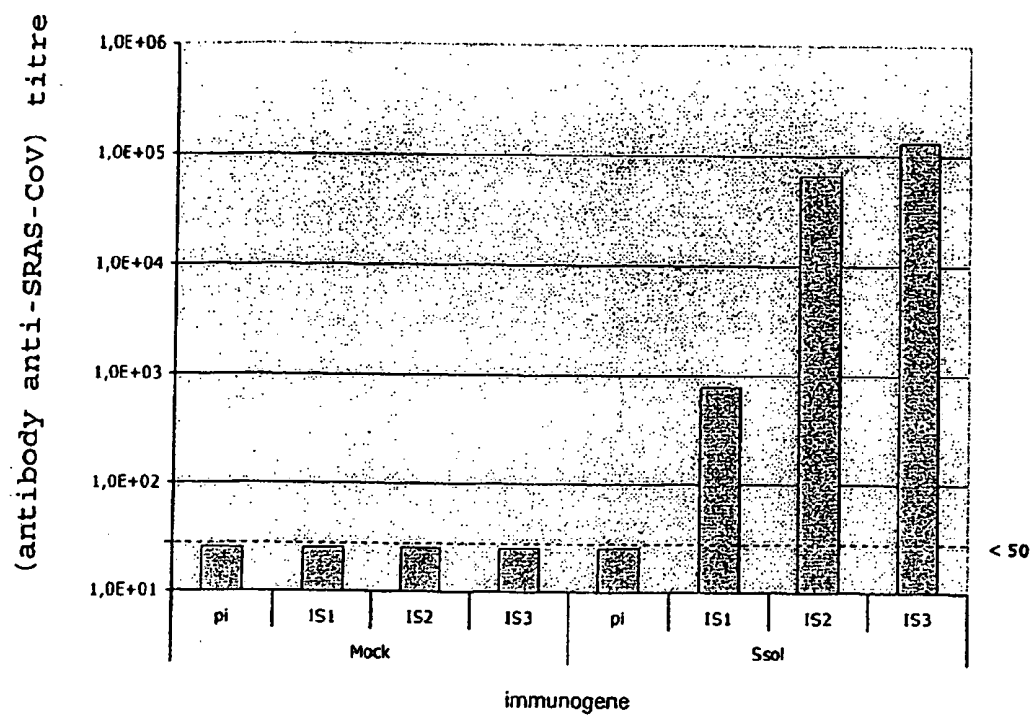


FIGURE 31

I-3059 S-040530	1	CTCTTCTGGAAAAAGGTAGGCTTTATCATTTAGAGAAAAACAACAGAGTTGTGGTTTCAAGTG
I-3059 S-040530	61 1	ATATTCTTGTTAACAACATAAACGAACATGTTTATTTCTTATTATTCTTACTCTCACTA GG" T " C " C " " " " " C " " C " " C " G C " G " C " " G " C " " G " C " "
I-3059 S-040530	121 44	GTGGTACTGACCTTGACCGGTGCACCACCTTTTGATGATGTTCAAGCTCCTAATTACACTC " C " " C " " C " " " " G " " " " " " " " " " " C " " C " " C " " C " " G " " G " " C " " C " " C " " " " " C " "
I-3059 S-040530	181 104	AACATACTT_CATCTATGAGGGGGSTTACTATCCTGATGAAATTTTATAGATCAGACACT " G " " C " " CAG " " G " _ " " C " " " " C " " G " " " " C " " C " " G " " C " " " C " GAGC " " " " C "
I-3059 S-040530	240 163	CTTTATTTAACCTCAGGATTTATTTCTTCCATTTTATTCTAATGTTACAGGGTTTCATACT " G " " CC " G " " C " " " " " CC " G " " C " " G " " C " " C " " CAGC " " C " " G " " C " " C " " C " " C " " C "
I-3059 S-040530	300 223	ATTAATCATACGTTTGCAACCCCTGTCATACCTTTTAAAGGATGGTATTATTATTTGCTGCC " C " " C " " C " " C " C " " " " " " " " " C " " G " " C " " C " " C " " " " " C " " C " " C " " C " " C " " " "
I-3059 S-040530	360 283	ACAGAGAAATCAAATGTTGTCGGTGGTGGGTTTTGGTTCTACCATGAACAACAAGTCA " C " " " " " " GAGC " " C " " G " " G " " G " " C " " " " " G " " C " " CAGC " " " " " " " " " " " " " " " " AGC "
I-3059 S-040530	420 343	CAGTCGGTGATTATTATTAAACAATTCTACTAATGTTGTTATACGAGCATGTAACCTTTGAA " " " AGC " " " " C " " C " " C " " " " " CAGC " " C " " C " " G " " G " " C " " G " " C " " C " " " " " C " " G "
I-3059 S-040530	480 403	TGTGTGACAACCCTTTCTTTGCTGTTTCTAAACCCATGGGTACACAGACACATACTATG C " " " C " " " " " " " " " " " C " " " " C " " G " " C " " " " " " " " " " " " C " " C " " " " " C " " C " " " "
I-3059 S-040530	540 463	ATATTCGATAATGCATTTAATTGCACTTTTCAGTACATATCTGATGCCTTTTCGCTTGAT " C " " " " " C " " C " " C " " C " " C " " " " C " " " " " " " " " " " CAGC " " C " " " " CAGC " " G " " C "
I-3059 S-040530	600 523	GTTTCAGAAAAGTCAGGTAATTTTAAACACTTACGAGAGTTTGTGTTTAAAAATAAAGAT " GAGC " " G " " " AGC " " C " " C " " C " " G " " C " " G " " G " " " " C " " " " " C " " G " " C " " G " " C "
I-3059 S-040530	660 583	GGGTTTCTCTATGTTTATAAGGGCTATCAACCTATAGATGTAGTTCGTGATCTACCTTCT " C " " C " " G " " C " " G " " C " " " " " " " C " " G " " C " " C " " C " " G " " GA " " A " " C " " G " " CAGC "
I-3059 S-040530	720 643	GGTTTTAACACTTTGAAACCTATTTTAAAGTTGCCCTCTTGGTATTAACATTACAAATTTT " C " " C " " " " " CC " " " " G " " C " " C " " C " " C " " " " C " " G " " C " " C " " " " " C " " C " " C " " C "
I-3059 S-040530	780 703	AGAGCCATTCTTACAGCCCTTTTCACTGCTCAAGACATTTGGGGCAGCTCAGCTGCAGCC C " G " " " " C " " G " " C " " " " " AGC " " " " C " " G " " " " C " " " " " " " CAGC " " C " " C " " "
I-3059 S-040530	840 763	TATTTGTTGGCTATTTAAAGCCAACTACATTTATGCTCAAGTATGATGAAAATGGTACA " C " " C " " G " " " " CC " G " " " " T " C " " C " " C " " " " G " " " " C " " C " " G " " C " " C " " C "
I-3059 S-040530	900 823	ATCACAGATGCTGTTGATTGTTCTCAAAATCCACTTGCTGAACTCAAATGCTCTGTTAAG " " " " C " " C " " C " " G " " C " " CAGC " " G " " C " " C " " G " " C " " G " " G " " G " " AGC " " G " " "
I-3059 S-040530	960 883	AGCTTTGAGATTGACAAAGGAATTTACCAGACCTCTAATTTTCAAGGTTGTTCCCTCAGGA " " " " C " " " " " C " " " " " G " " C " " C " " " " " " " " " " AGC " " C " " " " A " " G " " G " " TAGC " " C "
I-3059 S-040530	1020 943	GATGTTGTGAGATTCCTAATATTACAAACTTGTGTCCTTTTGGAGAGGTTTTTAAATGCT " " " " G " " " C " G " " " " " C " " " " " C " " C " " C " " " " C " " C " " C " " A " G " " C " " C " " C "
I-3059 S-040530	1080 1003	ACTAAATTCCTTCTGTCTATGCATGGGAGAGAAAAAAATTTCTAATTGTTTGTGCTGAT " C " " G " " " " CAGC " " G " " C " " C " " " " " C " G " " G " " G " " CAGC " " C " " C " " G " " C " " C "
I-3059 S-040530	1140 1063	TACTCTGTGCTCTACAACCTCAACATTTTTTCAACCTTTAAGTGTCTATGGGGTTTCTGCC " " AGC " " " " G " " " " " " C " " C " " C " " CAGC " " " " C " " " " " " " " " " " " " " " " " " GAGC " " "
I-3059 S-040530	1200 1123	ACTAAGTTGAATGATCTTTTGTCTCTCCAATGTCTATGCAGATTCTTTTGTAGTCAAGGGA " C " " C " " " C " " C " " C " " G " " " " " AG " " C " " G " " C " " C " " CAGC " " C " " G " " G " " " " C "
I-3059 S-040530	1260 1183	GATGATGTAAGACAAATAGCGCCAGGACAAACTGGTGTATTGCTGATTATAATTATAAA " C " " C " " G " " " " G " " C " " C " " " " C " G " " C " " C " " C " " C " " C " " C " " C " " C " " C " " "

FIGURE 32.1

[illegible]

FIGURE 32.2

I-3059	2697	GGTGGCTGGCGCTGCTCTTCAAATACCTTTTGTATGCAATGGCATATAGGTTCAATGGC
S-040530	2620	"A"C"A"C"C"G"G"C"C"C"C"G"G"C"C"C"C"
I-3059	2757	ATTGGAGTTACCCAAATGTTCTCTATGAGAACCAAAACAAATCGCCACCAATTAAAC
S-040530	2680	"C"C"G"G"G"C"G"G"C"G"G"G"G"G"G"G"G"G"G"G"C"
I-3059	2817	AAGGCGATTAGTCAAATTCAGAATCACTTACAACAACATCAACTGCATTGGGCAAGCTG
S-040530	2740	"C"C"C"G"C"G"GAGC"G"C"CAGC"C"CC"G"G"G"
I-3059	2877	CAAGACGTTGTTAACCAGAATGCTCAAGCATTAAACACACTTGTTAAACAACCTAGCTCT
S-040530	2800	"G"G"G"G"C"C"C"G"CC"G"C"G"G"G"G"G"AGC
I-3059	2937	AATTTTGGTGCAATTTCAAGTGTGCTAAATGATATCCTTTGCGACTTGATAAAGTCGAG
S-040530	2860	"C"C"C"C"CAGCTC"G"C"C"GAGCA"G"G"C"G"
I-3059	2997	GCGAGGTACAATTGACAGGCTAATTACAGGCAGACTTCAAAGCCTTCAAACCTATGTA
S-040530	2920	"C"A"G"G"C"C"G"C"AC"C"G"GTG"G"G"C"G"
I-3059	3057	ACACAACAATAATCAGGGCTGCTGAAATCAGGGCTTCTGCTAATCTTGCTGCTACTAAA
S-040530	2980	"C"G"G"G"A"C"C"G"C"ACG"C"G"C"C"
I-3059	3117	ATGCTCTGAGTGTGTTCTTGGACAATCAAAAAGAGTTGACTTTTGTGGAAGGGCTACCAC
S-040530	3040	"AGC"C"G"G"C"GAGC"G"G"G"C"C"C"CT"
I-3059	3177	CTTATGTCCTTCCACAAGCAGCCCCGCATGGTGTGCTTCTTACATGTCACGTATGTG
S-040530	3100	"G"AG"C"G"C"C"C"G"G"G"G"C"G"C"
I-3059	3237	CCATCCCAGGAGAGGAACCTCACCACAGCGCCAGCAATTTGTCATGAAGGCAAAGCATAC
S-040530	3160	"TAG"C"C"C"C"C"C"G"G"C"
I-3059	3297	TTCCCTCGTGAAGGTGTTTGTGTTAATGGCACTTCTTGTTTATTACACAGAGGAAC
S-040530	3220	"C"G"G"C"G"C"CAGC"C"C"C"C"
I-3059	3357	TTCTTTTCTCCACAAATAATTACTACAGACAATACATTTGTCTCAGGAAATTGTGATGTC
S-040530	3280	"CAGC"C"G"C"C"C"C"C"G"C"C"C"
I-3059	3417	GTTATTGGCATCATTAACAACACAGTTTATGATCCTCTGCAACCTGAGCTTGACTCATTC
S-040530	3340	"G"C"C"C"C"C"C"C"C"G"C"C"G"AGC"
I-3059	3477	AAAGAAGAGCTGGACAAGTACTTCAAAAATCATACATCACCAGATGTTGATCTTGGCGAC
S-040530	3400	"G"G"A"G"C"C"C"C"C"G"C"G"
I-3059	3537	ATTTAGGCATTAAAGCTTCTGTGCTCAACATTCAAAAAGAAATTGACCGCTCAATGAG
S-040530	3460	"CAGC"C"C"G"G"C"G"G"C"G"AA"CA"
I-3059	3597	GTCGCTAAAAATTAAATGAATCACTCATTGACCTTCAAGAATTGGGAAAATATGAGCAA
S-040530	3520	"G"C"G"CC"G"C"GAGC"G"C"G"GC"C"G"
I-3059	3657	TATATTAAATGGCCTTGGTATGTTGGCTCGGCTTCATTGCTGGACTAATTGCCATCGTC
S-040530	3580	"C"C"G"C"C"G"G"G"C"C"C"G"C"
I-3059	3717	ATGGTTACAATCTTGCTTGTGTCATGACTAGTTGTTGCAGTTGCCTCAAGGGTGCATGC
S-040530	3640	"G"C"C"G"C"C"C"C"C"TC"C"G"AA"C"
I-3059	3777	TCTTGTGGTTCTTGCTGCAAGTTTGATGAGGATGACTCTGAGCCAGTTCTCAAGGGTGTG
S-040530	3700	AGC"CAGC"C"C"C"AGC"C"G"G"C"
I-3059	3837	AAATTACATTACACATAAAGCACTTATGGATTGTTTATGAGATTTTACTCTTGGAT
S-040530	3760	"GC"G"C"C"CGA"
I-3059	3897	CAATTACTGCACAGCCAGTAAAAATTGACAATGCTTCTCCTGCAAGT
S-040530		

FIGURE 32.3

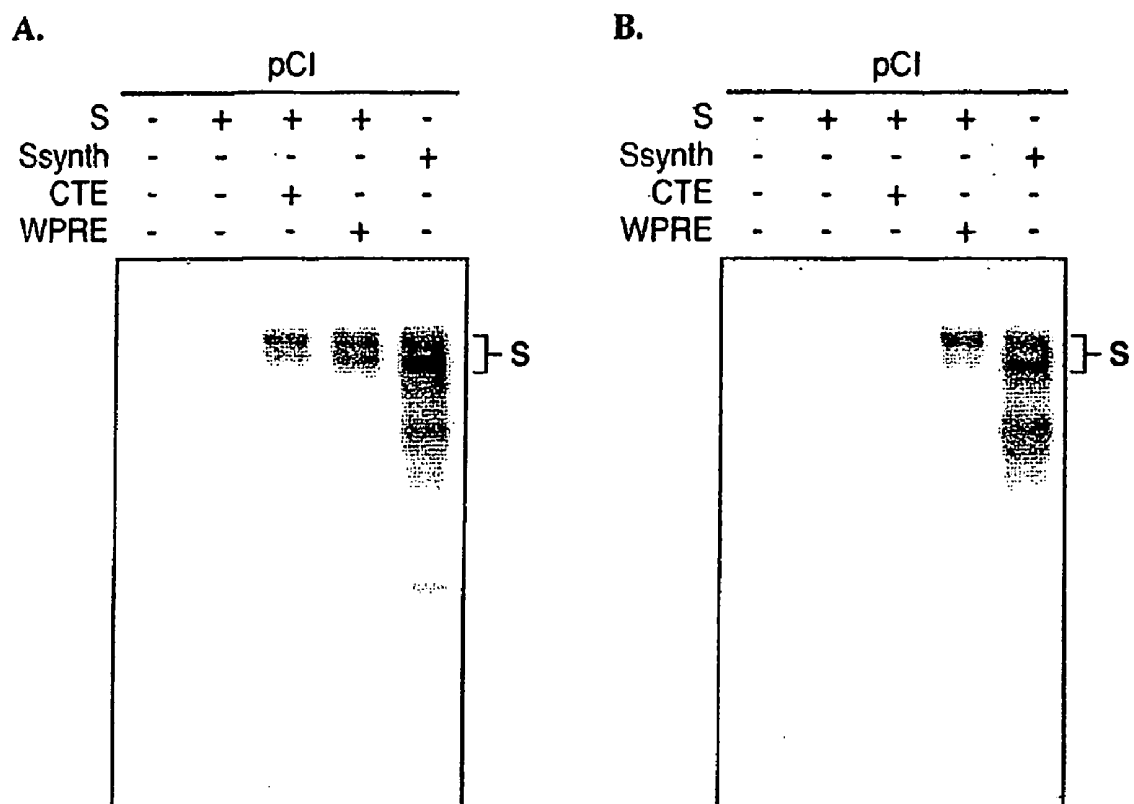
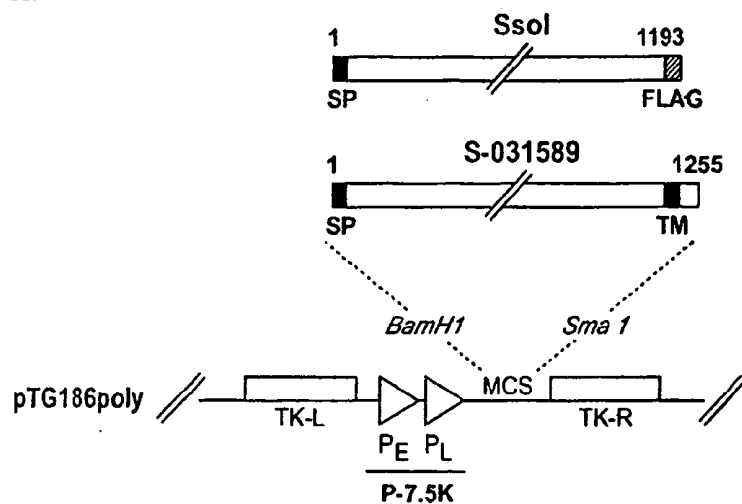
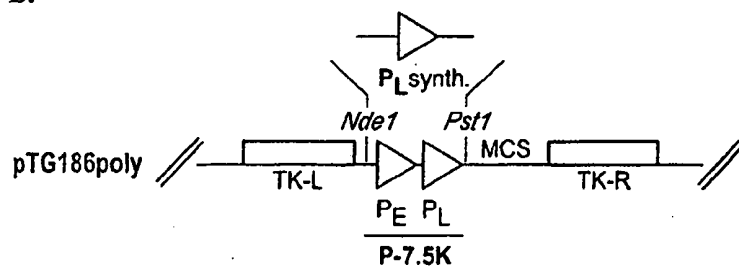


FIGURE 33

A.



B.



C.

$\frac{\text{CATATG}}{\text{NdeI}}$
 $\frac{\text{AGC}}{\text{promoteur 480}}$
 $\frac{[\text{T}]_{20}\text{GGCATATAAATA}}{\text{promoteur 480}}$
 $\frac{\text{GACTC}}{\text{Ascl}}$
 $\frac{\text{GGCGCGCC}}{\text{Ascl}}$
 $\frac{\text{AT}}{\text{PstI}}$
 $\frac{\text{CTGCAG}}{\text{PstI}}$

FIGURE 34 A-C

D.

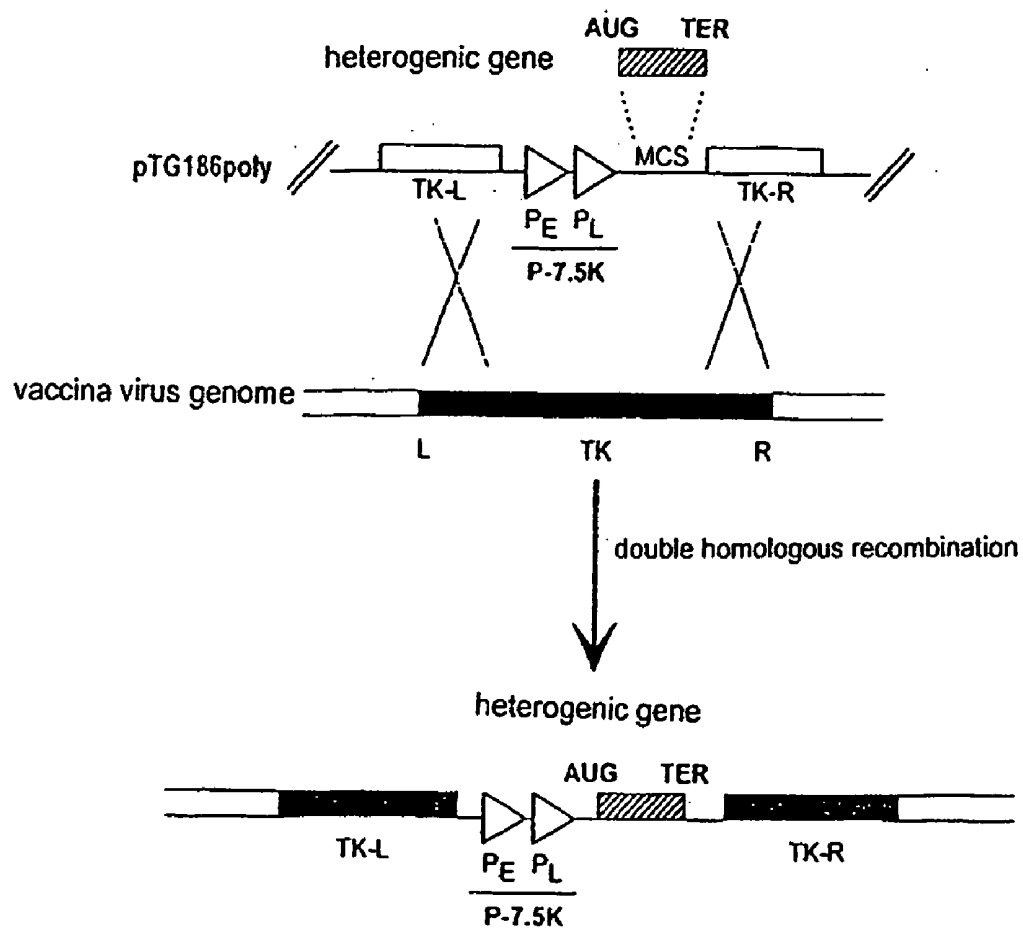


FIGURE 34 D

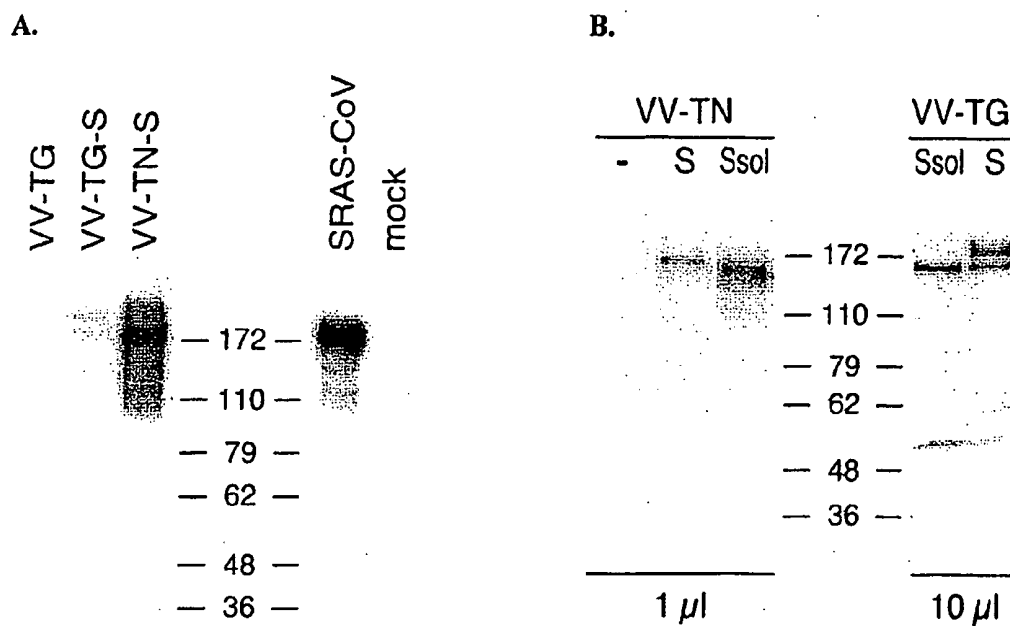
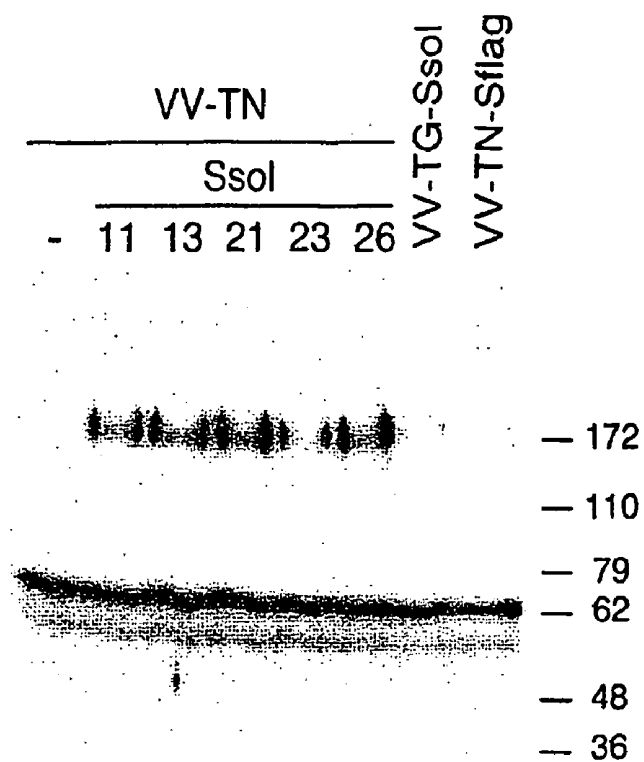


FIGURE 35

A.



B.

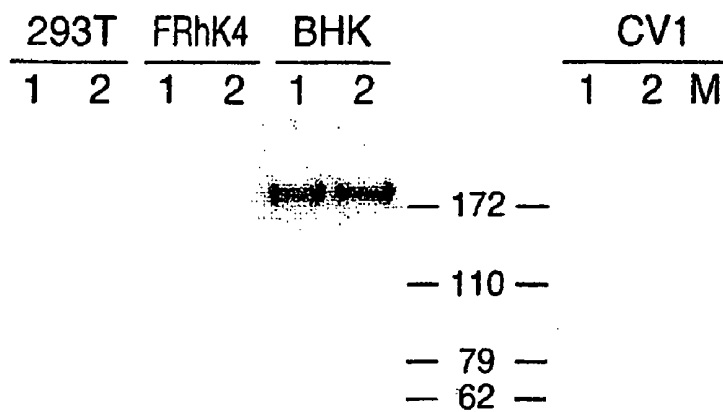


FIGURE 36



FIGURE 37

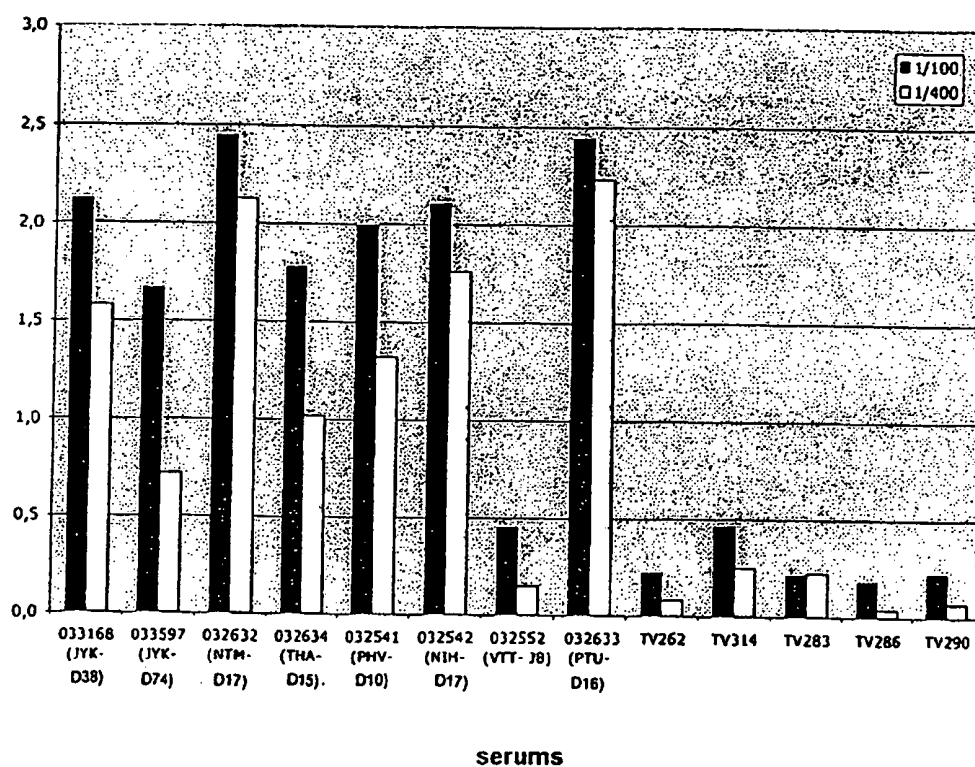
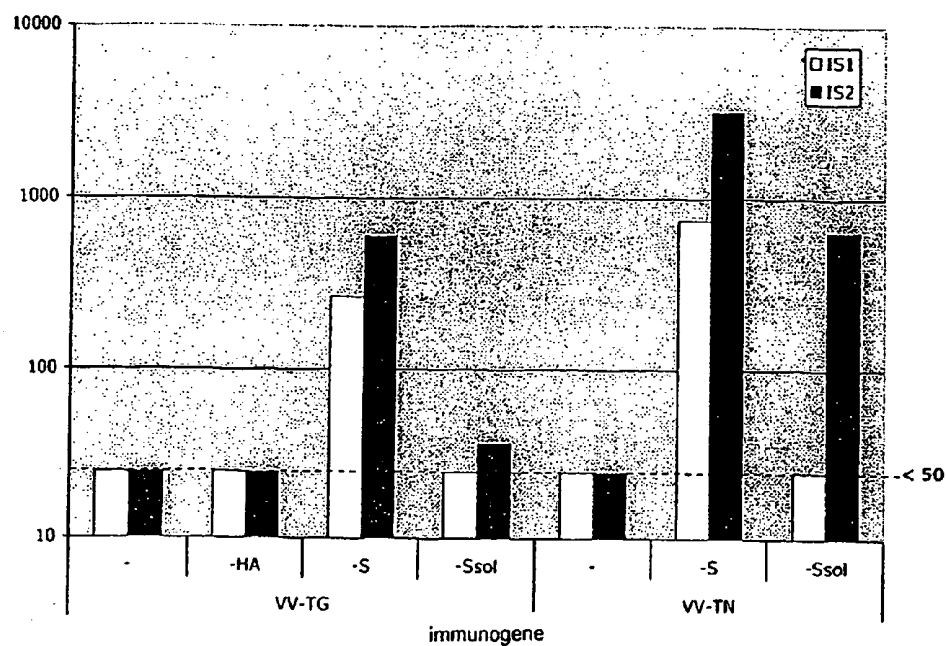


FIGURE 38

A.



B.

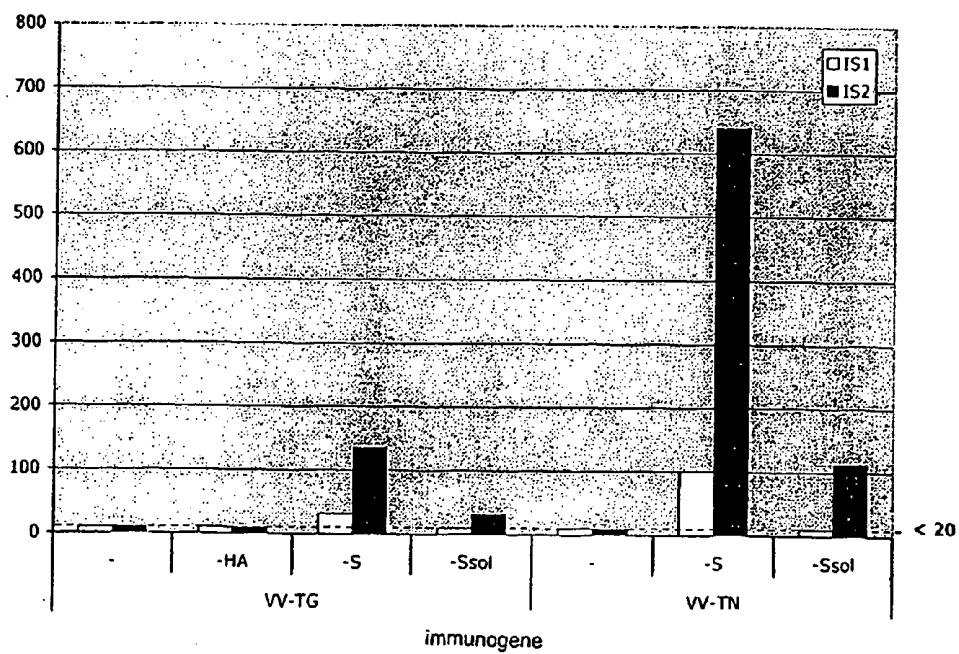


FIGURE 39

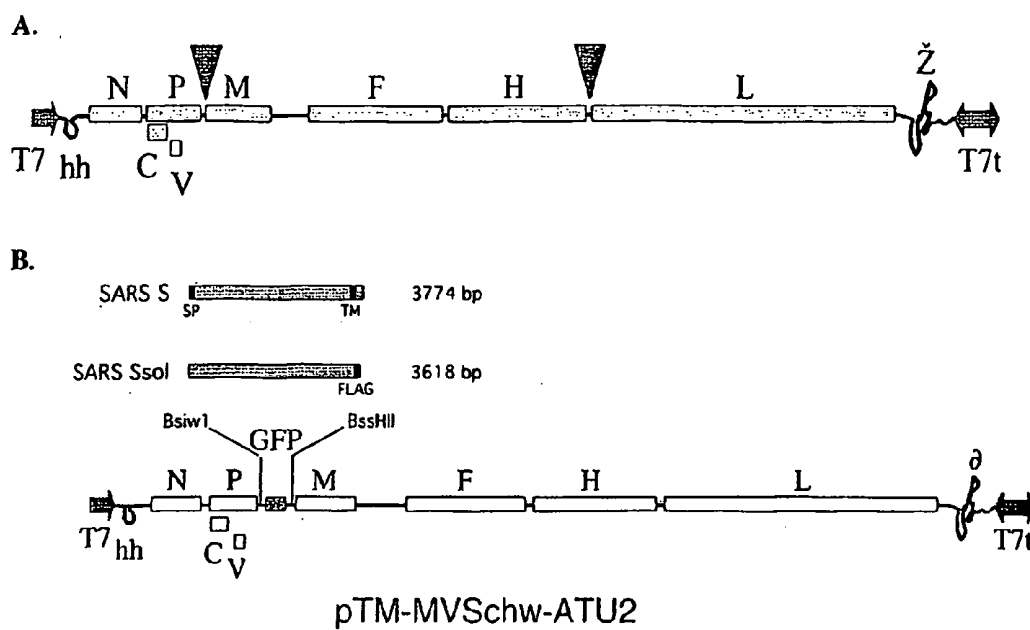


FIGURE 40

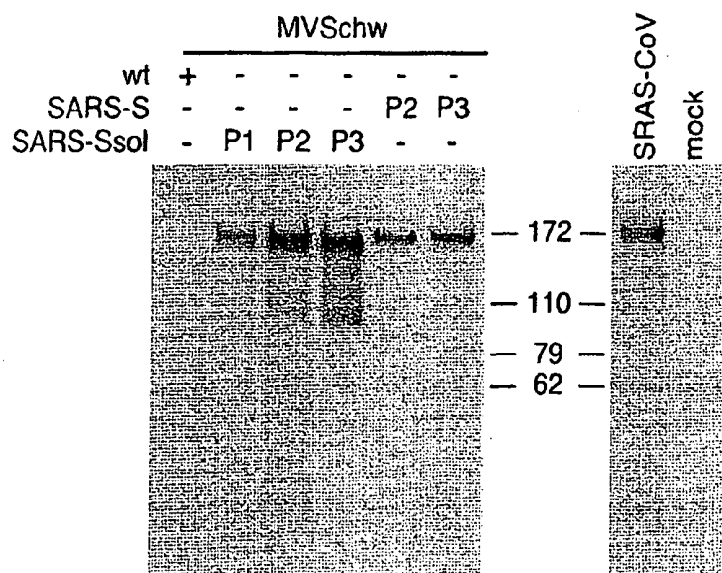


FIGURE 41

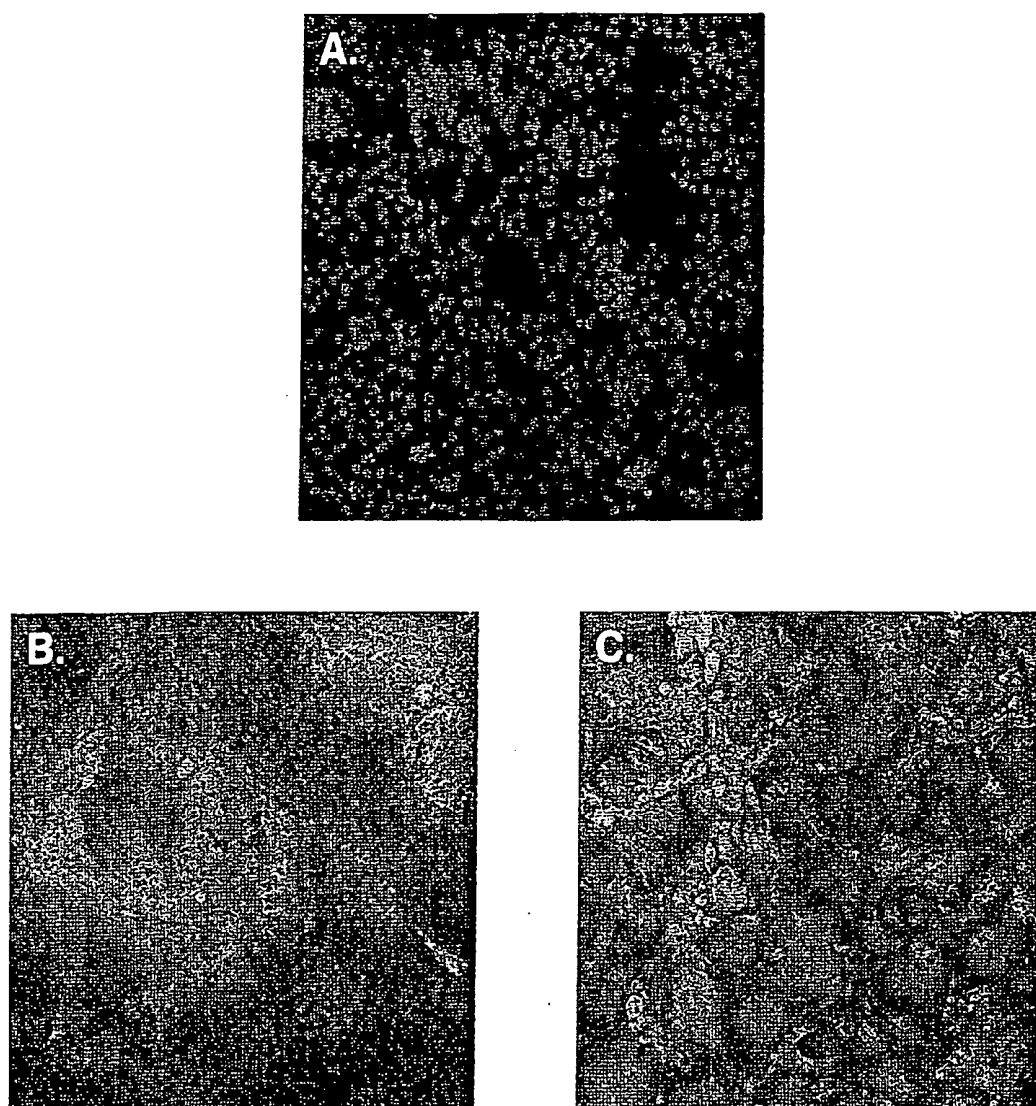


FIGURE 42

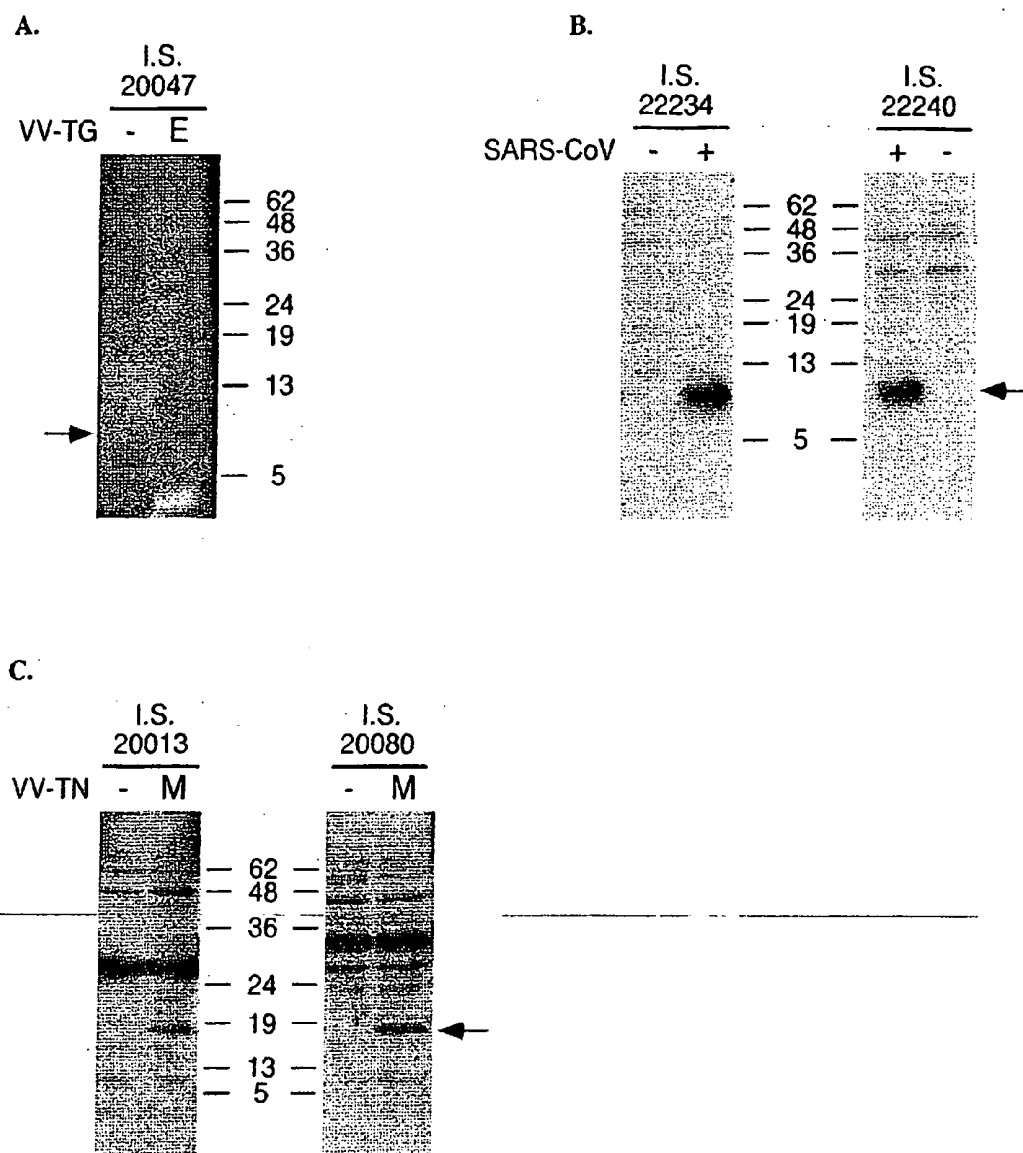


FIGURE 43

NOVEL STRAIN OF SARS-ASSOCIATED CORONAVIRUS AND APPLICATIONS THEREOF

[0001] The present invention relates to a novel strain of severe acute respiratory syndrome (SARS)-associated coronavirus derived from a sample recorded under No. 031589 and collected in Hanoi (Vietnam), to nucleic acid molecules derived from its genome, to the proteins and peptides encoded by said nucleic acid molecules and to their applications, in particular as diagnostic reagents and/or as vaccine.

[0002] Coronavirus is a virus containing single-stranded RNA, of positive polarity, of approximately 30 kilobases which replicates in the cytoplasm of the host cells; the 5' end of the genome has a capped structure and the 3' end contains a polyA tail. This virus is enveloped and comprises, at its surface, peplomeric structures called spicules.

[0003] The genome comprises the following open reading frames or ORFs, from its 5' end to its 3' end: ORF1a and ORF1b corresponding to the proteins of the transcription-replication complex, and ORF-S, ORF-E, ORF-M and ORF-N corresponding to the structural proteins S, E, M and N. It also comprises ORFs corresponding to proteins of unknown function encoded by: the region situated between ORF-S and ORF-E and overlapping the latter, the region situated between ORF-M and ORF-N, and the region included in ORF-N.

[0004] The S protein is a membrane glycoprotein (200-220 kDa) which exists in the form of spicules or spikes emerging from the surface of the viral envelope. It is responsible for the attachment of the virus to the receptors of the host cell and for inducing the fusion of the viral envelope with the cell membrane.

[0005] The small envelope protein (E), also called sM (small membrane), which is a nonglycosylated transmembrane protein of about 10 kDa, is the protein present in the smallest quantity in the virion. It plays a powerful role in the coronavirus budding process which occurs at the level of the intermediate compartment in the endoplasmic reticulum and the Golgi apparatus.

[0006] The M protein or matrix protein (25-30 kDa) is a more abundant membrane glycoprotein which is integrated into the viral particle by an M/E interaction, whereas the incorporation of S into the particles is directed by an S/M interaction. It appears to be important for the viral maturation of coronaviruses and for the determination of the site where the viral particles are assembled.

[0007] The N protein or nucleocapsid protein (45-50 kDa) which is the most conserved among the coronavirus structural proteins is necessary for encapsidating the genomic RNA and then for directing its incorporation into the virion. This protein is probably also involved in the replication of the RNA.

[0008] When the host cell is infected, the reading frame (ORF) situated in 5' of the viral genome is translated into a polyprotein which is cleaved by the viral proteases and then releases several nonstructural proteins such as the RNA-dependent RNA polymerase (Rep) and the ATPase helicase (Hel). These two proteins are involved in the replication of the viral genome and in the generation of transcripts which are used in the synthesis of the viral proteins. The mechanisms

by which these subgenomic mRNAs are produced are not completely understood; however, recent facts indicate that the sequences for regulation of transcription at the 5' end of each gene represent signals which regulate the discontinuous transcription of the subgenomic mRNAs.

[0009] The proteins of the viral membrane (S, E and M proteins) are inserted into the intermediate compartment, whereas the replicated RNA (+ strand) is assembled with the N (nucleocapsid) protein. This protein-RNA complex then combines with the M protein contained in the membranes of the endoplasmic reticulum and the viral particles form when the nucleocapsid complex buds into the endoplasmic reticulum. The virus then migrates across the Golgi complex and eventually leaves the cell, for example by exocytosis. The site of attachment of the virus to the host cell is at the level of the S protein.

[0010] Coronaviruses are responsible for 15 to 30% of colds in humans and for respiratory and digestive infections in animals, especially cats (FIPV: Feline infectious peritonitis virus), poultry (IBV: Avian infectious bronchitis virus), mice (MHV: Mouse hepatitis virus), pigs (TGEV: Transmissible gastroenteritis virus, PEDV: Porcine Epidemic diarrhea virus, PRCoV: Porcine Respiratory Coronavirus, HEV: Hemagglutinating encephalomyelitis Virus) and bovines (BCoV: Bovine coronavirus).

[0011] In general, each coronavirus affects only one species; in immunocompetent individuals, the infection induces optionally neutralizing antibodies and cell immunity, capable of destroying the infected cells.

[0012] An epidemic of atypical pneumonia, called severe acute respiratory syndrome (SARS) has spread in various countries (Vietnam, Hong Kong, Singapore, Thailand and Canada) during the first quarter of 2003, from an initial focus which appeared in China in the last quarter of 2002. The severity of this disease is such that its mortality rate is about 3 to 6%. The determination of the causative agent of this disease is underway by numerous laboratories worldwide.

[0013] In March 2003, a new coronavirus (SARS-CoV or SARS virus) was isolated, in association with cases of severe acute respiratory syndrome (T. G. KSIAZEK et al., *The New England Journal of Medicine*, 2003, 348, 1319-1330; C. DROSTEN et al., *The New England Journal of Medicine*, 2003, 348, 1967-1976; Peiris et al., *Lancet*, 2003, 361, 1319).

[0014] Genomic sequences of this new coronavirus have thus been obtained, in particular those of the Urbani isolate (Genbank accession No. AY274119.3 and A. MARRA et al., *Science*, May 1, 2003, 300, 1399-1404) and the Toronto isolate (Tor2, Genbank accession No. AY278741 and A. ROTA et al., *Science*, 2003, 300, 1394-1399).

[0015] The organization of the genome is comparable with that of other known coronaviruses, thus making it possible to confirm that SARS-CoV belongs to the Coronaviridae family; open reading frames ORF1a and 1b and open reading frames corresponding to the S, E, M and N proteins, and to proteins encoded by: the region situated between ORF-S and ORF-E (ORF3), the region situated between ORF-S and ORF-E and overlapping ORF-E (ORF4), the region situated between ORF-M and ORF-N (ORF7) to

ORF11) and the region corresponding to ORF-N (ORF13 and ORF14), have in particular been identified.

[0016] Seven differences have been identified between the sequences of the Tor2 and Urbani isolates; 3 correspond to silent mutations (c/t at position 16622 and a/g at position 19064 of ORF1b, t/c at position 24872 of ORF-S) and 4 modify the amino acid sequence of respectively: the proteins encoded by ORF1a (c/t at position 7919 corresponding to the A/V mutation), the S protein (g/t at position 23220 corresponding to the A/S mutation), the protein encoded by ORF3 (a/g at position 25298 corresponding to the R/G mutation) and the M protein (t/c at position 26857 corresponding to the S/P mutation).

[0017] In addition, phylogenetic analysis shows that SARS-CoV is distant from other coronaviruses and that it did not appear by mutation of human respiratory coronaviruses nor by recombination between known coronaviruses (for a review, see Holmes, J. C. I., 2003, 111, 1605-1609).

[0018] The determination and the taking into account of new variants are important for the development of reagents for the detection and diagnosis of SARS which are sufficiently sensitive and specific, and immunogenic compositions capable of protecting populations against epidemics of SARS.

[0019] The inventors have now identified another strain of SARS-associated coronavirus which is distinguishable from the Tor2 and Urbani isolates.

[0020] The subject of the present invention is therefore an isolated or purified strain of severe acute respiratory syndrome-associated human coronavirus, characterized in that its genome has, in the form of complementary DNA, a serine codon at position 23220-23222 of the gene for the S protein or a glycine codon at position 25298-25300 of the gene for ORF3, and an alanine codon at position 7918-7920 of ORF1a or a serine codon at position 26857-26859 of the gene for the M protein, said positions being indicated in terms of reference to the Genbank sequence AY274119.3.

[0021] According to an advantageous embodiment of said strain, the DNA equivalent of its genome has a sequence corresponding to the sequence SEQ ID No: 1; this coronavirus strain is derived from the sample collected from the bronchoalveolar washings from a patient suffering from SARS, recorded under the No. 031589 and collected at the Hanoi (Vietnam) French hospital.

[0022] In accordance with the invention, said sequence SEQ ID No: 1 is that of the deoxyribonucleic acid corresponding to the ribonucleic acid molecule of the genome of the isolated coronavirus strain as defined above.

[0023] The sequence SEQ ID No: 1 is distinguishable from the Genbank sequence AY274119.3 (Tor2 isolate) in that it possesses the following mutations:

[0024] g/t at position 23220; the alanine codon (gct) at position 577 of the amino acid sequence of the Tor2 S protein is replaced by a serine codon (tct),

[0025] a/g at position 25298; the arginine codon (aga) at position 11 of the amino acid sequence of the protein encoded by the Tor2 ORF3 is replaced by a glycine codon (gga).

[0026] In addition, the sequence SEQ ID No: 1 is distinguishable from the Genbank sequence AY278741 (Urbani isolate) in that it possesses the following mutations:

[0027] t/c at position 7919; the valine codon (gtt) in position 2552 of the amino acid sequence of the protein encoded by ORF1a is replaced by an alanine codon (gct),

[0028] t/c at position 16622: this mutation does not modify the amino acid sequence of the proteins encoded by ORF1b (silent mutation),

[0029] g/a at position 19064: this mutation does not modify the amino acid sequence of the proteins encoded by ORF1b (silent mutation),

[0030] c/t at position 24872: this mutation does not modify the amino acid sequence of the S protein, and

[0031] c/t at position 26857: the proline codon (ccc) at position 154 of the amino acid sequence of the M protein is replaced by a serine codon (tcc).

[0032] Unless otherwise stated, the positions of the nucleotide and peptide sequences are indicated with reference to the Genbank sequence AY274119.3.

[0033] The subject of the present invention is also an isolated or purified polynucleotide, characterized in that its sequence is that of the genome of the isolated coronavirus strain as defined above.

[0034] According to an advantageous embodiment of said polynucleotide, it has the sequence SEQ ID No: 1.

[0035] The subject of the present invention is also an isolated or purified polynucleotide, characterized in that its sequence hybridizes under high stringency conditions with the sequence of the polynucleotide as defined above.

[0036] The terms "isolated or purified" mean modified "by the hand of humans" from the natural state; in other words if an object exists in nature, it is said to be isolated or purified if it is modified or extracted from its natural environment or both. For example, a polynucleotide or a protein/peptide naturally present in a living organism is neither isolated nor purified; on the other hand, the same polynucleotide or protein/peptide separated from coexisting molecules in its natural environment, obtained by cloning, amplification and/or chemical synthesis is isolated for the purposes of the present invention. Furthermore, a polynucleotide or a protein/peptide which is introduced into an organism by transformation, genetic manipulation or by any other method, is "isolated" even if it is present in said organism. The term purified as used in the present invention means that the proteins/peptides according to the invention are essentially free of association with the other proteins or polypeptides, as is for example the product purified from the culture of recombinant host cells or the product purified from a nonrecombinant source.

[0037] For the purposes of the present invention, high stringency hybridization conditions are understood to mean temperature and ionic strength conditions chosen such that they make it possible to maintain the specific and selective hybridization between complementary polynucleotides.

[0038] By way of illustration, high stringency conditions for the purposes of defining the above polynucleotides are

advantageously the following: the DNA-DNA or DNA-RNA hybridization is performed in two steps: (1) prehybridization at 42° C. for 3 hours in phosphate buffer (20 mM, pH 7.5) containing 5×SSC (1×SSC corresponds to a 0.15 M NaCl+0.015 M sodium citrate solution), 50% formamide, 7% sodium dodecyl sulfate (SDS), 10× Denhardt's, 5% dextran sulfate and 1% salmon sperm DNA; (2) hybridization for 20 hours at 42° C. followed by 2 washings of 20 minutes at 20° C. in 2×SSC+2% SDS, 1 washing of 20 minutes at 20° C. in 0.1×SSC+0.1% SDS. The final washing is performed in 0.1×SSC+0.1% SDS for 30 minutes at 60° C.

[0039] The subject of the present invention is also a representative fragment of the polynucleotide as defined above, characterized in that it is capable of being obtained either by the use of restriction enzymes whose recognition and cleavage sites are present in said polynucleotide as defined above, or by amplification with the aid of oligonucleotide primers specific for said polynucleotide as defined above, or by transcription *in vitro*, or by chemical synthesis.

[0040] According to an advantageous embodiment of said fragment, it is selected from the group consisting of: the cDNA corresponding to at least one open reading frame (ORF) chosen from: ORF1a, ORF1b, ORF-S, ORF-E, ORF-M, ORF-N, ORF3, ORF4, ORF7 to ORF11, ORF13 and ORF14 and the cDNA corresponding to the noncoding 5' or 3' ends of said polynucleotide.

[0041] According to an advantageous feature of this embodiment, said fragment has a sequence selected from the group consisting of:

[0042] the sequences SEQ ID NO: 2 and 4 representing the cDNA corresponding to the ORF-S which encodes the S protein,

[0043] the sequences SEQ ID NO: 13 and 15 representing the cDNA corresponding to the ORF-E which encodes the E protein,

[0044] the sequences SEQ ID NO: 16 and 18 representing the cDNA corresponding to the ORF-M which encodes the M protein,

[0045] the sequences SEQ ID NO: 36 and 38 representing the cDNA corresponding to the ORF-N which encodes the N protein,

[0046] the sequences representing the cDNA corresponding respectively: to ORF1a and ORF1b (SEQ ID NO: 31), to ORF3 and ORF4 (SEQ ID NO: 7, 8), to ORF7 to 11 (SEQ ID NO: 19, 20) to ORF13 (SEQ ID NO: 32) and to ORF14 (SEQ ID NO: 34), and

[0047] the sequences representing the cDNAs corresponding respectively to the noncoding 5' (SEQ ID NO: 39 and 72) and 3' (SEQ ID NO: 40, 73) ends of said polynucleotide.

[0048] The subject of the present invention is also a cDNA fragment encoding the S protein, as defined above, characterized in that it has a sequence selected from the group consisting of the sequences SEQ ID NO: 5 and 6 (Sa and Sb fragments).

[0049] The subject of the present invention is also a cDNA fragment corresponding to ORF1a and ORF1b as defined

above, characterized in that it has a sequence selected from the group consisting of the sequences SEQ ID NO: 41 to 54 (L0 to L12 fragments).

[0050] The subject of the present invention is also a polynucleotide fragment as defined above, characterized in that it has at least 15 consecutive bases or base pairs of the sequence of the genome of said strain including at least one of those situated in position 7979, 16622, 19064, 23220, 24872, 25298 and 26857. Preferably this is a fragment of 20 to 2500 bases or base pairs, preferably from 20 to 400.

[0051] According to an advantageous embodiment of said fragment, it includes at least one pair of bases or base pairs corresponding to the following positions: 7919 and 23220, 7919 and 25298, 16622 and 23220, 19064 and 23220, 16622 and 25298, 19064 and 25298, 23220 and 24872, 23220 and 26857, 24872 and 25298, 25298 and 26857.

[0052] The subject of the present invention is also primers of at least 18 bases capable of amplifying a fragment of the genome of a SARS-associated coronavirus or of the DNA equivalent thereof.

[0053] According to an embodiment of said primers, they are selected from the group consisting of:

[0054] the pair of primers No. 1 corresponding respectively to positions 28507 to 28522 (sense primer, SEQ ID NO: 60) and 28774 to 28759 (antisense primer, SEQ ID NO: 61) of the sequence of the polynucleotide as defined above,

[0055] the pair of primers No. 2 corresponding respectively to positions 28375 to 28390 (sense primer, SEQ ID NO: 62) and 28702 to 28687 (antisense primer, SEQ ID NO: 63) of the sequence of the polynucleotide as defined above, and

[0056] the pair of primers consisting of the primers SEQ ID Nos: 55 and 56.

[0057] The subject of the present invention is also a probe capable of detecting the presence of the genome of a SARS-associated coronavirus or of a fragment thereof, characterized in that it is selected from the group consisting of: the fragments as defined above and the fragments corresponding to the following positions of the polynucleotide sequence as defined above: 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 (SEQ ID NO: 64 to 67).

[0058] The probes and primers according to the invention may be labeled directly or indirectly with a radioactive or nonradioactive compound by methods well known to persons skilled in the art so as to obtain a detectable and/or quantifiable signal. Among the radioactive isotopes used, there may be mentioned ³²P, ³³P, ³⁵S, ³H or ¹²⁵I. The nonradioactive entities are selected from ligands such as biotin, avidin, streptavidin, digoxigenin, haptens, dyes, luminescent agents such as radioluminescent, chemoluminescent, bioluminescent, fluorescent and phosphorescent agents.

[0059] The invention encompasses the labeled probes and primers derived from the preceding sequences.

[0060] Such probes and primers are useful for the diagnosis of infection by a SARS-associated coronavirus.

[0061] The subject of the present invention is also a method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

[0062] (a) the extraction of nucleic acids present in said biological sample,

[0063] (b) the amplification of a fragment of ORF-N by RT-PCR with the aid of a pair of primers as defined above, and

[0064] (c) the detection, by any appropriate means, of the amplification products obtained in (b).

[0065] The amplification products (amplicons) in (b) are 268 bp for the pair of primers No. 1 and 328 bp for the pair of primers No. 2.

[0066] According to an advantageous embodiment of said method, the step (b) of detection is carried out with the aid of at least one probe corresponding to positions 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 of the sequence of the polynucleotide as defined above.

[0067] Preferably, the SARS-associated coronavirus genome is detected and optionally quantified by PCR in real time with the aid of the pair of primers No. 2 and probes corresponding to positions 28541 to 28563 and 28565 to 28589 labeled with different compounds, in particular different fluorescent agents.

[0068] The real time RT-PCR which uses this pair of primers and this probe is very sensitive since it makes it possible to detect 10^2 copies of RNA and up to 10 copies of RNA; it is in addition reliable and reproducible.

[0069] The invention encompasses the single-stranded, double-stranded and triple-stranded polydeoxyribonucleotides and polyribonucleotides corresponding to the sequence of the genome of the isolated strain of coronavirus and its fragments as defined above, and to their sense or antisense complementary sequences, in particular the RNAs and cDNAs corresponding to the sequence of the genome and of its fragments as defined above.

[0070] The present invention also encompasses the amplification fragments obtained with the aid of primers specific for the genome of the purified or isolated strain as defined above, in particular with the aid of primers or pairs of primers as defined above, the restriction fragments formed by or comprising the sequence of fragments as defined above, the fragments obtained by transcription in vitro from a vector containing the sequence SEQ ID NO: 1 or a fragment as defined above, and fragments obtained by chemical synthesis. Examples of restriction fragments are deduced from the restriction map of the sequence SEQ ID NO: 1 illustrated by FIG. 13. In accordance with the invention, said fragments are either in the form of isolated fragments, or in the form of mixtures of fragments. The invention also encompasses fragments modified, in relation to the preceding ones, by removal or addition of nucleotides in a proportion of about 15%, relative to the length of the above fragments and/or modified in terms of the nature of the nucleotides, as long as the modified nucleotide fragments retain a capacity for hybridization with the genomic or antigenomic RNA sequences of the isolate as defined above.

[0071] The nucleic acid molecules according to the invention are obtained by conventional methods, known per se, following standard protocols such as those described in *Current Protocols in Molecular Biology* (Frederick M. AUSUBEL, 2000, Wiley and son Inc., Library of Congress, USA). For example, they may be obtained by amplification of a nucleic sequence by PCR or RT-PCR or alternatively by total or partial chemical synthesis.

[0072] The subject of the present invention is also a DNA or RNA chip or filter, characterized in that it comprises at least one polynucleotide or one of its fragments as defined above.

[0073] The DNA or RNA chips or filters according to the invention are prepared by conventional methods, known per se, such as for example chemical or electrochemical grafting of oligonucleotides on a glass or nylon support.

[0074] The subject of the present invention is also a recombinant cloning and/or expression vector, in particular a plasmid, a virus, a viral vector or a phage comprising a nucleic acid fragment as defined above. Preferably, said recombinant vector is an expression vector in which said nucleic acid fragment is placed under the control of appropriate elements for regulating transcription and translation. In addition, said vector may comprise sequences (tags) fused in phase with the 5' and/or 3' end of said insert, which are useful for the immobilization and/or detection and/or purification of the protein expressed from said vector.

[0075] These vectors are constructed and introduced into host cells by conventional recombinant DNA and genetic engineering methods which are known per se. Numerous vectors into which a nucleic acid molecule of interest may be inserted in order to introduce it and to maintain it in a host cell are known per se; the choice of an appropriate vector depends on the use envisaged for this vector (for example replication of the sequence of interest, expression of this sequence, maintenance of the sequence in extrachromosomal form or alternatively integration into the chromosomal material of the host), and on the nature of the host cell.

[0076] In accordance with the invention, said plasmid is selected in particular from the following plasmids:

[0077] the plasmid, called SARS-S, contained in the bacterial strain deposited under the No. I-3659, on Jun. 20, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, said sequence corresponding to the nucleotides at positions 21406 to 25348 (SEQ ID NO: 4), with reference to the Genbank sequence AY274119.3,

[0078] the plasmid, called SARS-S1, contained in the bacterial strain deposited under the No. I-3020, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 5' fragment of the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment corresponding to the nucleotides at positions 21406 to 23454 (SEQ ID NO: 5), with reference to the Genbank sequence AY274119.3 Tor2,

- [0079] the plasmid, called SARS-S2, contained in the bacterial strain deposited under the No. I-3019, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 3' fragment of the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the number No. 031589, as defined above, said fragment corresponding to the nucleotides at positions 23322 to 25348 (SEQ ID NO: 6), with reference to the Genbank sequence accession No. AY274119.3,
- [0080] the plasmid, called SARS-SE, contained in the bacterial strain deposited under the No. I-3126, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the region situated between ORF-S and ORF-E and overlapping ORF-E of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said region corresponding to the nucleotides at positions 25110 to 26244 (SEQ ID NO: 8), with reference to the Genbank sequence accession No. AY274119.3,
- [0081] the plasmid, called SARS-E, contained in the bacterial strain deposited under the No. I-3046, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the E protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 26082 to 26413 (SEQ ID NO: 15), with reference to the Genbank sequence accession No. AY274119.3,
- [0082] the plasmid, called SARS-M, contained in the bacterial strain deposited under the No. I-3047, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above; said sequence corresponding to the nucleotides at positions 26330 to 27098 (SEQ ID NO: 18), with reference to the Genbank sequence accession No. AY274119.3,
- [0083] the plasmid, called SARS-MN, contained in the bacterial sequence deposited under the No. I-3125, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the region situated between ORF-M and ORF-N of the SARS-CoV strain derived from the sample recorded under the No. 031589 and collected in Hanoi, as defined above, said sequence corresponding to the nucleotides at positions 26977 to 28218 (SEQ ID NO: 20), with reference to the Genbank accession No. AY274119.3,
- [0084] the plasmid, called SARS-N, contained in the bacterial strain deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA encoding the N protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 28054 to 29430 (SEQ ID NO: 38), with reference to the Genbank sequence accession No. AY274119.3; thus, this plasmid comprises an insert of sequence SEQ ID NO: 38 and is contained in a bacterial strain which was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15,
- [0085] the plasmid, called SARS-5'NC, contained in the bacterial strain deposited under the No. I-3124, on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the noncoding 5' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 1 to 204 (SEQ ID NO: 39), with reference to the Genbank sequence accession No. AY274119.3,
- [0086] the plasmid called SARS-3'NC, contained in the bacterial strain deposited under the No. I-3123 on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the noncoding 3' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to that situated between the nucleotide and position 28933 to 29727 (SEQ ID NO: 40), with reference to the Genbank sequence accession No. AY274119.3, ends with a series of nucleotides a.,
- [0087] the expression plasmid, called pIV2.3N, containing a cDNA fragment encoding a C-terminal fusion of the N protein (SEQ ID NO: 37) with a polyhistidine tag,
- [0088] the expression plasmid, called pIV2.3S_C, containing a cDNA fragment encoding a C-terminal fusion of the fragment corresponding to positions 475 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag,
- [0089] the expression plasmid, pIV2.3S_L, containing a cDNA fragment encoding a C-terminal fusion of the fragment corresponding to positions 14 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag,
- [0090] the expression plasmid, called pIV2.4N, containing a cDNA fragment encoding a N-terminal fusion of the N protein (SEQ ID NO: 3) with a polyhistidine tag,
- [0091] the expression plasmid, called pIV2.4S_C or pIV2.4S_L, containing an insert encoding a N-terminal fusion of the fragment corresponding to positions 475 to 1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag, and
- [0092] the expression plasmid, called pIV2.4S_L, containing a cDNA fragment encoding an N-terminal fusion of the fragment corresponding to positions 14 to

1193 of the amino acid sequence of the S protein (SEQ ID NO: 3) with a polyhistidine tag.

[0093] According to an advantageous feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited under the No. I-3117, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

[0094] According to another advantageous feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited under the No. I-3118, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

[0095] According to another feature of the expression plasmid as defined above, it is contained in a bacterial strain which was deposited at the CNCM, 25 rue du Docteur Roux, 75724 Paris Cedex 15 under the following numbers:

- [0096] a) strain No. I-3118, deposited on Oct. 23, 2003,
- [0097] b) strain No. I-3019, deposited on May 12, 2003,
- [0098] c) strain No. I-3020, deposited on May 12, 2003,
- [0099] d) strain No. I-3059, deposited on Jun. 20, 2003,
- [0100] e) strain No. I-3323, deposited on Nov. 22, 2004,
- [0101] f) strain No. I-3324, deposited on Nov. 22, 2004,
- [0102] g) strain No. I-3326, deposited on Dec. 1, 2004,
- [0103] h) strain No. I-3327, deposited on Dec. 1, 2004,
- [0104] i) strain No. I-3332, deposited on Dec. 1, 2004,
- [0105] j) strain No. I-3333, deposited on Dec. 1, 2004,
- [0106] k) strain No. I-3334, deposited on Dec. 1, 2004,
- [0107] l) strain No. I-3335, deposited on Dec. 1, 2004,
- [0108] m) strain No. I-3336, deposited on Dec. 1, 2004,
- [0109] n) strain No. I-3337, deposited on Dec. 1, 2004,
- [0110] o) strain No. I-3338, deposited on Dec. 2, 2004,
- [0111] p) strain No. I-3339, deposited on Dec. 2, 2004,
- [0112] q) strain No. I-3340, deposited on Dec. 2, 2004,
- [0113] r) strain No. I-3341, deposited on Dec. 2, 2004.

[0114] The subject of the present invention is also a nucleic acid insert of viral origin, characterized in that it is contained in any of the strains as defined above in a)-r).

[0115] The subject of the present invention is also a nucleic acid containing a synthetic gene allowing optimized expression of the S protein in eukaryotic cells, characterized in that it possesses the sequence SEQ ID NO: 140.

[0116] The subject of the present invention is also an expression vector containing a nucleic acid containing a synthetic gene allowing optimized expression of the S protein, which vector is contained in the bacterial strain deposited at the CNCM, on Dec. 1, 2004, under the No. I-3333.

[0117] According to one embodiment of said expression vector, it is a viral vector, in the form of a viral particle or in the form of a recombinant genome.

[0118] According to an advantageous feature of this embodiment, this is a recombinant viral particle or a recombinant viral genome capable of being obtained by transfection of a plasmid according to paragraphs g), h) and k) to r) as defined above, in an appropriate cellular system, that is to say, for example, cells transfected with one or more other plasmids intended to transcomplement certain functions of the virus that are deleted in the vector and that are necessary for the formation of the viral particles.

[0119] The expression "S protein family" is understood here to mean the complete S protein, its ectodomain and fragments of this ectodomain which are preferably produced in a eukaryotic system.

[0120] The subject of the present invention is also a lentiviral vector encoding a polypeptide of the S protein family, as defined above.

[0121] The subject of the present invention is also a recombinant measles virus encoding a polypeptide of the S protein family, as defined above.

[0122] The subject of the present invention is also a recombinant vaccinia virus encoding a polypeptide of the S protein family, as defined above.

[0123] The subject of the present invention is also the use of a vector according to paragraphs e) to r) as defined above, or of a vector containing a synthetic gene for the S protein, as defined above, for the production, in a eukaryotic system, of the SARS-associated coronavirus S protein or of a fragment of this protein.

[0124] The subject of the present invention is also a method for producing the S protein in a eukaryotic system, comprising a step of transfecting eukaryotic cells in culture with a vector chosen from the vectors contained in the bacterial strains mentioned in paragraphs e) to r) above or a vector containing a synthetic gene allowing optimized expression of the S protein.

[0125] The subject of the present invention is also a cDNA library characterized in that it comprises fragments as defined above, in particular amplification fragments or restriction fragments, cloned into a recombinant vector, in particular an expression vector (expression library).

[0126] The subject of the present invention is also cells, in particular prokaryotic cells, modified by a recombinant vector as defined above.

[0127] The subject of the present invention is also a genetically modified eukaryotic cell expressing a protein or a polypeptide as defined above. Quite obviously, the terms "genetically modified eukaryotic cell" do not denote a cell modified with a wild-type virus.

[0128] According to an advantageous embodiment of said cell, it is capable of being obtained by transfection with any of the vectors mentioned in paragraphs i) to l) above.

[0129] According to an advantageous feature of this embodiment, this is the cell FRhK4-Sso1-30, deposited at the CNCM on Nov. 22, 2004, under the No. I-3325.

[0130] The recombinant vectors as defined above and the cells transformed with said expression vectors are advantageously used for the production of the corresponding proteins and peptides. The expression libraries derived from

said vectors, and the cells transformed with said expression libraries are advantageously used to identify the immunogenic epitopes (B and T epitopes) of the SARS-associated coronavirus proteins.

[0131] The subject of the present invention is also the purified or isolated proteins and peptides, characterized in that they are encoded by the polynucleotide or one of its fragments as defined above.

[0132] According to an advantageous embodiment of the invention, said protein is selected from the group consisting of:

[0133] the S protein having the sequence SEQ ID NO: 3 or its ectodomaine

[0134] the E protein having the sequence SEQ ID NO: 14

[0135] the M protein having the sequence SEQ ID NO: 17

[0136] the N protein having the sequence SEQ ID NO: 37

[0137] the proteins encoded by the ORFs: ORF1a, ORF1b, ORF3, ORF4 and ORF7 to ORF11, ORF13 and ORF14 and having the respective sequence, SEQ ID NO: 74, 75, 10, 12, 22, 24, 26, 28, 30, 33 and 35.

[0138] The terms "ectodomaine of the S protein" and "soluble form of the S protein" will be used interchangeably below.

[0139] According to an advantageous embodiment of the invention, said polypeptide consists of the amino acids corresponding to positions 1 to 1193 of the amino acid sequence of the S protein.

[0140] According to another advantageous embodiment of the invention, said peptide is selected from the group consisting of:

[0141] a) the peptides corresponding to positions 14 to 1193 and 475 to 1193 of the amino acid sequence of the S protein,

[0142] b) the peptides corresponding to positions 2 to 14 (SEQ ID NO: 69) and 100 to 221 of the amino acid sequence of the M protein; these peptides correspond respectively to the ectodomaine and to the endodomaine of the M protein, and

[0143] c) the peptides corresponding to positions 1 to 12 (SEQ ID NO: 70) and 53 to 76 (SEQ ID NO: 71) of the amino acid sequence of the E protein; these peptides correspond respectively to the ectodomaine and to the C-terminal end of the E protein, and

[0144] d) the peptides of 5 to 50 consecutive amino acids, preferably of 10 to 30 amino acids, inclusive or partially or completely overlapping the sequence of the peptides as defined in a), b) or c).

[0145] The subject of the present invention is also a peptide, characterized in that it has a sequence of 7 to 50 amino acids including an amino acid residue selected from the group consisting of:

[0146] the alanine situated at position 2552 of the amino acid sequence of the protein encoded by ORF1a,

[0147] the serine situated at position 577 of the amino acid sequence of the S protein of the SARS-CoV strain as defined above,

[0148] the glycine at position 11 of the amino acid sequence of the protein encoded by ORF3 of the SARS-CoV strain as defined above,

[0149] the serine at position 154 of the amino acid sequence of the M protein of the SARS-CoV strain as defined above.

[0150] The subject of the present invention is also an antibody or a polyclonal or monoclonal antibody fragment which can be obtained by immunization of an animal with a recombinant vector as defined above, a cDNA library as defined above or alternatively a protein or a peptide as defined above, characterized in that it binds to at least one of the proteins encoded by SARS-CoV as defined above.

[0151] The invention encompasses the polyclonal antibodies, the monoclonal antibodies, the chimeric antibodies such as the humanized antibodies, and fragments thereof (Fab, Fv, scFv).

[0152] A subject of the present invention is also a hybridoma producing a monoclonal antibody against the N protein, characterized in that it is chosen from the following hybridomas:

[0153] the hybridoma producing the monoclonal antibody 87, deposited at the CNCM on Dec. 1, 2004 under the number I-3328,

[0154] the hybridoma producing the monoclonal antibody 86, deposited at the CNCM on Dec. 1, 2004 under the number I-3329,

[0155] the hybridoma producing the monoclonal antibody 57, deposited at the CNCM on Dec. 1, 2004 under the number I-3330, and

[0156] the hybridoma producing the monoclonal antibody 156, deposited at the CNCM on Dec. 1, 2004 under the number I-3331.

[0157] The subject of the present invention is also a polyclonal or monoclonal antibody or antibody fragment directed against the N protein, characterized in that it is produced by a hybridoma as defined above.

[0158] For the purposes of the present invention, the expression chimeric antibody is understood to mean, in relation to an antibody of a particular animal species or of a particular class of antibody, an antibody comprising all or part of a heavy chain and/or of a light chain of an antibody of another animal species or of another class of antibody.

[0159] For the purposes of the present invention, the expression humanized antibody is understood to mean a human immunoglobulin in which the residues of the CDRs (Complementary Determining Regions) which form the antigen-binding site are replaced by those of a nonhuman monoclonal antibody possessing the desired specificity, affinity or activity. Compared with the nonhuman antibodies, the humanized antibodies are less immunogenic and possess a prolonged half-life in humans because they possess only a small proportion of nonhuman sequences given that practically all the residues of the FR (Framework) regions and of

the constant (Fc) region of these antibodies are those of a consensus sequence of human immunoglobulins.

[0160] A subject of the present invention is also a protein chip or filter, characterized in that it comprises a protein, a peptide or alternatively an antibody as defined above.

[0161] The protein chips according to the invention are prepared by conventional methods known per se. Among the appropriate supports on which proteins may be immobilized, there may be mentioned those made of plastic or glass, in particular in the form of microplates.

[0162] The subject of the present invention is also reagents derived from the isolated strain of SARS-associated coronavirus, derived from the sample recorded under the No. 031589, which are useful for the study and diagnosis of the infection caused by a SARS-associated coronavirus, said reagents are selected from the group consisting of:

[0163] (a) a pair of primers, a probe or a DNA chip as defined above,

[0164] (b) a recombinant vector or a modified cell as defined above,

[0165] (c) an isolated coronavirus strain or a polynucleotide as defined above,

[0166] (d) a protein or a peptide as defined above,

[0167] (e) an antibody or an antibody fragment as defined above, and

[0168] (f) a protein chip as defined above.

[0169] These various reagents are prepared and used according to conventional molecular biology and immunology techniques following standard protocols such as those described in *Current Protocols in Molecular Biology* (Frederick M. AUSUBEL, 2000, Wiley and Son Inc., Library of Congress, USA), in *Current Protocols in Immunology* (John E. Coligan, 2000, Wiley and Son Inc., Library of Congress, USA) and in *Antibodies: A Laboratory Manual* (E. Howell and D. Lane, Cold Spring Harbor Laboratory, 1988).

[0170] The nucleic acid fragments according to the invention are prepared and used according to conventional techniques as defined above. The peptides and proteins according to the invention are prepared by recombinant DNA techniques, known to persons skilled in the art, in particular with the aid of the recombinant vectors as defined above. Alternatively, the peptides according to the invention may be prepared by conventional techniques of solid or liquid phase synthesis, known to persons skilled in the art.

[0171] The polyclonal antibodies are prepared by immunizing an appropriate animal with a protein or a peptide as defined above, optionally coupled to KLH or to albumin and/or combined with an appropriate adjuvant such as (complete or incomplete) Freund's adjuvant or aluminum hydroxide; after obtaining a satisfactory antibody titer, the antibodies are harvested by collecting serum from the immunized animals and enriched with IgG by precipitation, according to conventional techniques, and then the IgGs specific for the SARS-CoV proteins are optionally purified by affinity chromatography on an appropriate column to which said peptide or said protein is attached, as defined above, so as to obtain a monospecific IgG preparation.

[0172] The monoclonal antibodies are produced from hybridomas obtained by fusion of B lymphocytes from an animal immunized with a protein or a peptide as defined above with myelomas, according to the Köhler and Milstein technique (Nature, 1975, 256, 495-497); the hybridomas are cultured in vitro, in particular in fermenters or produced in vivo, in the form of ascites; alternatively, said monoclonal antibodies are produced by genetic engineering as described in American U.S. Pat. No. 4,816,567.

[0173] The humanized antibodies are produced by general methods such as those described in International application WO 98/45332.

[0174] The antibody fragments are produced from the cloned V_H and V_L regions, from the mRNAs of hybridomas or splenic lymphocytes of an immunized mouse; for example, the Fv, scFv or Fab fragments are expressed at the surface of filamentous phages according to the Winter and Milstein technique (Nature, 1991, 349, 293-299); after several selection steps, the antibody fragments specific for the antigen are isolated and expressed in an appropriate expression system, by conventional techniques for cloning and expression of recombinant DNA.

[0175] The antibodies or fragments thereof as defined above are purified by conventional techniques known to persons skilled in the art, such as affinity chromatography.

[0176] The subject of the present invention is additionally the use of a product selected from the group consisting of: a pair of primers, a probe, a DNA chip, a recombinant vector, a modified cell, an isolated coronavirus strain, a polynucleotide, a protein or a peptide, an antibody or an antibody fragment and a protein chip as defined above, for the preparation of a reagent for the detection and optionally genotyping/serotyping of a SARS-associated coronavirus.

[0177] The proteins and peptides according to the invention, which are capable of being recognized and/or of inducing the production of antibodies specific for the SARS-associated coronavirus, are useful for the diagnosis of infection with such a coronavirus; the infection is detected, by an appropriate technique—in particular EIA, ELISA, RIA, immunofluorescence—in a biological sample collected from an individual capable of being infected.

[0178] According to an advantageous feature of said use, said proteins are selected from the group consisting of the S, E, M and/or N proteins and the peptides as defined above.

[0179] The S, E, M and/or N proteins and the peptides derived from these proteins as defined above, for example the N protein, are used for the indirect diagnosis of a SARS-associated coronavirus infection (serological diagnosis; detection of an antibody specific for SARS-CoV), in particular by an immunoenzymatic method (ELISA).

[0180] The antibodies and antibody fragments according to the invention, in particular those directed against the S, E, M and/or N proteins and the derived peptides as defined above, are useful for the direct diagnosis of a SARS-associated coronavirus infection; the detection of the protein(s) of SARS-CoV is carried out by an appropriate technique, in particular EIA, ELISA, RIA, immunofluorescence, in a biological sample collected from an individual capable of being infected.

[0181] The subject of the present invention is also a method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

[0182] (a) bringing said biological sample into contact with at least one antibody or one antibody fragment, one protein, one peptide or alternatively one protein or peptide chip or filter as defined above, and

[0183] (b) visualizing by any appropriate means antigen-antibody complexes formed in (a), for example by EIA, ELISA, RIA, or by immunofluorescence.

[0184] According to one advantageous embodiment of said process, step (a) comprises:

[0185] (a₁) bringing said biological sample into contact with at least a first antibody or an antibody fragment which is attached to an appropriate support, in particular a microplate,

[0186] (a₂) washing the solid phase, and

[0187] (a₃) adding at least a second antibody or an antibody fragment, different from the first, said antibody or antibody fragment being optionally appropriately labeled.

[0188] This method, which makes it possible to capture the viral particles present in the biological sample, is also called immunocapture method.

[0189] For example:

[0190] step (a₁) is carried out with at least a first monoclonal or polyclonal antibody or a fragment thereof, directed against the S, M and/or E protein, and/or a peptide corresponding to the ectodomain of one of these proteins (M2-14 or E1-12 peptides)

[0191] step (a₃) is carried out with at least one antibody or an antibody fragment directed against another epitope of the same protein or preferably against another protein, preferably against an inner protein such as the N nucleoprotein or the endodomain of the E or M protein, more preferably still these are antibodies or antibody fragments directed against the N protein which is very abundant in the viral particle; when an antibody or an antibody fragment directed against an inner protein (N) or against the endodomain of the E or M proteins is used, said antibody is incubated in the presence of detergent, such as Tween 20 for example, at concentrations of the order of 0.1%.

[0192] step (b) for visualizing the antigen-antibody complexes formed is carried out, either directly with the aid of a second antibody labeled for example with biotin or an appropriate enzyme such as peroxidase or alkaline phosphatase, or indirectly with the aid of an anti-immunoglobulin serum labeled as above. The complexes thus formed are visualized with the aid of an appropriate substrate.

[0193] According to a preferred embodiment of this aspect of the invention, the biological sample is mixed with the visualizing monoclonal antibody prior to its being brought into contact with the capture monoclonal antibodies. Where appropriate, the serum-visualizing antibody mixture is incubated for at least 10 minutes at room temperature before being applied to the plate.

[0194] The subject of the present invention is also an immunocapture test intended to detect an infection by the SARS-associated coronavirus by detecting the native nucleoprotein (N protein), in particular characterized in that the antibody used for the capture of the native viral nucleoprotein is a monoclonal antibody specific for the central region and/or for a conformational epitope.

[0195] According to one embodiment of said test, the antibody used for the capture of the N protein is the monoclonal antibody mAb87, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3328.

[0196] According to another embodiment of said immunocapture test, the antibody used for the capture of the N protein is the monoclonal antibody mAb86, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3329.

[0197] According to another embodiment of said immunocapture test, the monoclonal antibodies mAb86 and mAb87 are used for the capture of the N protein.

[0198] In the immunocapture tests according to the invention, it is possible to use, for visualizing the N protein, the monoclonal antibody mAb57, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3330, said antibody being conjugated with a visualizing molecule or particle.

[0199] In accordance with said immunocapture test, a combination of the antibodies mAb57 and mAb87, conjugated with a visualizing molecule or particle, is used for the visualization of the N protein.

[0200] A visualizing molecule may be a radioactive atom, a dye, a fluorescent molecule, a fluorophore, an enzyme; a visualizing particle may be for example: colloidal gold, a magnetic particle or a latex bead.

[0201] The subject of the present invention is also a reagent for detecting a SARS-associated coronavirus, characterized in that it is selected from the group consisting of:

[0202] (a) a pair of primers or a probe as defined above,

[0203] (b) a recombinant vector as defined above or a modified cell as defined above,

[0204] (c) an isolated coronavirus strain as defined above or a polynucleotide as defined above,

[0205] (d) an antibody or an antibody fragment as defined above,

[0206] (e) a combination of antibodies comprising the monoclonal antibodies mAb86 and/or mAb87, and the monoclonal antibody mAb57, as defined above,

[0207] (f) a chip or a filter as defined above.

[0208] The subject of the present invention is also a method for the detection of a SARS-associated coronavirus infection, from a biological sample, by indirect IgG ELISA using the N protein, which method is characterized in that the plates are sensitized with an N protein solution at a concentration of between 0.5 and 4 µg/ml, preferably to 2 µg/ml, in a 10 mM PBS buffer pH 7.2, phenol red at 0.25 ml/l.

[0209] The subject of the present invention is additionally a method for the detection of a SARS-associated coronavirus infection, from a biological sample, by double epitope ELSA, characterized in that the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

[0210] According to one variant of the tests for detecting SARS-associated coronaviruses, these tests combine an ELSA using the N protein, and another ELSA using the S protein, as described below.

[0211] The subject of the present invention is also an immune complex formed of a polyclonal or monoclonal antibody or antibody fragment as defined above, and of a SARS-associated coronavirus protein or peptide.

[0212] The subject of the present invention is additionally a SARS-associated coronavirus detection kit, characterized in that it comprises at least one reagent selected from the group consisting of: a pair of primers, a probe, a DNA or RNA chip, a recombinant vector, a modified cell, an isolated coronavirus strain, a polynucleotide, a protein or a peptide, an antibody, and a protein chip as defined above.

[0213] The subject of the present invention is additionally an immunogenic composition, characterized in that it comprises at least one product selected from the group consisting of:

- [0214] a) a protein or a peptide as defined above,
- [0215] b) a polynucleotide of the DNA or RNA type or one of its representative fragments as defined above, having a sequence chosen from:
 - [0216] (i) the sequence SEQ ID NO: 1 or its RNA equivalent
 - [0217] (ii) the sequence hybridizing under high stringency conditions with the sequence SEQ ID NO: 1,
 - [0218] (iii) the sequence complementary to the sequence SEQ ID NO: 1 or to the sequence hybridizing under high stringency conditions with the sequence SEQ ID NO: 1,
 - [0219] (iv) the nucleotide sequence of a representative fragment of the polynucleotide as defined in (i), (ii) or (iii),
 - [0220] (v) the sequence as defined in (i), (ii), (iii) or (iv), modified, and
- [0221] c) a recombinant expression vector comprising a polynucleotide as defined in b), and
- [0222] d) a cDNA library as defined above,

said immunogenic composition being capable of inducing protective humoral or cellular immunity specific for the SARS-associated coronavirus, in particular the production of an antibody directed against a specific epitope of the SARS-associated coronavirus.

[0223] The proteins and peptides as defined above, in particular the S, M, E and/or N proteins and the derived peptides, and the nucleic acid (DNA or RNA) molecules encoding said proteins or said peptides are good candidate vaccines and may be used in immunogenic compositions for the production of a vaccine against the SARS-associated coronavirus.

[0224] According to an advantageous embodiment of the compositions according to the invention, they additionally contain at least one pharmaceutically acceptable vehicle and optionally carrier substances and/or adjuvants.

[0225] The pharmaceutically acceptable vehicles, the carrier substances and the adjuvants are those conventionally used.

[0226] The adjuvants are advantageously chosen from the group consisting of oily emulsions, saponin, mineral substances, bacterial extracts, aluminum hydroxide and squalene.

[0227] The carrier substances are advantageously selected from the group consisting of unilamellar liposomes, multilamellar liposomes, micelles of saponin or solid microspheres of a saccharide or auriferous nature.

[0228] The compositions according to the invention are administered by the general route, in particular by the intramuscular or subcutaneous route or alternatively by the local, in particular nasal (aerosol) route.

[0229] The subject of the present invention is also the use of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75 to form an immune complex with an antibody specifically directed against an epitope of the SARS-associated coronavirus.

[0230] The subject of the present invention is also an immune complex consisting of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75, and of an antibody specifically directed against an epitope of the SARS-associated coronavirus.

[0231] The subject of the present invention is also the use of an isolated or purified protein or peptide having a sequence selected from the group consisting of the sequences SEQ ID NO: 3, 10, 12, 14, 17, 22, 24, 26, 28, 30, 33, 35, 37, 69, 70, 71, 74 and 75 to induce the production of an antibody capable of specifically recognizing an epitope of the SARS-associated coronavirus.

[0232] The subject of the present invention is also the use of an isolated or purified polynucleotide having a sequence selected from the group consisting of the sequences SEQ ID NO: 1, 2, 4, 7, 8, 13, 15, 16, 18, 19, 20, 31, 36 and 38 to induce the production of an antibody directed against the protein encoded by said polynucleotide and capable of specifically recognizing an epitope of the SARS-associated coronavirus.

[0233] The subject of the present invention is also monoclonal antibodies recognizing the native S protein of a SARS-associated coronavirus.

[0234] The subject of the present invention is also the use of a protein or a polypeptide of the S protein family, as defined above, or of an antibody recognizing the native S protein, as defined above, to detect an infection by a SARS-associated coronavirus, in a biological sample.

[0235] The subject of the present invention is also a method for detecting an infection by a SARS-associated coronavirus, in a biological sample, characterized in that the

detection is carried out by ELISA using the recombinant S protein, expressed in a eukaryotic system.

[0236] According to an advantageous embodiment of said method, it is a double epitope ELISA method, and the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

[0237] The subject of the present invention is also an immune complex consisting of a monoclonal antibody or antibody fragment recognizing the native S protein, and of a protein or a peptide of the SARS-associated coronavirus.

[0238] The subject of the present invention is also an immune complex consisting of a protein or a polypeptide of the S protein family, as defined above, and of an antibody specifically directed against an epitope of the SARS-associated coronavirus.

[0239] The subject of the present invention is additionally a SARS-associated coronavirus detection kit or box, characterized in that it comprises at least one reagent selected from the group consisting of: a protein or polypeptide of the S protein family, as defined above, a nucleic acid encoding a protein or peptide of the S protein family, as defined above,

a cell expressing a protein or polypeptide of the S protein family, as defined above, or an antibody recognizing the native S protein of a SARS-associated coronavirus.

[0240] The subject of the present invention is an immunogenic and/or vaccine composition, characterized in that it comprises a polypeptide or a recombinant protein of the S protein family, as defined above, obtained in a eukaryotic expression system.

[0241] The subject of the present invention is also an immunogenic and/or vaccine composition, characterized in that it comprises a vector or recombinant virus, expressing a protein or a polypeptide of the S protein family, as defined above.

[0242] In addition to the preceding features, the invention further comprises other features, which will emerge from the description which follows, which refers to examples of use of the polynucleotide representing the genome of the SARS-CoV strain derived from the sample recorded under the number 031589, and derived cDNA fragments which are the subject of the present invention, and to Table I presenting the sequence listing:

TABLE I

Identification number	Sequence	Sequence listing	
		Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the of the CNCM corresponding plasmid
SEQ ID NO: 1	genome of the strain derived from the sample 031589	—	—
SEQ ID NO: 2	ORF-S*	21406–25348	—
SEQ ID NO: 3	S protein	—	—
SEQ ID NO: 4	ORF-S**	21406–25348	I-3059
SEQ ID NO: 5	Sa fragment	21406–23454	I-3020
SEQ ID NO: 6	Sb fragment	23322–25348	I-3019
SEQ ID NO: 7	ORF-3 + ORF-4*	25110–26244	—
SEQ ID NO: 8	ORF-3 + ORF-4**	25110–26244	I-3126
SEQ ID NO: 9	ORF3	—	—
SEQ ID NO: 10	ORF-3 protein	—	—
SEQ ID NO: 11	ORF4	—	—
SEQ ID NO: 12	ORF-4 protein	—	—
SEQ ID NO: 13	ORF-E*	26082–26413	—
SEQ ID NO: 14	E protein	—	—
SEQ ID NO: 15	ORF-E**	26082–26413	I-3046
SEQ ID NO: 16	ORF-M*	26330–27098	—
SEQ ID NO: 17	M protein	—	—
SEQ ID NO: 18	ORF-M**	26330–27098	I-3047
SEQ ID NO: 19	ORF7 to 11*	26977–28218	—
SEQ ID NO: 20	ORF7 to 11**	26977–28218	I-3125
SEQ ID NO: 21	ORF7	—	—
SEQ ID NO: 22	ORF7 protein	—	—
SEQ ID NO: 23	ORF8	—	—
SEQ ID NO: 24	ORF8 protein	—	—
SEQ ID NO: 25	ORF9	—	—
SEQ ID NO: 26	ORF9 protein	—	—
SEQ ID NO: 27	ORF10	—	—
SEQ ID NO: 28	ORF10 protein	—	—
SEQ ID NO: 29	ORF11	—	—
SEQ ID NO: 30	ORF11 protein	—	—
SEQ ID NO: 31	OrF1ab	265–21485	—
SEQ ID NO: 32	ORF13	28130–28426	—
SEQ ID NO: 33	ORF13 protein	—	—
SEQ ID NO: 34	ORF14	—	—
SEQ ID NO: 35	ORF14 protein	28583–28795	—

TABLE I-continued

Identification number	Sequence	Sequence listing	
		Position of the cDNA with reference to Genbank AY274119.3	Deposit number at the of the CNCM corresponding plasmid
SEQ ID NO: 36	ORF-N*	28054–29430	
SEQ ID NO: 37	N protein	—	—
SEQ ID NO: 38	ORF-N**	28054–29430	I-3048
SEQ ID NO: 39	noncoding 5**	1–204	I-3124
SEQ ID NO: 40	noncoding 3**	28933–29727	I-3123
SEQ ID NO: 41	ORF1ab	30–500	—
	Fragment L0		
SEQ ID NO: 42	Fragment L1	211–2260	—
SEQ ID NO: 43	Fragment L2	2136–4187	—
SEQ ID NO: 44	Fragment L3	3892–5344	—
SEQ ID NO: 45	Fragment L4b	4932–6043	—
SEQ ID NO: 46	Fragment L4	5305–7318	—
SEQ ID NO: 47	Fragment L5	7275–9176	—
SEQ ID NO: 48	Fragment L6	9032–11086	—
SEQ ID NO: 49	Fragment L7	10298–12982	—
SEQ ID NO: 50	Fragment L8	12815–14854	—
SEQ ID NO: 51	Fragment L9	14745–16646	—
SEQ ID NO: 52	Fragment L10	16514–18590	—
SEQ ID NO: 53	Fragment L11	18500–20602	—
SEQ ID NO: 54	Fragment L12	20319–22224	—
SEQ ID NO: 55	Sense N primer	—	—
SEQ ID NO: 56	Antisense N primer	—	—
SEQ ID NO: 57	Sense S _C primer	—	—
SEQ ID NO: 58	Sence S _L primer	—	—
SEQ ID NO: 59	Antisense S _C and S _L primer	—	—
SEQ ID NO: 60	Sense primer series 1	28507–28522	—
SEQ ID NO: 61	Antisense primer series 1	28774–28759	—
SEQ ID NO: 62	Sense primer series 2	28375–28390	—
SEQ ID NO: 63	Antisense primer series 2	28702–28687	—
SEQ ID NO: 64	Probe 1/series 1	28561–28586	—
SEQ ID NO: 65	Probe 2/series 1	28588–28608	—
SEQ ID NO: 66	Probe 1/series 2	28541–28563	—
SEQ ID NO: 67	Probe 2/series 2	28565–28589	—
SEQ ID NO: 68	Anchor primer 14T		
SEQ ID NO: 69	Peptide M2–14	—	—
SEQ ID NO: 70	Peptide E1–12	—	—
SEQ ID NO: 71	Peptide E53–76	—	—
SEQ ID NO: 72	Noncoding 5*	1–204	—
SEQ ID NO: 73	Noncoding 3*	28933–29727	—
SEQ ID NO: 74	ORF1a protein	—	—
SEQ ID NO: 75	ORF1b protein	—	—
SEQ ID NO: 76–139	Primers		
SEQ ID NO: 140	Pseudogene of S		
SEQ ID NO: 141–148	Primers		
SEQ ID NO: 149	Aa1–13 of S		
SEQ ID NO: 150	Polypeptide		
SEQ ID NO: 151–158	Primers		

* PCR amplification product (amplicon)

** Insert cloned into the plasmid deposited at the CNCM and to the appended drawings in which:

[0243] FIG. 1 illustrates Western-blot analysis of the expression in vitro of the recombinant proteins N, S_C and S_L from the expression vectors pIVEX. Lane 1: pIV2.3N. Lane 2: pIV2.3S_C. Lane 3: pIV2.3S_L. Lane 4: pIV2.4N. Lane 5: pIV2.4S_L or pIV2.4S_C. Lane 6: pIV2.4S_L. The expression of the GFP protein expressed from the same vector is used as a control.

[0244] FIG. 2 illustrates the analysis, by polyacrylamide gel electrophoresis under denaturing conditions (SDS-PAGE) and staining with Coomassie blue, of the expression in vivo of the N protein from the expression vectors pIVEX. The *E. coli* BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX is cultured at 30° C. in LB medium, in the presence or in the absence of inducer (IPTG 1 mM). Lane 1: pIV2.3N. Lane 2: pIV2.4N.

[0245] FIG. 3 illustrates the analysis, by polyacrylamide gel electrophoresis under denaturing conditions (SDS-

PAGE) and staining with Coomassie blue, of the expression in vivo of the S_L and S_C polypeptides from the expression vectors pIVEX. The *E. coli* BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX is cultured at 30° C. in LB medium, in the presence or in the absence of inducer (IPTG 1 mM). Lane 1: pIV2.3 S_C . Lane 2: pIV2.3 S_L . Lane 3: pIV2.4 S_L . Lane 4: pIV2.4 S_L .

[0246] FIG. 4 illustrates the antigenic activity of the recombinant N, S_L and S_C proteins produced in the *E. coli* BL21(DE3)pDIA17 strain transformed with the recombinant vectors pIVEX. A: electrophoresis (SDS-PAGE) of the bacterial lysates. B and C: Western-blot with the sera, obtained from the same patient infected with SARS-CoV, collected 6 days (B: serum M12) and 29 days (C: serum M13) respectively after the onset of the SARS symptoms. Lane 1: pIV2.3N. Lane 2: pIV2.4N. Lane 3: pIV2.3 S_C . Lane 4: pIV2.4 S_L . Lane 5: pIV2.3 S_L . Lane 6: pIV2.4 S_L .

[0247] FIG. 5 illustrates the purification on an Ni-NTA agarose column of the recombinant N protein produced in the *E. coli* BL21(DE3)pDIA17 strain from the vector pIV2.3N. Lane 1: total bacterial extract. Lane 2: soluble extract. Lane 3: insoluble extract. Lane 4: extract deposited on the Ni-NTA column. Lane 5: unbound proteins. Lane 6: fractions of peak 1. Lane 7: fractions of peak 2.

[0248] FIG. 6 illustrates the purification of the recombinant S_C protein from the inclusion bodies produced in the *E. coli* BL21(DE3)pDIA17 strain transformed with pIV2.4 S_L . A: Treatment with Triton X-100 (2%): Lane 1: total bacterial extract. Lane 2: soluble extract. Lane 3: insoluble extract. Lane 4: supernatant after treatment with Triton X-100 (2%). Lanes 5 and 6: pellet after treatment with Triton X-100 (2%). B: Treatment with 4 M, 5 M, 6 M and 7 M urea of the soluble and insoluble extracts.

[0249] FIG. 7 represents the immunoblot produced with the aid of a lysate of cells infected with SARS-CoV and a serum from a patient suffering from atypical pneumopathy.

[0250] FIG. 8 represents immunoblots produced with the aid of a lysate of cells infected with SARS-CoV and rabbit immunosera specific for the nucleoprotein N (A) and for the spicule protein S (B). I.S.: immune serum. p.i.: preimmune serum. The anti-N immune serum was used at 1/50 000 and the anti-S immune serum at 1/10 000.

[0251] FIG. 9 illustrates the ELISA reactivity of the rabbit monospecific polyclonal sera directed against the N protein or the short fragment of the S protein (S_C), toward the corresponding recombinant proteins used for immunization. A: rabbits P13097, P13081 and P13031 immunized with the purified recombinant N protein. B: rabbits P11135, P13042 and P14001 immunized with a preparation of inclusion bodies corresponding to the short fragment of the S protein (S_C). I.S.: immune serum. p.i.: preimmune serum.

[0252] FIG. 10 illustrates the ELISA reactivity of the purified recombinant N protein, toward sera from patients suffering from atypical pneumonia caused by SARS-CoV. FIG. 10a: ELISA plates prepared with the N protein at the concentration of 4 µg/ml and 2 µg/ml. FIG. 10b: ELISA plate prepared with the N protein at the concentration of 1 µg/ml. The sera designated A, B, D, E, F, G, H correspond to those of Table IV.

[0253] FIG. 11 illustrates the amplification by RT-PCR of decreasing quantities of synthetic RNA of the SARS-CoV N

gene (10^7 to 1 copy), with the aid of pairs of primers No. 1 (N/+28507, N/-28774) (A) and No. 2 (N/+28373, N/-28702) (B). T: amplification performed in the absence of RNA. MW: DNA marker.

[0254] FIG. 12 illustrates the amplification by RT-PCR in real time of synthetic RNA for the SARS-CoV N gene: decreasing quantities of synthetic RNA as replica (repli.; lanes 16 to 29) and of viral RNA diluted $1/20 \times 10^4$ (lane 32) were amplified by RT-PCR in real time with the aid of the kit "Light Cycler RNA Amplification Kit Hybridization Probes" and pairs of primers and probes of the No. 2 series, under the conditions described in Example 8.

[0255] FIG. 13 (FIGS. 13.1 to 13.7) represents the restriction map of the sequence SEQ ID NO: 1 corresponding to the DNA equivalent of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589.

[0256] FIG. 14 shows the result of the SARS serology test by indirect N ELISA (1st series of sera tested).

[0257] FIG. 15 shows the result of the SARS serology test by indirect N ELISA (2nd series of sera-tested).

[0258] FIG. 16 presents the result of the SARS serology test by double epitope N ELISA (1st series of sera tested).

[0259] FIG. 17 shows the result of the SARS serology test by double epitope N ELISA (2nd series of sera tested).

[0260] FIG. 18 illustrates the test of reactivity of the anti-N monoclonal antibodies by ELISA on the native nucleoprotein N of SARS-CoV. The antibodies were tested in the form of hybridoma culture supernatants by indirect ELISA using an irradiated lysate of VeroE6 cells infected with SARS-CoV as antigen (SARS lysate curves). A negative control for reactivity is performed for each antibody on a lysate of uninfected VeroE6 cells (negative lysate curves). Several monoclonal antibodies of known specificity were used as negative control antibodies: para1-3 directed against the antigens of the parainfluenza viruses type 1-3 (Bio-Rad) and influenza B directed against the antigens of the influenza virus type B (Bio-Rad).

[0261] FIG. 19 illustrates the test of reactivity of the anti-N of SARS-CoV monoclonal antibodies by ELISA on the native antigens of the human coronavirus 229E (HCoV-229E). The antibodies were tested in the form of hybridoma culture supernatants by an indirect ELISA test using a lysate of MRC-5 cells infected with the human coronavirus 229E as antigen (229E lysate curves). A negative control for immunoreactivity was performed for each antibody on a lysate of noninfected MRC-5 cells (negative lysate curves). The monoclonal antibody 5-11H.6 directed against the S protein of the human coronavirus 229E (Sizun et al. 1998, J. Virol. Met. 72: 145-152) is used as positive control antibody. The antibodies para1-3 directed against the antigens of the parainfluenza virus type 1-3 (Bio-Rad) and influenza B directed against the antigens of the influenza virus type B (Bio-Rad) were added to the panel of monoclonal antibodies tested.

[0262] FIG. 20 shows a test of reactivity of the anti-N of SARS-CoV monoclonal antibodies by Western blotting on the denatured native nucleoprotein N of SARS-CoV. A lysate of VeroE6 cells infected with SARS-CoV was prepared in the loading buffer according to Laemmli and caused to migrate in a 12% SDS polyacrylamide gel and then the

proteins were transferred onto PVDF membrane. The anti-N monoclonal antibodies tested were used for the immunoassay at the concentration of 0.05 µg/ml. The visualization is carried out with anti-mouse IgG(H+L) antibodies coupled to peroxidase (NA93IV, Amersham) and the ECL+ system. Two monoclonal antibodies were used as negative controls for reactivity: influenza B directed against the antigens of the influenza virus type B (Bio-Rad) and para1-3 directed against the antigens of the parainfluenza virus type 1-3 (Bio-Rad).

[0263] FIG. 21 presents the plasmids for expression in mammalian cells of the SARS-CoV S protein. The cDNA for the SARS-CoV S was inserted between the BamHI and XhoI sites of the expression plasmid pcDNA3.1(+) (Clontech) in order to obtain the plasmid pcDNA-S and between the NheI and XhoI sites of the expression plasmid pCI (Promega) in order to obtain the plasmid pCI-S. The WPRE and CTE sequences were inserted between each of the two plasmids pcDNA-S and pCI-S between the XhoI and XbaI sites in order to obtain the plasmids pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S-CTE and pCI-S-WPRE, respectively.

[0264] SP: signal peptide predicted (aa 1-13) with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10: 1-6)

[0265] TM: transmembrane region predicted (aa 1196-1218) with the software TMHMM v2.0 (Sonnhammer et al., 1998, Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, pp. 175-182, AAAI Press). It should be noted that the amino acids W-1194 and P1195 are possibly part of the transmembrane region with the respective probabilities of 0.13 and 0.42

[0266] P-CMV: cytomegalovirus immediate/early promoter. BGH pA: polyadenylation signal of the bovine growth hormone gene

[0267] SV40 late pA: SV40 virus late polyadenylation signal

[0268] SD/SA: splice donor and acceptor sites

[0269] WPRE: sequences of the "Woodchuck Hepatitis Virus posttranscriptional regulatory element" of the woodchuck hepatitis virus

[0270] CTE: sequences of the "constitutive transport element" of the Mason-Pfizer simian retrovirus

[0271] FIG. 22 illustrates the expression of the S protein after transfection of VeroE6 cells. Cellular extracts were prepared 48 hours after transfection of VeroE6 cells with the plasmids pcDNA, pcDNA-S, pCI and pCI-S. Cellular extracts were also prepared 18 hours after infection with the recombinant vaccinia virus VV-TF7.3 and transfection with the plasmids pcDNA or pcDNA-S. As a control, extracts of VeroE6 cells were prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 3. They were separated on an 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). A molecular mass ladder (kDa) is presented in the figure.

[0272] SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

[0273] Mock: control extract of noninfected cells

[0274] FIG. 23 illustrates the effect of the CTE and WPRE sequences on the expression of the S protein after transfection of VeroE6 and 293T cells. Cellular extracts were prepared 48 hours after transfection of VeroE6 cells (A) or 293T cells (B) with the plasmids pcDNA, pcDNA-S, pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S, pCI-S-CTE and pCI-S-WPRE separated on 8% SDS polyacrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). A molecular mass ladder (kDa) is presented in the figure.

[0275] SARS-CoV: extract of VeroE6 cells prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 3.

[0276] Mock: control extract of noninfected VeroE6 cells

[0277] FIG. 24 presents defective lentiviral vectors with central DNA flap for the expression of SARS-CoV S. The cDNA for the SARS-CoV S protein was cloned in the form of a BamHI-XhoI fragment into the plasmid pTRIPΔU3-CMV containing a defective lentiviral vector TRIP with central DNA flap (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmid pTRIP-S. The optimum expression cassettes consisting of the CMV virus immediate/early promoter, a splice signal, cDNA for S and either of the posttranscriptional signals CTE or WPRE were substituted for the cassette EF1α-EGFP of the defective lentiviral expression vector with central DNA flap TRIPΔU3-EF1α (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmids pTRIP-SD/SA-S-CTE and pTRIP-SD/SA-S-WPRE.

[0278] SP: signal peptide

[0279] TM: transmembrane region

[0280] P-CMV: cytomegalovirus immediate/early promoter

[0281] P-EF1α: EF1α gene promoter

[0282] SD/SA: splice donor and acceptor sites

[0283] WPRE: sequences of the "Woodchuck Hepatitis Virus posttranscriptional regulatory element" of the woodchuck hepatitis virus

[0284] CTE: sequences of the "constitutive transport element" of the Mason-Pfizer simian retrovirus

[0285] LTR: long terminal repeat

[0286] ΔU3: LTR deleted for the "promoter/enhancer" sequences

[0287] cPPT: "polypurine tract cis-active sequence"

[0288] CTS: "central termination sequence"

[0289] FIG. 25 shows the Western-blot analysis of the expression of the SARS-CoV S by cell lines transduced with the lentiviral vectors TRIP-SD/SA-S-WPRE and TRIP-SD/SA-S-CTE. Cellular extracts were prepared from established lines FrhK4-S-CTE and FrhK4-S-WPRE after transduction with the lentiviral vectors TRIP-SD/SA-S-CTE and TRIP-SD/SA-S-WPRE respectively. They were separated on an 8% SDS acrylamide gel and analyzed by Western blotting

with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) conjugate coupled to peroxidase. A molecular mass ladder (kDa) is presented in the figure.

[0290] T-: control extract of FrhK-4 cells

[0291] T+: extract of FrhK-4 cells prepared 24 hours after infection with SARS-CoV at a multiplicity of infection of 3.

[0292] FIG. 26 relates to the analysis of the expression of Ssol polypeptide by cell lines transduced with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. The secretion of the Ssol polypeptide was determined in the supernatant of a series of cell clones isolated after transduction of FrhK-4 cells with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. 5 μ l of supernatant, diluted 1/2 in loading buffer according to Laemmli, were analyzed by Western blotting, visualized with an anti-FLAG monoclonal antibody (M2, Sigma) and an anti-mouse IgG(H+L) conjugate coupled to peroxidase. T-: supernatant of the parental FRhK-4 line. T+: supernatant of BHK cells infected with a recombinant vaccinia virus expressing the Ssol polypeptide. The solid arrow indicates the Ssol polypeptide, while the empty arrow indicates a cross reaction with a protein of cellular origin.

[0293] FIG. 27 shows the results relating to the analysis of the purified Ssol polypeptide

[0294] A. 8, 2, 0.5 and 0.125 μ g of recombinant Ssol polypeptide purified by anti-FLAG affinity chromatography and gel filtration (G75) were separated on 8% SDS polyacrylamide gel. The Ssol polypeptide and variable quantities of molecular mass markers (MM) were visualized by staining with silver nitrate (Gelcode SilverSNAP stain kit II, Pierce).

[0295] B. Standard markers for analysis by SELDI-TOF mass spectrometry

[0296] IgG: bovine IgG of MM 147300

[0297] ConA: conalbumin of MM 77490

[0298] HRP: horseradish peroxidase analyzed as a control and of MM 43240

[0299] C. Analysis by mass spectrometry (SELDI-TOF) of the recombinant Ssol polypeptide.

[0300] The peaks A and B correspond to the single and double charged Ssol polypeptide.

[0301] D. Sequencing of the N-terminal end of the recombinant Ssol polypeptide. 5 Edman degradation cycles in liquid phase were carried out on an ABI494 sequencer (Applied Biosystems).

[0302] FIG. 28 illustrates the influence of a splicing signal and of the CTE and WPRE sequences on the efficacy of the gene immunization with the aid of plasmid DNA encoding the SARS-CoV S

[0303] A. Groups of 7 BALB/c mice were immunized twice at 4 weeks' interval with the aid of 50 μ g of plasmid DNA of pCI, pcDNA-S, pCI-S, pcDNA-N and pCI-HA.

[0304] B. Groups of 6 BALB/c mice were immunized twice at 4 weeks' interval with the aid of 2 μ g, 10 μ g or 50 μ g of plasmid DNA of pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE.

[0305] The immune sera collected 3 weeks after the second immunization were analyzed by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (NA931V, Amersham) and TMB (KPL).

[0306] FIG. 29 shows the seroneutralization of the infectivity of SARS-CoV with the antibodies induced in mice after gene immunization with the aid of plasmid DNA encoding SARS-CoV S. Pools of immune sera collected 3 weeks after the second immunization were prepared for each of the groups of experiments described in FIG. 28 and evaluated for their capacity to seroneutralize the infectivity of 100 TCID₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

[0307] A. Groups by BALB/c mice immunized twice at 4 weeks' interval with the aid of 50 μ g of plasmid DNA of pCI, pcDNA-S, pCI-S, pcDNA-N and pCI-HA. □: pre-immune serum. ■: immune serum.

[0308] B. Groups of BALB/c mice immunized twice at 4 weeks' interval with the aid of 2 μ g, 10 μ g or 50 μ g of plasmid DNA of pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE.

[0309] FIG. 30 illustrates the immunoreactivity of the recombinant Ssol polypeptide toward sera from patients suffering from SARS. The reactivity of sera from patients was analyzed by indirect ELISA test against solid phases prepared with the aid of the purified recombinant Ssol polypeptide. The antibodies from patients reacting with the solid phase at a dilution of 1/400 are visualized with a human anti-IgG(H+L) polyclonal antibody coupled to peroxidase (Amersham NA933V) and TMB plus, H202 (KPL). The sera of probable SARS cases are identified by a National Reference Center for Influenza Viruses serial number and by the initials of the patient and the number of days elapsed since the onset of symptoms, where appropriate. The TV sera are control sera from subjects which were collected in France before the SARS epidemic which occurred in 2003.

[0310] FIG. 31 shows the induction of antibodies directed against SARS-CoV after immunization with the recombinant Ssol polypeptide. Two groups of 6 mice were immunized at 3 weeks' interval with 10 μ g of recombinant Ssol polypeptide (Ssol group) adjuvanted with aluminum hydroxide or, as a control, of adjuvant alone (mock group). Three successive immunizations were performed and the immune sera were collected 3 weeks after each of the three immunizations (IS1, IS2, IS3). The immune sera were analyzed per pool for each of the 2 groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (Amersham) and TMB (KPL).

[0311] FIG. 32 presents the nucleotide alignment of the sequences of the synthetic gene 040530 with the sequence of the wild-type gene of the SARS-CoV isolate 031589. I-3059

corresponds to nucleotides 21406-25348 of the SARS-CoV isolate 031589 deposited at the C.N.C.M. under the number I-3059 (SEQ ID NO: 4, plasmid pSARS-S) S-040530 is the sequence of the synthetic gene 040530.

[0312] FIG. 33 illustrates the use of a synthetic gene for the expression of the SARS-CoV S. Cellular extracts prepared 48 hours after transfection of VeroE6 cells (A) or 293T cells (B) with the plasmids pCI, PCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-Ssynth were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The levels of expression of the S protein were measured by quantifying the 2 predominant bands identified on the image.

[0313] FIG. 34 presents a diagram for the construction of recombinant vaccinia viruses VV-TG-S, VV-TG-Ssol, VV-TN-S and VV-TN-Ssol

[0314] A. The cDNAs for the S protein and the Ssol polypeptide of SARS-CoV were inserted between the BamHI and SmaI sites of the transfer plasmid pTG186 in order to obtain the plasmids pTG-S and pTG-Ssol.

[0315] B. The sequences of the synthetic promoter 480 were then substituted for those of the 7.5 promoter by exchange of the NdeI-PstI fragments of the plasmids pTG186poly, pTG-S and pTG-Ssol in order to obtain the transfer plasmids pTN480, pTN-S and pTN-Ssol.

[0316] C. Sequence of the synthetic promoter 480 as contained between the NdeI and PstI sites of the transfer plasmids of the pTN series. An AscI site was inserted in order to facilitate subsequent handling. The restriction sites and the promoter sequence are underlined.

[0317] D. The recombinant vaccinia viruses are obtained by double homologous recombination in vivo between the TK cassette of the transfer plasmids of the pTG and pTN series and the TK gene of the Copenhagen strain of the vaccinia virus.

[0318] SP: signal peptide predicted (aa 1-13) with the software signalP v2.0 (Nielsen et al., 1997, Protein Engineering, 10: 1-6)

[0319] TM: transmembrane region predicted (aa 1196-1218) with the software TMHMM v2.0 (Sonnhammer et al., 1998, Proc. of Sixth Int. Conf. on Intelligent Systems for Molecular Biology, pp. 175-182, AAAI Press). It should be noted that the amino acids W1194 and P1195 possibly form part of the transmembrane region with respective probabilities of 0.13 and 0.42.

[0320] TK-L, TK-R: left- and right-hand parts of the vaccinia virus thymidine kinase gene

[0321] MCS: multiple cloning site

[0322] PE: early promoter

[0323] PL: late promoter

[0324] PL synth: synthetic late promoter 480

[0325] FIG. 35 illustrates the expression of the S protein by recombinant vaccinia viruses, analyzed by Western blot-

ting. Cellular extracts were prepared 18 hours after infection of CV1 cells with the recombinant vaccinia viruses VV-TG, VV-TG-S and VV-TN-S at an M.O.I. of 2 (A). As a control, extracts of VeroE6 cells were prepared 8 hours after infection with SARS-CoV at a multiplicity of infection of 2. Cellular extracts were also prepared 18 hours after infection of CV1 cells with the recombinant vaccinia viruses VV-TG-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol (B). They were separated on 8% SDS acrylamide gels and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham). "1 μ l" and "10 μ l" indicates the quantities of cellular extracts deposited on the gel. A molecular mass ladder (kDa) is presented in the figure.

[0326] SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

[0327] Mock: control extract of noninfected cells

[0328] FIG. 36 shows the result of a Western-blot analysis of the secretion of the Ssol polypeptide by the recombinant vaccinia viruses.

[0329] A. Supernatants of CV1 cells infected with the recombinant vaccinia virus VV-TN, various clones of the VV-TN-Ssol virus and with the viruses VV-TG-Ssol or VV-TN-Sflag were harvested 18 hours after infection of CV1 cells at an M.O.I. of 2.

[0330] B. Supernatants of 293T, FRhK-4, BHK-21 and CV1 cells infected in duplicate (1.2) with the recombinant vaccinia virus VV-TN-Ssol at an M.O.I. of 2 were harvested 18 hours after infection. The supernatant of CV1 cells infected with the virus VV-TN was also harvested as a control (M).

[0331] All the supernatants were separated on 8% SDS acrylamide gel according to Laemmli and analyzed by Western blotting with the aid of an anti-FLAG mouse monoclonal antibody and an anti-mouse IgG(H+L) polyclonal antibody coupled to peroxidase (NA931V, Amersham) (A) or with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham) (B).

[0332] A molecular mass ladder (kDa) is presented in the figure.

[0333] FIG. 37 shows the analysis of the Ssol polypeptide, purified on SDS polyacrylamide gel

[0334] 10, 5 and 2 μ l of recombinant Ssol polypeptide purified by anti-FLAG affinity chromatography were separated on 4 to 15% gradient SDS polyacrylamide gel. The Ssol polypeptide and variable quantities of molecular mass markers (MM) were visualized by staining with silver nitrate (Gelcode SilverSNAP stain kit II, Pierce).

[0335] FIG. 38 illustrates the immunoreactivity of the recombinant Ssol polypeptide produced by the recombinant vaccinia virus VV-TN-Ssol toward sera of patients suffering from SARS. The reactivity of sera from patients was analyzed by indirect ELISA test against solid phases prepared with the aid of the purified recombinant Ssol polypeptide. The antibodies from patients reacting with the solid phase at a dilution of 1/100 and 1/400 are visualized with a human anti-IgG(H+L) polyclonal antibody coupled to peroxidase

(Amersham NA933V) and TMB plus H2O2 (KPL). The sera of probable SARS cases are identified by a National Reference Center for Influenza Virus serial number and by the initials of the patient and the number of days elapsed since the onset of symptoms, where appropriate. The TV sera are control sera from subjects which were collected in France before the SARS epidemic which occurred in 2003.

[0336] FIG. 39 shows the anti-SARS-CoV antibody response in mice after immunization with the recombinant vaccinia viruses. Groups of 7 BALB/c mice were immunized by the i.v. route twice at 4 weeks' interval with 106 pfu of recombinant vaccinia viruses VV-TG, VV-TG-HA, VV-TG-S, W-TG-Ssol, VV-TN, VV-TN-S, VV-TN-Ssol.

[0337] A. Pools of immune sera collected 3 weeks after each of the two immunizations were prepared for each of the groups and were analyzed by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG polyclonal antibody coupled to peroxidase (NA931V, Amersham) and TMB (KPL).

[0338] B. The pools of immune sera were evaluated for their capacity to seroneutralize the infectivity of 100 TCID50 of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.

[0339] FIG. 40 describes the construction of the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol.

[0340] A. The measles vector is a complete genome of the Schwarz vaccine strain of the measles virus (MV) into which an additional transcription unit has been introduced (Combredet, 2003, Journal of Virology, 77: 11546-11554). The expression of the additional open reading frames (ORF) is controlled by cis-acting elements necessary for the transcription, for the formation of the cap and for the polyadenylation of the transgene which were copied from the elements present at the N/P junction. 2 different vectors allow the insertion between the P (phosphoprotein) and M (matrix) genes on the one hand and the H (hemagglutinin) and L (polymerase) genes on the other hand.

[0341] B. The recombinant genomes MVSchw2-SARS-S and MVSchw2-SARS-Ssol of the measles virus were constructed by inserting the ORFs of the S protein and of the Ssol polypeptide into an additional transcription unit located between the P and M genes of the vector.

[0342] The various genes of the measles virus (MV) are indicated: N (nucleoprotein), PVC (V/C phosphoprotein and protein), M (matrix), F (fusion), H (hemagglutinin), L (polymerase). T7=T7 RNA polymerase promoter, hh=hammerhead ribozyme, T7=T7 phage RNA polymerase terminator sequence, δ =ribozyme of the hepatitis δ virus, (2), (3)=additional transcription units (ATU).

[0343] Size of the MV genome: 15 894 nt.

[0344] SP: signal peptide

[0345] TM: transmembrane region

[0346] FLAG: FLAG tag

[0347] FIG. 41 illustrates the expression of the S protein by the recombinant measles viruses, analyzed by Western blotting.

[0348] Cytoplasmic extracts were prepared after infection of Vero cells by different passages of the viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and the wild-type virus MWSchw as control. Cellular extracts in loading buffer according to Laemmli were also prepared 8 hours after infection of VeroE6 cells with SARS-CoV at a multiplicity of infection of 3. They were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H+L) polyclonal antibody coupled to peroxidase (NA934V, Amersham).

[0349] A molecular mass ladder (kDa) is presented in the figure.

[0350] Pn: nth passage of the virus after coculture of 293-3-46 and Vero cells

[0351] SARS-CoV: extract of VeroE6 cells infected with SARS-CoV

[0352] Mock: control extract of noninfected VeroE6 cells

[0353] FIG. 42 shows the expression of the S protein by the recombinant measles viruses, analyzed by immunofluorescence

[0354] Vero cells in monolayers on glass slides were infected with the wild-type virus MWSchw (A) or the viruses MVSchw2-SARS-S (B) and MVSchw2-SARS-Ssol (C). When the syncytia have reached 30 to 40% confluence (A., B.) or 90-100% (C), the cells were fixed, permeabilized and labeled with anti-SARS-CoV rabbit polyclonal antibodies and an anti-rabbit IgG(H+L) conjugate coupled to FITC (Jackson).

[0355] FIG. 43 illustrates the Western-blot analysis of the immunoreactivity of rabbit sera directed against the peptides E1-12, E53-76 and M2-14. The rabbit 20047 was immunized with the peptide E1-12 coupled to KLH. The rabbits 22234 and 22240 were immunized with the peptide E53-76 coupled to KLH. The rabbits 20013 and 20080 were immunized with the peptide M2-14 coupled to KLH. The immune sera were analyzed by Western blotting with the aid of extracts of cells infected with SARS-CoV (B) or with the aid of extracts of cells infected with a recombinant vaccinia virus expressing the protein E (A) or M (C) of the SARS-CoV 031589 isolate. The immunoblots were visualized with the aid of an anti-rabbit IgG(H+L) conjugate coupled to peroxidase (NA934V, Amersham).

[0356] The position of the E and M proteins is indicated by an arrow.

[0357] A molecular mass ladder (kDa) is presented in the figure.

[0358] It should be understood, however, that these examples are given solely by way of illustration of the subject of the invention, and do not constitute in any manner a limitation thereto.

EXAMPLE 1

Cloning and Sequencing of the Genome of the
SARS-CoV Strain Derived from the Sample
Recorded Under the Number 031589

[0359] The RNA of the SARS-CoV strain was extracted from the sample of bronchoalveolar washing recorded under the number 031589, performed on a patient at the Hanoi (Vietnam) French hospital suffering from SARS.

[0360] The isolated RNA was used as template to amplify the cDNAs corresponding to the various open reading frames of the genome (ORF1a, ORF1b, ORF-S, ORF-E, ORF-M, ORF-N (including ORF-13 and ORF-14), ORF3, ORF4, ORF7 to ORF11), and at the noncoding 5' and 3' ends. The sequences of the primers and of the probes used for the amplification/detection were defined based on the available SARS-CoV nucleotide sequence.

[0361] In the text which follows, the primers and the probes are identified by: the letter S, followed by a letter which indicates the corresponding region of the genome (L for the 5' end including ORF1a and ORF1b; S, M and N for ORF-S, ORF-M, ORF-N, SE and MN for the corresponding intergene regions), and then optionally by Fn, Rn, with n between 1 and 6 corresponding to the primers used for the nested PCR (F1+R1 pair for the first amplification, F2+R2 pair for the second amplification, and the like), and then by +/- or -/- corresponding to a sense or antisense primer and finally by the positions of the primers with reference to the Genbank sequence AY27411.3; for the sense and antisense S and N primers and the other sense primers only, when a single position is indicated, it corresponds to that of the 5' end of a probe or of a primer of about 20 bases; for the antisense primers other than the S and N primers, when a single position is indicated, it corresponds to that of the 3' end of a probe or of a primer of about 20 bases.

[0362] The amplification products thus generated were sequenced with the aid of specific primers in order to determine the complete sequence of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589. These amplification products, with the exception of those corresponding to ORF1a and ORF1b, were then cloned into expression vectors in order to produce the corresponding viral proteins and the antibodies directed against these proteins, in particular by DNA-based immunization.

1. Extraction of the RNAs

[0363] The RNAs were extracted with the aid of the QIamp viral RNA extraction mini kit (QIAGEN) according to the manufacturer's recommendations. More specifically: 140 μ l of the sample and 560 μ l of AVL buffer were vigorously mixed for 15 seconds, incubated for 10 minutes at room temperature and then briefly centrifuged at maximum speed. 560 μ l of 100% ethanol were added to the supernatant and the mixture thus obtained was very vigorously stirred for 15 sec. 630 μ l of the mixture were then deposited on the column.

[0364] The column was placed on a 2 ml tube, centrifuged for 1 min at 8000 rpm, and then the remainder of the preceding mixture was deposited on the same column, centrifuged again, for 1 min at 8000 rpm, and the column was transferred over a clean 2 ml tube. Next, 500 μ l of AW1

buffer were added to the column, and then the column was centrifuged for 1 min at 8000 rpm and the eluate was discarded. 500 μ l of AW2 buffer were added to the column which was then centrifuged for 3 min at 14 000 rpm and transferred onto a 1.5 ml tube. Finally, 60 μ l of AVE buffer were added to the column which was incubated for 1 to 2 min at room temperature and then centrifuged for 1 min at 8000 rpm. The eluate corresponding to the purified RNA was recovered and frozen at -20° C.

2. Amplification, Sequencing and Cloning of the cDNAs

2.1) cDNA Encoding the S Protein

[0365] The RNAs extracted from the sample were subjected to reverse transcription with the aid of random sequence hexameric oligonucleotides (pdN6), so as to produce cDNA fragments.

[0366] The sequence encoding the SARS-CoV S glycoprotein was amplified in the form of two overlapping DNA fragments: 5' fragment (SARS-Sa, SEQ ID NO: 5) and 3' fragment (SARS-Sb, SEQ ID NO: 6), by carrying out two successive amplifications with the aid of nested primers. The amplicons thus obtained were sequenced, cloned into the PCR plasmid vector 2.1-TOPO™ (INVITROGEN), and then the sequence of the cloned cDNAs was determined.

a) Cloning and Sequencing of the Sa and Sb Fragments

a.1) Synthesis of the cDNA

[0367] The reaction mixture containing: RNA (5 μ l), H_2O for injection (3.5 μ l), 5 $\times$ reverse transcriptase buffer (4 μ l), 5 mM dNTP (2 μ l), pdN6 100 μ g/ml (4 μ l), RNasin 40 IU/ μ l (0.5 μ l) and reverse transcriptase AMV-RT, 10 IU/ μ l, PROMEGA (1 μ l) was incubated in a thermocycler under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and then the cDNA obtained was kept at $+4^{\circ}$ C.

a.2) First PCR Amplification

[0368] The 5' and 3' ends of the S gene were respectively amplified with the pairs of primers S/F1/+21350-21372 and S/R1/-23518-23498, S/F3/+23258-23277 and S/R3/-25382-25363. The 50 μ l reaction mixture containing: cDNA (2 μ l), 50 μ M primers (0.5 μ l), 10 $\times$ buffer (5 μ l), 5 mM dNTP (2 μ l), Taq Expand High Fidelity, Roche (0.75 μ l) and H_2O (39, 75 μ l) was amplified in a thermocycler, under the following conditions: an initial step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising: a step of denaturation at 94° C. for 30 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 2 min 30 sec, with 10 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 5 min.

a.3) Second PCR Amplification

[0369] The products of the first PCR amplification (5' and 3' amplicons) were subjected to a second PCR amplification step (nested PCR) under conditions identical to those of the first amplification, with the pairs of primers S/F2/+21406-21426 and S/R2/-23454-23435 and S/F4/+23322-23341 and S/R4/-25348-25329, respectively for the 5' amplicon and the 3' amplicon.

a.4) Cloning and Sequencing of the Sa and Sb Fragments

[0370] The Sa (5' end) and Sb (3' end) amplicons thus obtained were purified with the aid of the QIAquick PCR

purification kit (QIAGEN), following the manufacturer's instructions, and then they were cloned into the vector PCR2.1-TOPO (Invitrogen kit), to give the plasmids called SARS-S1 and SARS-S2.

[0371] The DNA of the Sa and Sb clones was isolated and then the corresponding insert was sequenced with the aid of the Big Dye kit, Applied Biosystem® and universal primers M13 forward and M13 reverse, and primers: S/S/+21867, S/S/+22353, S/S/+22811, S/S/+23754, S/S/+24207, S/S/+24699, S/S/+24348, S/S/-24209, S/S/-23630, S/S/-23038, S/S/-22454, S/S/-21815, S/S/-24784, S/S/+21556, S/S/+23130 and S/S/+24465 following the manufacturer's instructions; the sequences of the Sa and Sb fragments thus obtained correspond to the sequences SEQ ID NO: 5 and SEQ ID NO: 6 in the sequence listing appended as an annex.

[0372] The plasmid, called SARS-S1, was deposited under the No. I-3020, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 5' fragment of the sequence of the S gene of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment called Sa corresponding to the nucleotides at positions 21406 to 23454 (SEQ ID NO: 5), with reference to the Genbank sequence AY274119.3 Tor2.

[0373] The plasmid, called TOP10F'-SARS-S2, was deposited under the No. I-3019, on May 12, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains a 3' fragment of the sequence of the S gene of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said fragment called Sb corresponding to the nucleotides at positions 23322 to 25348 (SEQ ID NO: 6), with reference to the Genbank sequence accession No. AY274119.3.

b) Cloning and Sequencing of the Complete cDNA (SARS-S Clone of 4 kb)

[0374] The complete S cDNA was obtained from the abovementioned clones SARS-S1 and SARS-S2, in the following manner:

[0375] 1) A PCR amplification reaction was carried out on a SARS-S2 clone in the presence of the abovementioned primer S/R4/-/25348-25329 and of the primer S/S/+24696-24715: an amplicon of 633 bp was obtained,

[0376] 2) Another PCR amplification reaction was carried out on another SARS-S2 clone, in the presence of the primers S/F4/+23322-23341 mentioned above and S/S/-24803-24784: an amplicon of 1481 bp was obtained.

[0377] The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, with the exception that 30 amplification cycles comprising a step of denaturation at 94° C. for 20 sec and a step of extension at 72° C. for 2 min 30 sec were carried out.

[0378] 3) The 2 amplicons (633 bp and 1481 bp) were purified under the conditions as defined above for the Sa and Sb fragments.

[0379] 4) Another PCR amplification reaction with the aid of the abovementioned primers S/F4/+23322-23341 and

S/R4/-/25348-25329 was carried out on the purified amplicons obtained in 3). The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, except that 30 amplification cycles were performed.

[0380] The 2026 bp amplicon thus obtained was purified, cloned into the vector PCR2.1-TOPO and then sequenced as above, with the aid of the primers as-defined above for the Sa and Sb fragments. The clone thus obtained was called clone 3'.

[0381] 5) The clone SARS-S1 obtained above and the clone 3"were digested with EcoR I, the bands of about 2 kb thus obtained were gel purified and then amplified by PCR with the abovementioned primers S/F2/+21406-21426 and S/R4/-/25348-25329. The amplification reaction was carried out under the conditions as defined above for the amplification of the Sa and Sb fragments, except that 30 amplification cycles were performed. The amplicon of about 4 kb was purified and sequenced. It was then cloned into the vector PCR2.1-TOPO in order to give the plasmid, called SARS-S, and the insert obtained in this plasmid was sequenced as above, with the aid of the primers as defined above for the Sa and Sb fragments. The cDNA sequences of the insert and of the amplicon encoding the S protein correspond respectively to the sequences SEQ ID NO: 4 and SEQ ID NO: 2 in the sequence listing appended as an annex, they encode the S protein (SEQ ID NO: 3).

[0382] The sequence of the amplicon corresponding to the cDNA encoding the S protein of the SARS-CoV strain derived from the sample No. 031589 has the following two mutations compared with the corresponding sequences of respectively the Tor2 and Urbani isolates, the positions of the mutations being indicated with reference to the complete sequence of the genome of the Tor2 isolate (Genbank AY274119.3):

[0383] g/t in position 23220; the alanine codon (gct) in position 577 of the amino acid sequence of the S protein of Tor2 is replaced with a serine codon (tct),

[0384] c/t in position 24872: this mutation does not modify the amino acid sequence of the S protein, and

the plasmid, called SARS-S, was deposited under the No. I-3059, on Jun. 20, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the S protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, said sequence corresponding to the nucleotides at positions 21406 to 25348 (SEQ ID NO: 4), with reference to the Genbank sequence AY274119.3.

2.2) cDNA Encoding the M and E Proteins

[0385] The RNAs derived from the sample 031589, extracted as above, were subjected to a reverse transcription, combined, during the same step (Titan One Step RT-PCR® kit, Roche), with a PCR amplification reaction, with the aid of the pairs of primers:

[0386] S/E/F1/+26051-26070 and S/E/R1/-/26455-26436 in order to amplify ORF-E, and

[0387] S/M/F1/+26225-26244 and S/M/R1/-/27148-27129 in order to amplify ORF-M.

[0388] A first reaction mixture containing: 8.6 µl of H₂O for injection, 1 µl of dNTP (5 mM), 0.2 µl of each of the primers (50 µM), 1.25 µl of DTT (100 mM) and 0.25 µl of RNAsin (40 IU/µl) was combined with a second reaction mixture containing: 1 µl of RNA, 7 µl of H₂O for injection, 5 µl of 5×RT-PCR buffer and 0.5 µl of enzyme mixture and the combined mixtures were incubated in a thermocycler under the following conditions: 30 min at 42° C., 10 min at 55° C., 2 min at 94° C. followed by 40 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 55° C. for 30 sec and a step of extension at 68° C. for 45 sec, with 3 sec increment per cycle and finally a step of terminal extension at 68° C. for 7 min.

[0389] The amplification products thus obtained (M and E amplicons) were subjected to a second PCR amplification (nested PCR) using the Expand High-Fi® kit, (Roche), with the aid of the pairs of primers:

[0390] S/E/F2/+26082-26101 and S/E/R2/-26413-26394 for the amplicon E, and

[0391] S/M/F2/+26330-26350 and S/M/R2/-27098-27078 for the amplicon M.

[0392] The reaction mixture containing: 2 µl of the product of the first PCR, 39.25 µl of H₂O for injection, 5 µl of 10× buffer containing MgCl₂, 2 µl of dNTP (5 mM), 0.5 µl of each of the primers (50 µM) and 0.75 µl of enzyme mixture was incubated in a thermocycler under the following conditions: a step of denaturation at 94° C. for 2 min was followed by 30 cycles comprising a step of denaturation at 94° C. for 15 sec, a step of annealing at 60° C. for 30 sec and a step of extension at 72° C. for 45 sec, with 3 sec increment per cycle, and finally a step of terminal extension at 72° C. for 7 min. The amplification products obtained corresponding to the cDNAs encoding the E and M proteins were sequenced as above, with the aid of the primers: S/E/F2/+26082 and S/E/R2/-26394, S/M/F2/+26330, S/M/R2/-27078 cited above and the primers S/M/+26636-26655 and S/M/-26567-26548. They were then cloned, as above, in order to give the plasmids called SARS-E and SARS-M. The DNA of these clones was then isolated and sequenced with the aid of the universal primers M13 forward and M13 reverse and the primers S/M/+26636 and S/M/-26548 mentioned above.

[0393] The sequence of the amplicon representing the cDNA encoding the E protein (SEQ ID NO: 13) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani. The sequence of the E protein of the SARS-CoV 031589 strain corresponds to the sequence SEQ ID NO: 14 in the sequence listing appended as an annex.

[0394] The plasmid, called SARS-E, was deposited under the No. I-3046, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the E protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 26082 to 26413 (SEQ ID NO: 15), with reference to the Genbank sequence accession No. AY274119.3.

[0395] The sequence of the amplicon representing the cDNA encoding M (SEQ ID NO: 16) from the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequence of the isolate AY274119.3-Tor2. By contrast, at position 26857, the

isolate AY278741-Urbani contains a c and the sequence of the SARS-CoV strain derived from the sample recorded under the No. 031589 contains a t. This mutation results in a modification of the amino acid sequence of the corresponding protein: at position 154, a proline (AY278741-Urbani) is changed to serine in the SARS-CoV strain derived from the sample recorded under the No. 031589. The sequence of the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 17 in the sequence listing appended as an annex.

[0396] The plasmid, called SARS-M, was deposited under the No. I-3047, on May 28, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence encoding the M protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above; said sequence corresponding to the nucleotides at positions 26330 to 27098 (SEQ ID NO: 18), with reference to the Genbank sequence accession No. AY274119.3.

2.3) cDNA Corresponding to ORF3, ORF4, ORF7 to ORF11

[0397] The same amplification, cloning and sequencing strategy was used to obtain the cDNA fragments corresponding respectively to the following ORFs: ORF3, ORF4, ORF7, ORF8, ORF9, ORF10 and ORF11. The pairs of primers used for the first amplification are:

[0398] ORF3 and ORF4: S/SE/F1/+25069-25088 and S/SE/R1/-26300-26281

[0399] ORF7 to ORF11: S/MN/F1/+26898-26917 and S/MN/R1/-28287-28266

[0400] The pairs of primers used for the second amplification are:

[0401] ORF3 and ORF4: S/SE/F2/+25110-25129 and S/SE/R2/-26244-26225

[0402] ORF7 to ORF11: S/MN/F2/+26977-26996 and S/MN/R2/-28218-28199

[0403] The conditions for the first amplification (RT-PCR) are the following: 45 min at 42° C., 10 min at 55° C., 2 min at 94° C. followed by 40 cycles comprising a step of denaturation at 94° C. for 15 sec, a step of annealing at 58° C. for 30 sec and a step of extension at 68° C. for 1 min, with 5 sec increment per cycle and finally a step of terminal extension at 68° C. for 7 min. The conditions for the nested PCR are the following: a step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 58° C. for 30 sec and a step of extension at 72° C. for 50 sec, with 4 sec increment per cycle and finally a step of terminal extension at 72° C. for 7 min.

[0404] The amplification products obtained corresponding to the cDNAs containing respectively ORF3 and 4 and ORF7 to 11 were sequenced with the aid of the primers: S/SE/+25363, S/SE/+25835, S/SE/-25494, S/SE/-25875, S/MN/+27839, S/MN/+27409, S/MN/-27836, S/MN/-27799 and cloned as above for the other ORFs, to give the plasmids called SARS-SE and SARS-MN. The DNA of these clones was isolated and sequenced with the aid of these same primers and of the universal primers M13 sense and M13 antisense.

[0405] The sequence of the amplicon representing the cDNA of the region containing ORF3 and ORF4 (SEQ ID NO: 7) of the SARS-CoV strain derived from the sample No.

031589 contains a nucleotide difference in relation to the corresponding sequence of the isolate AY274119-Tor2. This mutation at position 25298 results in a modification of the amino acid sequence of the corresponding protein (ORF3): at position 11, an arginine (AY274119-Tor2) is changed to glycine in the SARS-CoV strain derived from the sample No. 031589. By contrast, no mutation was identified in relation to the corresponding sequence of the isolate AY278741-Urbani. The sequences of ORF3 and 4 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 10 and 12 in the sequence listing appended as an annex.

[0406] The plasmid, called SARS-SE, was deposited under the No. I-3126, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the region situated between ORF-S and ORF-E and overlapping ORF-E of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said region corresponding to the nucleotides at positions 25110 to 26244 (SEQ ID NO: 8), with reference to the Genbank sequence accession No. AY274119.3.

[0407] The sequence of the amplicon representing the cDNA corresponding to the region containing ORF7 to ORF11 (SEQ ID NO: 19) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119-Tor2 and AY278741-Urbani. The sequences of ORF7 to 11 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 22, 24, 26, 28 and 30 in the sequence listing appended as an annex.

[0408] The plasmid, called SARS-MN, was deposited under the No. I-3125, on Nov. 13, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the region situated between ORF-M and ORF-N of the SARS-CoV strain derived from the sample recorded under the No. 031589 and collected in Hanoi, as defined above, said sequence corresponding to the nucleotides at positions 26977 to 28218 (SEQ ID NO: 20), with reference to the Genbank sequence accession No. AY274119.3.

[0409] The sequence of the amplicon representing the cDNA corresponding to the region containing ORF7 to ORF11 (SEQ ID NO: 19) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119-Tor2 and AY278741-Urbani. The sequences of ORF7 to 11 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 22, 24, 26, 28 and 30 in the sequence listing appended as an annex.

2.4) cDNA Encoding the N Protein and Including ORF13 and ORF14

[0410] The cDNA was synthesized and amplified as described above for the fragments Sa and Sb. More specifically, the reaction mixture containing: 5 µl of RNA, 5 µl of H₂O for injection, 4 µl of 5× reverse transcriptase buffer, 2 µl of dNTP (5 mM), 2 µl of oligo 20T (5 µM), 0.5 µl of RNasin (40 IU/µl) and 1.5 µl of AMV-RT (10 IU/µl Promega) was incubated in a thermocycler under the fol-

lowing conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and it was then kept at +4° C.

[0411] A first PCR amplification was performed with the pair of primers S/N/F3/+28023 and S/N/R3/-29480.

[0412] The reaction mixture as above for the amplification of the S1 and S2 fragments was incubated in a thermocycler, under the following conditions: an initial step of denaturation at 94° C. for 2 min was followed by 40 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 1 min 30 sec with 10 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 5 min.

[0413] The amplicon obtained at the first PCR amplification was subjected to a second PCR amplification step (nested PCR) with the pairs of primer S/N/F4/+28054 and S/N/R4/-29430 under conditions identical to those of the first amplification.

[0414] The amplification product obtained, corresponding to the cDNA encoding the N protein of the SARS-CoV strain derived from the sample No. 031589, was sequenced with the aid of the primers: S/N/F4/+28054, S/N/R4/-29430, S/N/+28468, S/N/+28918 and S/N/-28607 and cloned as above for the other ORFs, to give the plasmid called SARS-N. The DNA of these clones was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense, and the primers S/N/+28468, S/N/+28918 and S/N/-28607.

[0415] The sequence of the amplicon representing the cDNA corresponding to ORF-N and including ORF13 and ORF14 (SEQ ID NO: 36) of the SARS-CoV strain derived from the sample No. 031589 does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani. The sequence of the N protein of the SARS-CoV strain derived from the sample No. 031589 corresponds to the sequence SEQ ID NO: 37 in the sequence listing appended as an annex.

[0416] The sequences of ORF13 and 14 of the SARS-CoV strain derived from the sample No. 031589 correspond respectively to the sequences SEQ ID NO: 32 and 34 in the sequence listing appended as an annex.

[0417] The plasmid, called SARS-N, was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA encoding the N protein of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 28054 to 29430 (SEQ ID NO: 38), with reference to the Genbank sequence accession No. AY274119.3.

2.5) Noncoding 5' and 3' Ends

a) Noncoding 5' end (5'NC)

a₁) Synthesis of the cDNA

[0418] The RNAs derived from the sample 031589, extracted as above, were subjected to reverse transcription under the following conditions:

[0419] The RNA (15 µl) and the primer S/L/-443 (3 µl at the concentration of 5 µM) were incubated for 10 min at 75° C.

[0420] Next, the 5× reverse transcriptase buffer (6 µl, INVITROGEN), 10 mM dNTP (1 µl), 0.1 M DTT (3 µl) were added and the mixture was incubated at 50° C. for 3 min.

[0421] Finally, the reverse transcriptase (3 µl of Superscript®, INVITROGEN) was added to the preceding mixture which was incubated at 50° C. for 1 h 30 min and then at 90° C. for 2 min.

[0422] The cDNA thus obtained was purified with the aid of the QIAquick PCR purification kit (QIAGEN) according to the manufacturer's recommendations.

b₁) Terminal Transferase Reaction (TdT)

[0423] The cDNA (10 µl) is incubated for 2 min at 100° C., stored in ice, and the following are then added: H₂O (2.5 µl), 5× TdT buffer (4 µl, AMERSHAM), 5 mM dATP (2 µl) and TdT (1.5 µl, AMERSHAM). The mixture thus obtained is incubated for 45 min at 37° C. and then for 2 min at 65° C.

[0424] The product obtained is amplified by a first PCR reaction with the aid of the primers: S/L/-225-206 and anchor 14T: 5'-AGATGAATTCGGTAC-CTTTTTTTTTTTTTT-3' (SEQ ID NO: 68). The amplification conditions are the following: an initial step of denaturation at 94° C. for 2 min is followed by 10 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 30 sec and then by 30 cycles comprising a step of denaturation at 94° C. for 10 sec, a step of annealing at 50° C. for 30 sec and then a step of extension at 72° C. for 30 sec, and then a final step of extension at 72° C. for 5 min.

[0425] The product of the first PCR amplification was subjected to a second amplification step with the aid of the primers: S/L/-204-185 and anchor 14T mentioned above under conditions identical to those of the first amplification. The amplicon thus obtained was purified, sequenced with the aid of the primer S/L/-182-163 and it was then cloned as above for the different ORFs, to give the plasmid called SARS-5'NC. The DNA of this clone was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense and the primer S/L/-182-163 mentioned above.

[0426] The amplicon representing the cDNA corresponding to the 5'NC end of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 72 in the sequence listing appended as an annex; this sequence does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani.

[0427] The plasmid, called SARS-5'NC, was deposited under the No. I-3124, on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA corresponding to the noncoding 5' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to the nucleotides at positions 1 to 204 (SEQ ID NO: 39), with reference to the Genbank sequence accession No. AY274119.3.

b) Noncoding 3' end (3'NC)

a₁) Synthesis of the cDNA

[0428] The RNAs derived from the sample 031589, extracted as above, were subjected to reverse transcription, according to the following protocol: the reaction mixture containing: RNA (5 µl), H₂O (5 µl), 5× reverse transcriptase buffer (4 µl), 5 mM dNTP (2 µl), 5 µM Oligo 20T (2 µl), 40 U/µl RNasin (0.5 µl) and 10 IU/µl RT-AMV (1.5 µl, PROMEGA) was incubated in a thermocycler, under the following conditions: 45 min at 42° C., 15 min at 55° C., 5 min at 95° C., and it was then kept at +4° C.

[0429] The cDNA obtained was amplified by a first PCR reaction with the aid of the primers S/N/+28468-28487 and anchor 14T mentioned above. The amplification conditions are the following: an initial step of denaturation at 94° C. for 2 min is followed by 10 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 50 sec and then 30 cycles comprising a step of denaturation at 94° C. for 20 sec, a step of annealing at 50° C. for 30 sec and then a step of extension at 72° C. for 50 sec, and then a final step of extension at 72° C. for 5 min.

[0430] The product of the first PCR amplification was subjected to a second amplification step with the aid of the primers S/N/+28933-28952 and anchor 14T mentioned above, under conditions identical to those of the first amplification. The amplicon thus obtained was purified, sequenced with the aid of the primer S/N/+29257-29278 and cloned as above for the different ORFs, to give the plasmid called SARS-3'NC. The DNA of this clone was isolated and sequenced with the aid of the universal primers M13 sense and M13 antisense and the primer S/N/+29257-29278 mentioned above.

[0431] The amplicon representing the cDNA corresponding to the 3'NC end of the SARS-CoV strain derived from the sample recorded under the No. 031589 corresponds to the sequence SEQ ID NO: 73 in the sequence listing appended as an annex; this sequence does not contain differences in relation to the corresponding sequences of the isolates AY274119.3-Tor2 and AY278741-Urbani.

[0432] The plasmid called SARS-3'NC was deposited under the No. I-3123 on Nov. 7, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15; it contains the cDNA sequence corresponding to the noncoding 3' end of the genome of the SARS-CoV strain derived from the sample recorded under the No. 031589, as defined above, said sequence corresponding to that situated between the nucleotide at positions 28933 to 29727 (SEQ ID NO: 40), with reference to the Genbank sequence accession No. AY274119.3, ends with a series of nucleotides a.

2.6) ORF1a and ORF1b

[0433] The amplification of the 5' region containing ORF1a and ORF1b of the SARS-CoV genome derived from the sample 031589 was performed by carrying out RT-PCR reactions followed by nested PCRs according to the same principles as those described above for the other ORFs. The amplified fragments overlap over several tenths of bases, thus allowing computer reconstruction of the complete sequence of this part of the genome. On average, the amplified fragments are of two kilobases.

[0434] 14 overlapping fragments, called L0 to L12, were thus amplified with the aid of the following primers:

TABLE II

Primers used for the amplification of the 5' region (ORF1a and ORF1b)				
REGION AMPLIFIED AND SEQUENCED (does not include the primers)	RT-PCR sense primer	RT-PCR antisense primer	Nested PCR sense primer	Nested PCR antisense primer
L0 50-480	S/L0/F1/+30	S/L0/R1/-481		
L1 231-2240	S/L1/F1/+147	S/L1/R1/-2336	S/L1/F2/+211	S/L1/R2/-2241
L2 2156-4167	S/L2/F1/+2033	S/L2/R1/-4192	S/L2/F2/+2136	S/L2/R2/-4168
L3 3913-5324	S/L3bis/F1/+3850	S/L3bis/R1/-5365	S/L3bis/F2/+3892	S/L3bis/R2/-5325
L4b 4952-6023	S/L4b/F1/+4878	S/L4b/R1/-6061	S/L4b/F2/+4932	S/L4b/R2/-6024
L4 5325-7318	S/L4/F1/+5272	S/L4/R1/-7392	S/L4/F2/+5305	S/L4/R2/-7323
L5 7296-9156	S/L5/F1/+7111	S/L5/R1/-9253	S/L5/F2/+7275	S/L5/R2/-9157
L6 9053-11066	S/L6/F1/+8975	S/L6/R1/-11151	S/L6/F2/+9032	S/L6/R2/-11067
L7 10928-12962	S/L7/F1/+10883	S/L7/R1/-13050	S/L7/F2/+10928	S/L7/R2/-12963
L8 12835-14834	S/L8/F1/+12690	S/L8/R1/-14857	S/L8/F2/+12815	S/L8/R2/-14835
L9 14765-16624	S/L9/F1/+14688	S/L9/R1/-16678	S/L9/F2/+14745	S/L9/R2/-16625
L10 16534-18570	S/L10/F1/+16451	S/L10/R1/-18594	S/L10/F2/+16514	S/L10/R2/-18571
L11 18521-20582	S/L11/F1/+18441	S/L11/R1/-20612	S/L11/F2/+18500	S/L11/R2/-20583
L12 20338-22205	S/L12/F1/+20279	S/L12/R1/-22229	S/L12/F2/+20319	S/L12/R2/-22206

All the fragments were amplified under the following conditions, except fragment L0 which was amplified as described above for ORF-M:
RT-PCR: 30 min at 42° C., 15 min at 55° C., 2 min at 94° C., and then the cDNA obtained is amplified under the following conditions: 40 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 58° C. for 30 sec and then a step of extension at 68° C. for 1 min 30 sec, with 5 sec additional extension at each cycle, and then a final step of extension at 68° C. for 7 min.

Nested PCR: An initial step of denaturation at 94° C. for 2 min is followed by 35 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 60° C. for 30 sec and then a step of extension at 72° C. for 1 min 30 sec, with 5 sec of additional extension at each cycle, and then a final step of extension at 72° C. for 7 min.

[0435] The amplification products were sequenced with the aid of the primers defined in table III below:

TABLE III

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L3/+4932	5'-CCACACACAGCTTGTGGATA-3'
S/L4/+6401	5'-CCGAAGTTGTAGGCAATGTC-3'
S/L4/+6964	5'-TTTGGTGTCTCTTCTTATTG-3'
S/L4/-6817	5'-CCGGCATCCAAACATAATT-3'
S/L5/-7633	5'-TGGTCAGTAGGGTTGATTGG-3'
S/L5/-8127	5'-CATCCTTTGTGTCAACATCG-3'
S/L5/-8633	5'-GTCACGAGTGACACCATCCT-3'
S/L5/+7839	5'-ATGCGACGAGTCTGCTTCTA-3'
S/L5/+8785	5'-TTCATAGTGCCTGGCTTACC-3'
S/L5/+8255	5'-ATCTTGGCGCATGTATTGAC-3'
S/L6/-9422	5'-TGCAATTAGCAGCAACAACAT-3'

TABLE III-continued

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L6/-9966	5'-TCTGCAGAACAGCAGAAGTG-3'
S/L6/-10542	5'-CCTGTGCAGTTTGTCTGTCA-3'
S/L6/+10677	5'-CCTTGTGGCAATGAAGTACA-3'
S/L6/+10106	5'-ATGTCAATTGCACAGCAGAA-3'
S/L6/+9571	5'-CTTCAATGGTTTGCCATGTT-3'
S/L7/-11271	5'-TGCGAGCTGTCATGAGAATA-3'
S/L7/-11801	5'-AACCGAGAGCAGTACCACAG-3'
S/L7/-12383	5'-TTTGGCTGCTGTAGTCAATG-3'
S/L7/+12640	5'-CTACGACAGATGCTCTGTGC-3'
S/L7/+12088	5'-GAGCAGGCTGTAGCTAATGG-3'
S/L7/+11551	5'-TTAGGCTATTGTTGCTGCTG-3'
S/L8/-13160	5'-CAGACAACATGAAGCACCAC-3'
S/L8/-13704	5'-CGTGACGTGATATATGTGG-3'

TABLE III-continued

Primers used for the sequencing of the 5' region (ORF1a and ORF1b)	
Names	Sequences (SEQ ID NO: 76 to 139)
S/L8/-14284	5'-TGCACAATGAAGGATACACC-3'
S/L8/+14453	5'-ACATAGCTCGCGTCTCAGTT-3'
S/L8/+13968	5'-GGCATTGTAGGCGTACTGAC-3'
S/L8/+13401	5'-GTTTGCGGTGTAAAGTGCAG-3'
S/L9/-15098	5'-TAGTGGCGGCTATTGACTTC-3'
S/L9/-15677	5'-CTAAACCTTGAGCCGCATAG-3'
S/L9/-16247	5'-CATGGTCATAGCAGCACTTG-3'
S/L9/+16323	5'-CCAGGTTGTGATGTCAGTAT-3'
S/L9/+15858	5'-CCTTACCCAGATCCATCAAG-3'
S/L9/+15288	5'-CGCAAACATAACACTTGCTG-3'
S/L10/-16914	5'-AGTGTGGGTACAGCCAGT-3'
S/L10/-17466	5'-GTTCCAAGGAACATGTCTGG-3'
S/L10/-18022	5'-AGGTGCCTGTGTAGGATGAA-3'
S/L10/+18245	5'-GGGCTGTGTCATGCACTAGAG-3'
S/L10/+17663	5'-TCTTACACGCAATCCTGCTT-3'
S/L10/+17061	5'-TACCATCTGCTCGCATAGT-3'
S/L11/-18877	5'-GCAAGCAGAAATTAACCCTCA-3'
S/L11/-19396	5'-AGCACCACTAAATGTCATC-3'
S/L11/-20002	5'-TGGTCCCTTTGAAGGTGTTA-3'
S/L11/+20245	5'-TCGAACACATCGTTTATGGA-3'
S/L11/+19611	5'-GAAGCACCTGTTTCCATCAT-3'
S/L11/+19021	5'-ACGATGCTCAGCCATGTAGT-3'
SARS/L1/F3/+800	5'-GAGGTGCAGTCACTCGCTAT-3'
SARS/L1/F4/+1391	5'-CAGAGATTGGACCTGAGCAT-3'
SARS/L1/F5/+1925	5'-CAGCAAACCACTCAATTCCT-3'
SARS/L1/R3/-1674	5'-AAATGATGGCAACCTCTTCA-3'
SARS/L1/R4/-1107	5'-CACGTGGTTGAATGACTTTG-3'
SARS/L1/R5/-520	5'-ATTCTGCAACCAGCTCAAC-3'
SARS/L2/F3/+2664	5'-CGCATGTCTCCTGGTTTAC-3'
SARS/L2/F4/+3232	5'-GAGATTGAGCCAGAACCCAGA-3'
SARS/L2/F5/+3746	5'-ATGAGCAGGTTGTCATGGAT-3'
SARS/L2/R3/-3579	5'-CTGCCCTAAGAAGCTGGATG-3'
SARS/L2/R4/-2991	5'-TTTCTTACCAGCATCATCA-3'
SARS/L2/R5/-2529	5'-CACCGTTCTTGAGAACCAACC-3'
SARS/L3/F3/+4708	5'-TCTTTGGCTGGCTCTTACAG-3'
SARS/L3/F4/+5305	5'-GCTGGTGATGCTGCTAACTT-3'
SARS/L3/F5/+5822	5'-CCATCAAGCCTGTGTCGTAT-3'
SARS/L3/R3/-5610	5'-CAGGTGGTGCGACATCATA-3'
SARS/L3/R4/-4988	5'-AACATCAGCACCATCCAAGT-3'
SARS/L3/R5/-4437	5'-ATCGGACACCATAGTCAACG-3'

[0436] The sequences of the fragments L0 to L12 of the SARS-CoV strain derived from the sample recorded under the No. 031589 correspond respectively to the sequences SEQ ID NO: 41 to SEQ ID NO: 54 in the sequence listing appended as an annex. Among these sequences, only that corresponding to the fragments L5 contains a nucleotide difference in relation to the corresponding sequence of the isolate AY278741-Urbani. This t/c mutation at position 7919 results in a modification of the amino acid sequence of the corresponding protein, encoded by ORF1a: at position 2552, a valine (gtt codon; AY278741) is changed to alanine (gct codon) in the SARS-CoV strain 031589. By contrast, no mutation was identified in relation to the corresponding sequence of the isolate AY274119.3-Urbani. The other fragments do not exhibit differences in relation to the corresponding sequences of the isolates Tor2 and Urbani.

EXAMPLE 2

Production and Purification of the Recombinant N and S Proteins of the SARS-CoV Strain Derived from the Sample Recorded Under the Number 031589

[0437] The entire N protein and two polypeptide fragments of the S protein of the SARS-CoV strain derived from

the sample recorded under the number 031589 were produced in *E. coli*, in the form of fusion proteins comprising an N- or C-terminal polyhistidine tag. In the two S polypeptides, the N- and C-terminal hydrophobic sequences of the S protein (signal peptide: positions 1 to 13 and transmembrane helix: positions 1196 to 1218) were deleted whereas the β helix (positions 565 to 687) and the two motifs of the coiled-coil type (positions 895 to 980 and 1155 to 1186) of the S protein were preserved. These two polypeptides consist of: a long fragment (S_L) corresponding to positions 14 to 1193 of the amino acid sequence of the S protein and a short fragment (S_C) corresponding to positions 475 to 1193 of the amino acid sequence of the S protein.

1) Cloning of the cDNAs N, S_L and S_C into the Expression Vectors pIVEX2.3 and pIVEX2.4

[0438] The cDNAs corresponding to the N protein and to the S_L and S_C fragments were amplified by PCR under standard conditions, with the aid of the DNA polymerase Platinum Pfx® (INVITROGEN). The plasmids SRAS-N and SRAS-S were used as template and the following oligonucleotides as primers:

5'- <u>CCCATATG</u> TCTCTGATAATGGACCCCAATCAAA C-3'	(N sense, SEQ ID NO: 55)
5'- <u>CCCCCGGG</u> TGCTGAGTTGAATCAGCAGAAG C-3'	(N antisense, SEQ ID NO: 56)
5'- <u>CCCATATG</u> AGTGACCTTGACCGGTGCACCA C-3'	(S_C sense, SEQ ID NO: 57)
5'- <u>CCCATATG</u> AACCTTGCACCCACCTGCT C-3'	(S_L sense, SEQ ID NO: 58)
5'- <u>CCCCCGGG</u> TTTAATATATTGCTCATATTTTC C-3'.	(S_C and S_L antisense, SEQ ID NO: 29)

[0439] The sense primers introduce an NdeI site (underlined) while the antisense primers introduce an XmaI or SmaI site (underlined). The 3 amplification products were column purified (QIAquick PCR Purification kit, QIAGEN) and cloned into an appropriate vector. The plasmid DNA purified from the 3 constructs (QIAfilter Midi Plasmid kit, QIAGEN) was verified by sequencing and digested with the enzymes NdeI and XmaI. The 3 fragments corresponding to the cDNAs N, S_L and S_C were purified on agarose gel and then inserted into the plasmids pIVEX2.3MCS (C-terminal polyhistidine tag) and pIVEX2.4d (N-terminal polyhistidine tag) digested beforehand with the same enzymes. After verification of the constructs, the 6 expression vectors thus obtained (pIV2.3N, pIV2.3 S_C , pIV2.3 S_L , pIV2.4N, pIV2.4 S_C also called pIV2.4 S_1 , pIV2.4 S_L) were then used, on the one hand to test the expression of the proteins in vitro, and on the other hand to transform the bacterial strain BL21(DE3)pDIA17 (NOVAGEN). These constructs encode proteins whose expected molecular mass is the following: pIV2.3N (47174 Da), pIV2.3 S_C (82897 Da), pIV2.3 S_L (132056 Da), pIV2.4N (48996 Da), pIV2.4 S_1 (81076 Da) and pIV2.4 S_L (133877 Da). Bacteria transformed with pIV2.3N were deposited at the CNCM on Oct. 23, 2003, under the number I-3117, and bacteria transformed with pIV2.4 S_1 were deposited at the CNCM on Oct. 23, 2003, under the number I-3118.

2) Analysis of the Expression of the Recombinant Proteins In Vitro and In Vivo

[0440] The expression of recombinant proteins from the 6 recombinant vectors was tested, in a first instance, in a

system in vitro (RTS100, Roche). The proteins produced in vitro, after incubation of the recombinant vectors pIVEX for 4 h at 30° C., in the RTS100 system, were analyzed by Western blotting with the aid of an anti-(his)₆ antibody coupled to peroxidase. The result of expression in vitro (FIG. 1) shows that only the N protein is expressed in large quantities, regardless of the position, N- or C-terminal, of the polyhistidine tag. In a second step, the expression of the N and S proteins was tested in vivo at 30° C. in LB medium in the presence or in the absence of inducer (1 mM IPTG). The N protein is very well produced in this bacterial system (FIG. 2) and is found mainly in a soluble fraction after lysis of the bacteria. By contrast, the long version of S (S_L) is very weakly produced and is completely insoluble (FIG. 3). The short version (S_C) also exhibits a very weak solubility, but an expression level that is much higher than that of the long version. Moreover, the construct S_C fused with a polyhistidine tag at the C-terminal position has a smaller size than that expected. An immunodetection experiment with an anti-polyhistidine antibody has shown that this construct was incomplete. In conclusion, the two constructs, pIV2.3N and pIV2.4S_L, which express respectively the entire N protein fused with the C-terminal polyhistidine tag and the short S protein fused with the N-terminal polyhistidine tag, were selected in order to produce the two proteins in a large quantity so as to purify them. The plasmids pIV2.3N and pIV2.4S were deposited respectively under the No. I-3117 and I-3118 at the CNCM, 25 rue du Docteur Roux, 75724 PARIS 15, on Oct. 23, 2003.

3) Analysis of the Antigenic Activity of the Recombinant Proteins

[0441] The antigenic activity of the N, S_L and S_C proteins was tested by Western blotting with the aid of two serum samples, obtained from the same patient infected with SARS-CoV, collected 8 days (M12) and 29 days (M13) after the onset of the SARS symptoms. The experimental protocol is as described in example 3. The results illustrated by FIG. 4 show (i) the seroconversion of the patient, and (ii) that the N protein possesses a higher antigenic reactivity than the short S protein.

4) Purification of the N Protein from pIV2.3N

[0442] Several experiments for purifying the N protein, produced from the vector pIV2.3N, were carried out according to the following protocol. The bacteria BL21(DE3)pDIA17, transformed with the expression vector pIV2.3N, were cultured at 30° C. in 1 liter of culture medium containing 0.1 mg/ml of ampicillin, and induced with 1 mM IPTG when the cell density equivalent to A₆₀₀=0.8 is reached (about 3 hours). After 2 hours of culture in the presence of inducer, the cells were recovered by centrifugation (10 min at 5000 rpm), resuspended in the lysis buffer (50 mM NaH₂PO₄, 0.3 M NaCl, 20 mM imidazole, pH 8, containing the mixture of protease inhibitors Complete®, Roche), and lysed with the French press (12 000 psi). After centrifugation of the bacterial lysate (15 min at 12 000 rpm), the supernatant (50 ml) was deposited at a flow rate of 1 ml/min on a metal chelation column (15 ml) (Ni-NTA superflow, Qiagen), equilibrated with the lysis buffer. After washing the column with 200 ml of lysis buffer, the N protein was eluted with an imidazole gradient (20→250 mM) in 10 column volumes. The fractions containing the N protein were assembled and analyzed by polyacrylamide gel electrophoresis under denaturing conditions followed by staining with Coomassie blue. The results illus-

trated by FIG. 5 show that the protocol used makes it possible to purify the N protein with a very satisfactory homogeneity (95%) and a mean yield of 15 mg of protein per liter of culture.

5) Purification of the S_C Protein from pIV2.4S_C (pIV2.4S_L)

[0443] The protocol followed for purifying the short S protein is very different from that described above because the protein is highly aggregated in the bacterial system (inclusion bodies). The bacteria BL21(DE3)pDIA17, transformed with the expression vector pIV2.4S_L, were cultured at 30° C. in 1 liter of culture medium containing 0.1 mg/ml of ampicillin, and induced with 1 mM IPTG when the cell density equivalent to A₆₀₀=0.8 is reached (about 3 hours). After 2 hours of culture in the presence of inducer, the cells were recovered by centrifugation (10 min at 5000 rpm), resuspended in the lysis buffer (0.1 M Tris-HCl, 1 mM EDTA, pH 7.5), and lysed with the French press (1200 psi). After centrifugation of the bacterial lysate (15 min at 12 000 rpm), the pellet was resuspended in 25 ml of lysis buffer containing 2% Triton X100 and 10 mM β-mercaptoethanol, and then centrifuged for 20 min at 12 000 rpm. The pellet was resuspended in 10 mM Tris-HCl buffer containing 7 M urea, and gently stirred for 30 min at room temperature. This final washing of the inclusion bodies with 7 M urea is necessary in order to remove most of the *E. coli* membrane proteins which co-sediment with the aggregated S_C protein. After a final centrifugation for 20 min at 12 000 rpm, the final pellet is resuspended in the 10 mM Tris-HCl buffer. The electrophoretic analysis of this preparation (FIG. 6) shows that the short S protein may be purified with a satisfactory homogeneity (about 90%) from the inclusion bodies (insoluble extract).

EXAMPLE 3

Immunodominance of the N Protein

[0444] The reactivity of the antibodies present in the serum of patients suffering from atypical pneumopathy caused by the SARS-associated coronavirus (SARS-CoV), toward the various proteins of this virus, was analyzed by Western blotting under the conditions described below.

1) Materials

a) Lysate of Cells Infected with SARS-CoV

[0445] Vero E6 cells (2×10⁶) were infected with SARS-CoV (isolate recorded under the number FFM/MA104) at a multiplicity of infection (M.O.I.) of 10⁻¹ or 10⁻² and then incubated in DMEM medium containing 2% FCS, at 35° C. in an atmosphere containing 5% CO₂. 48 hours later, the cellular lawn was washed with PBS and then lysed with 500 μl of loading buffer prepared according to Laemmli and containing β-mercaptoethanol. The samples were then boiled for 10 minutes and then sonicated for 3 times 20 seconds.

b) Antibodies

b₁) Serum from a Patient Suffering from Atypical Pneumopathy

[0446] The serum designated by a reference at the National Reference Center for Influenza Viruses (Northern region) under the No. 20033168 is that from a French patient suffering from atypical pneumopathy caused by SARS-CoV

collected on day 38 after the onset of the symptoms; the diagnosis of SARS-CoV infection was performed by nested RT-PCR and quantitative PCR.

b₂) Monospecific Rabbit Polyclonal Sera Directed Against the N Protein or the S Protein

[0447] The sera are those produced from the recombinant N and S_C proteins (example 2), according to the immunization protocol described in example 4; they are the rabbit P13097 serum (anti-N serum) and the rabbit P11135 serum (anti-S serum).

2) Method

[0448] 20 µl of lysate of cells infected with SARS-CoV at M.O.I. values of 10⁻¹ and 10⁻² and, as a control, 20 µl of a lysate of noninfected cells (mock) were separated on 10% SDS polyacrylamide gel and then transferred onto a nitrocellulose membrane. After blocking in a solution of PBS/5% milk/0.1% Tween and washing in PBS/0.1% Tween, this membrane was hybridized overnight at 4° C. with: (i) the immune serum No. 20033168 diluted 1/300, 1/1000 and 1/3000 in the buffer PBS/1% BSA/0.1% Tween, (ii) the rabbit P13097 serum (anti-N serum) diluted 1/50 000 in the same buffer and (iii) the rabbit P11135 serum (anti-S serum) diluted 1/10 000 in the same buffer. After washing in PBS/Tween, a secondary hybridization was performed with the aid of either sheep polyclonal antibodies directed against the heavy and light chains of human G immunoglobulins and coupled with peroxidase (NA933V, Amersham), or of donkey polyclonal antibodies directed against the heavy and light chains of the rabbit G immunoglobulins and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized with the aid of the ECL+ kit (Amersham) and of Hyperfilm MP autoradiography films (Amersham). A molecular mass ladder (kDa) is presented in the figure.

3) Results

[0449] FIG. 7 shows that three polypeptides of apparent molecular mass 35, 55 and 200 kDa are specifically detected in the extracts of cells infected with SARS-CoV.

[0450] In order to identify these polypeptides, two other immunoblots (FIG. 8) were prepared on the same samples and under the same conditions with rabbit polyclonal antibodies specific for the nucleoprotein N (rabbit P13097, FIG. 8A) and for the spicule protein S (rabbit P11135, FIG. 8B). This experiment shows that the 200 kDa polypeptide corresponds to the SARS-CoV spicule glycoprotein S, that the 55 kDa polypeptide corresponds to the nucleoprotein N while the 35 kDa polypeptide probably represents a truncated or degraded form of N.

[0451] The data presented in FIG. 7 therefore show that the serum 20033168 strongly reacts with N and a lot more weakly with the SARS-CoV S since the 35 and 55 kDa polypeptides are visualized in the form of intense bands for 1/300, 1/1000 and 1/3000 dilutions of the immunosera whereas the 200 kDa polypeptide is only weakly visualized for a dilution of 1/300. It is also possible to note that no other SARS-CoV polypeptide is detected for dilutions greater than 1/300 of the serum 20033168.

[0452] This experiment indicates that the antibody response specific for the SARS-CoV N dominates the antibody responses specific for the other SARS-CoV polypeptides and in particular the antibody response directed against

the S glycoprotein. It indicates an immuno-dominance of the nucleoprotein N during human infections with SARS-CoV.

EXAMPLE 4

Preparation of Monospecific Polyclonal Antibodies Directed Against the SRAS-associated Coronavirus (SARS-CoV) N and S Proteins

1) Materials and Method

[0453] Three rabbits (P13097, P13081, P13031) were immunized with the purified recombinant polypeptide corresponding to the entire nucleoprotein (N), prepared according to the protocol described in example 2. After a first injection of 0.35 mg per rabbit of protein emulsified in complete Freund's adjuvant (intradermal route), the animals received 3 booster injections at 3 and then 4 weeks' interval, of 0.35 mg of recombinant protein emulsified in incomplete Freund's adjuvant.

[0454] Three rabbits (P11135, P13042, P14001) were immunized with the recombinant polypeptide corresponding to the short fragment of the S protein (S_C) produced as described in example 2. As this polypeptide is found mainly in the form of inclusion bodies in the bacterial cytoplasm, the animals received 4 intradermal injections at 3-4 weeks' interval of a preparation of inclusion bodies corresponding to 0.5 mg of recombinant protein emulsified in incomplete Freund's adjuvant. The first 3 injections were made with a preparation of inclusion bodies prepared according to the protocol described in example 2, while the fourth injection was made with a preparation of inclusion bodies which were prepared according to the protocol described in example 2 and then purified on sucrose gradient and washed in 2% Triton X100.

[0455] For each rabbit, a preimmune (p.i.) serum was prepared before the first immunization and an immune serum (I.S.) 5 weeks after the fourth immunization.

[0456] In a first instance, the reactivity of the sera was analyzed by ELISA test on preparations of recombinant proteins similar to those used for the immunizations; the ELISA tests were carried out according to the protocol and with the reagents as described in example 6.

[0457] In a second instance, the reactivity of the sera was analyzed by preparing an immunoblot (Western blot) of a lysate of cells infected with SARS-CoV, according to the protocol as described in example 3.

2) Results

[0458] The ELISA tests (FIG. 9) demonstrate that the preparations of recombinant N protein and of inclusion bodies of the short fragment of the S protein (S_C) are immunogenic in animals and that the titer of the immune sera is high (more than 1/25 000).

[0459] The immunoblot (FIG. 8) shows that the rabbit P13097 immune serum recognizes two polypeptides present in the lysates of cells infected with SARS-CoV: a polypeptide whose apparent molecular mass (50-55 kDa based on experiments) is compatible with that of the nucleoprotein N (422 residues, predicted molecular mass of 46 kDa) and a polypeptide of 35 kDa, which probably represents a truncated or degraded form of N.

[0460] This experiment also shows that the rabbit P11135 serum mainly recognizes a polypeptide whose apparent molecular mass (180-220 kDa based on experiments) is compatible with a glycosylated form of S (1255 residues, nonglycosylated polypeptide chain of 139 kDa), as well as lighter polypeptides, which probably represent truncated and/or nonglycosylated forms of S.

[0461] In conclusion, all these experiments demonstrate that the recombinant polypeptides expressed in *E. coli* and corresponding to the SARS-CoV N and S proteins make it possible to induce, in animals, polyclonal antibodies capable of recognizing the native forms of these proteins.

EXAMPLE 5

Preparation of Monospecific Polyclonal Antibodies Directed Against the SARS-associated Coronavirus (SARS-CoV) M and E Proteins

1) Analysis of the Structure of the M and E Proteins

a) E Protein

[0462] The structure of the SARS-CoV E protein (76 amino acids) was analyzed in silico, with the aid of various software packages such as signalP v1.1, NetNGlyc 1.0, THMM 1.0 and 2.0 (Krogh et al., 2001, *J. Mol. Biol.*, 305(3):567-580) or alternatively TOPPRED (von Heijne, 1992, *J. Mol. Biol.* 225, 487-494). The analysis shows that this nonglycosylated polypeptide is a type 1 membrane protein, containing a single transmembrane helix (aa 12-34 according to THMM), and in which the majority of the hydrophilic domain (42 residues) is located at the C-terminal end and probably inside the viral particle (endodomain). It is possible to note an inversion in the topology predicted by versions 1.0 (N-ter is external) and 2.0 (N-ter is internal) of the THMM software, but that other algorithms, in particular TOPPRED and THUMBUP (Zhou et Zhou, 2003, *Protein Science* 12:1547-1555) confirm an external location of the N-terminal end of E.

b) M Protein

[0463] A similar analysis carried out on the SARS-CoV M protein (221 amino acids) shows that this polypeptide does not possess a signal peptide (according to the software signalP v1.1) but three transmembrane domains (residues 15-37, 50-72, 77-99 according to THMM2.0) and a large hydrophilic domain (aa 100-221) located inside the viral particle (endodomain). It is probably glycosylated on the asparagine at position 4 (according to NetNGlyc 1.0).

[0464] Thus, in agreement with the experimental data known for the other coronaviruses, it is remarkable that the two M and E proteins exhibit endodomains corresponding to the majority of the polypeptides and of the ectodomains that are very small in size.

[0465] The ectodomain of E probably corresponds to residues 1 to 11 or 1 to 12 of the protein: MYSFV-SEETGT(L), SEQ ID NO: 70. Indeed, the probability associated with the transmembrane location of residue 12 is intermediate (0.56 according to THMM 2.0).

[0466] The ectodomain of M probably corresponds to residues 2 to 14 of the protein: ADNGTITVEELKQ, SEQ ID NO: 69. Indeed, the N-terminal methionine of M is very probably cleaved from the mature polypep-

tide because the residue at position 2 is an alanine (Varshavsky, 1996, 93:12142-12149).

[0467] Moreover, the analysis of the hydrophobicity (Kyte & Doolittle, Hopp & Woods) of the E protein demonstrates that the C-terminal end of the endodomain of E is hydrophilic and therefore probably exposed at the surface of this domain. Thus, a synthetic peptide corresponding to this end is a good immunogenic candidate for inducing, in animals, antibodies directed against the endodomain of E. Consequently, a peptide corresponding to 24 C-terminal residues of E was synthesized.

2) Preparation of Antibodies Directed Against the Ectodomain of the M and E Proteins and the Endodomain of the E Protein

[0468] The peptides M2-14 (ADNGTITVEELKQ, SEQ ID NO: 69), E1-12 (MYSFVSEETGTL, SEQ ID NO: 70) and E53-76 (KPTVYVYSRV KNLNSSEGVP DLLV, SEQ ID NO: 71) were synthesized by Neosystem. They were coupled with KLH (Keyhole Limpet Hemocyanin) with the aid of MBS (m-maleimido-benzoyl-N-hydroxysuccinimide ester) via a cysteine added during the synthesis either at the N-terminus of the peptide (case for E53-76) or at the C-terminus (case of M2-14 and E1-12).

[0469] Two rabbits were immunized with each of the conjugates, according to the following immunization protocol: after a first injection of 0.5 mg of peptide coupled with KLH and emulsified in complete Freund's adjuvant (intradermal route), the animals receive 2 to 4 booster injections at 3 or 4 weeks' interval of 0.25 mg of peptide coupled to KLH and emulsified in incomplete Freund's adjuvant.

[0470] For each rabbit, a preimmune (p.i.) serum was prepared before the first immunization and an immune serum (I.S.) is prepared 3 to 5 weeks after the booster injections.

[0471] The reactivity of the sera was analyzed by Western blotting with the aid of extracts of cells infected with SARS-CoV (FIG. 43B) or with the aid of extracts of cells infected with a recombinant vaccinia virus expressing the protein E (VV-TG-E, FIG. 43A) or M (VV-TN-M, FIG. 43C) of the SARS-CoV 031589 isolate.

[0472] The immune sera of the rabbits 22234 and 22240, immunized with the conjugate KLH-E53-76, recognize a polypeptide of about 9 to 10 kD, which is present in the extracts of cells infected with SARS-CoV but absent from the extracts of noninfected cells (FIG. 43B). The apparent mass of this polypeptide is compatible with the predicted mass of the E protein, which is 8.4 kD. Similarly, the immune serum of the rabbit 20047, immunized with the conjugate KLH-E1-12, recognizes a polypeptide present in the extracts of cells infected with the VV-TG-E virus, whose apparent molar mass is compatible with that of the E protein (FIG. 43A).

[0473] The immune serum of the rabbits 20013 and 20080, immunized with the conjugate KLH-M2-14, recognizes a polypeptide present in the extracts of cells infected with the VV-TN-M virus (FIG. 43C), whose apparent molar mass (about 18 kD) is compatible with that of the glycoprotein M, which is 25.1 kD and has a high iso-electric point (9.1 for the naked polypeptide).

[0474] These results demonstrate that the peptides E1-12 and E53-76, on the one hand, and the peptide M2-14, on the other hand, make it possible to induce, in animals, polyclonal antibodies capable of recognizing the native forms of the SARS-CoV E and M proteins, respectively.

EXAMPLE 6

Analysis of the ELISA Reactivity of the Recombinant N Protein Toward Sera from Patients Suffering from SARS

1) Materials

[0475] The antigen used to prepare the solid phases is the purified recombinant nucleoprotein N prepared according to the protocol described in example 2.

[0476] The sera to be tested (table IV) were chosen on the basis of the results of analysis of their reactivity by immunofluorescence (IF-SARS titer), toward cells infected with SARS-CoV.

TABLE IV

Sera tested by ELISA				
Reference	Serum No.	Type of serum	Date of the serum***	IF-SARS titer
3050	A	Control	na*	nt**
3048	B	Control	na	nt
033168	D	Patient 1-SARS	Apr. 27, 2003 (D38)	320
033397	E	Patient-1 SARS	May 11, 2005 (D52)	320
032632	F	Patient-2 SARS	Mar. 21, 2003 (D17)	2500
032791	G	Patient-3 SARS	Apr. 04, 2003 (D3)	<40
033258	H	Patient-3 SARS	Apr. 28, 2003 (D27)	160

*na: not applicable.

**nt: not tested.

***the dates indicated correspond to the number of days after the onset of the SARS symptoms.

2) Method

[0477] The N protein (100 μ l) diluted at various concentrations in 0.1 M carbonate buffer, pH 9.6 (1, 2 or 4 μ g/ml) is distributed into the wells of ELISA plates, and then the plates are incubated overnight at laboratory temperature. The plates are washed with PBS-Tween buffer saturated with PBS-skimmed milk-sucrose (5%) buffer. The test sera (100 μ l), diluted beforehand (1/50, 1/100, 1/200, 1/400, 1/800, 1/1600 and 1/3200) are added and then the plates are incubated for 1 h at 37° C. After 3 washings, the peroxidase-labeled anti-human IgG conjugate (reference 209-035-098, JACKSON) diluted 1/18 000 is added and then the plates are incubated for 1 h at 37° C. After 4 washings, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 30 min at room temperature, protected from light. The reaction is then stopped and then the absorbance at 450 nm is measured with the aid of an automated reader.

3) Results

[0478] The ELISA tests (FIG. 10) demonstrate that the recombinant N protein preparation is specifically recognized by the antibodies of sera from patients suffering from SARS collected in the late phase of the infection (≥ 17 days after the onset of the symptoms) whereas it is not significantly recognized by the antibodies of a patient's serum collected

in the early phase of the infection (3 days after the onset of the symptoms) or by control sera from subjects not suffering from SARS.

EXAMPLE 7

ELISA Tests Prepared for a Very Specific and Sensitive Detection of a SARS-associated Coronavirus Infection, from Sera of Patients

1) Indirect ELISA IgG Test

a) Reagents

Preparation of the Plates

[0479] The plates are sensitized with a solution of N protein at 2 μ g/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l. 100 μ l of solution are deposited in the wells and left to incubate at room temperature overnight. Saturation is obtained by prewashing in 10 mM PBS/0.1% Tween buffer, followed by washing with a saturation solution PBS, 25% milk/sucrose.

Diluent Sera

[0480] Buffer 0.48 g/l TRIS, 10 mM PBS, 3.7 g/l EDTA, 15% v/v milk, pH 6.7

Diluent Conjugate

[0481] Citrate buffer (15 g/l), 0.5% Tween, 25% bovine serum, 12% NaCl, 6% v/v skimmed milk pH 6.5

Conjugate

[0482] 50 $\times$ anti-human IgG conjugate, marketed by Bio-Rad: *Platelia H. pylori* kit ref 72778

Other Solutions:

[0483] Washing solution R2, solutions for visualizing with TMB R8 diluent, R9 chromogen, R10 stopping solution: reagents marketed by Bio-Rad (e.g.: *Platelia pylori* kit, ref 72778)

b) Procedure

[0484] Dilute the sera 1/200 in the sample diluent

[0485] Distribute 100 μ l/well

[0486] Incubation 1 h at 37° C.

[0487] 3 washings in 10 $\times$ WASHING solution R2 diluted beforehand 10-fold in demineralized water (i.e., 1 $\times$ washing solution)

[0488] Distribute 100 μ l of conjugate (50 $\times$ conjugate to be diluted immediately before use in the diluent conjugate provided)

[0489] Incubation 1 h at 37° C.

[0490] 4 washings in 1 $\times$ washing solution

[0491] Distribute 200 μ l/well of visualization solution (to be diluted immediately before use e.g.: 1 ml of R9 in 10 ml of R8)

[0492] Incubation for 30 min at room temperature in the dark

[0493] Stop the reaction with 100 μ l/well of R10

[0494] READING at 450/620 nm

[0495] The results can be interpreted by taking a THRESHOLD serum giving a response above which the sera tested would be considered as positive. This serum is chosen and diluted so as to give a significantly higher signal than the background noise.

2) Double Epitope ELISA Test

a) Reagents

Preparation of the Plates

[0496] The plates are sensitized with a solution of N protein at 1 µg/ml in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l. 100 µl of solution are deposited in the wells and left to incubate at room temperature overnight. Saturation is obtained by prewashing in 10 mM PBS/0.1% Tween buffer, followed by washing with a saturation solution 10 mM PBS, 25% (V/V) milk.

Diluent Sera and Conjugate

[0497] Buffer 50 mM TRIS saline, pH 8, 2% milk

Conjugate

[0498] This is the purified recombinant N protein coupled with peroxidase according to the Nakane protocol (Nakane P. K. and Kawaoi A.; (1974): *Peroxydase-labeled antibody, a new method of conjugation. The Journal of Histochemistry and Cytochemistry* Vol. 22, N) 23, pp. 1084-1091), in respective molar ratios 1/2. This ProtN POD conjugate is used at a concentration of 2 µg/ml in serum/conjugate diluent.

Other Solutions:

[0499] Washing solution R2, solutions for visualization with TMB R8, diluent, R9 chromogen, R10 stopping solution: reagents marketed by Bio-Rad (e.g. *Platelia pylori* kit, ref 72778).

b) Procedure

1st Step in "Predilution" Plate

[0500] Dilute each serum 1/5 in the predilution plate (48 µl of diluent+12 µl of serum).

[0501] After having diluted all the sera, distribute 60 µl of conjugate.

[0502] Where appropriate, the serum+conjugate mix is left to incubate.

2nd Step in "Reaction" Plate

[0503] Transfer 100 µl of mixture/well into the reaction plate

[0504] Incubation 1 h 37° C.

[0505] 5 washings in 10× WASHING solution R2 diluted 10-fold beforehand in demineralized water (→1× washing solution)

[0506] Distribute 200 µl/well of visualization solution (to be diluted immediately before use e.g.: 1 ml of R9 in 10 ml of R8)

[0507] Incubation 30 min at room temperature and protected from light

[0508] Stop the reaction with 100 µl/well of R10

[0509] READING at 450/620 nm

[0510] Likewise as for the indirect ELISA test, the results can be interpreted using a "threshold value" serum. Any serum having a response greater than the threshold value serum will be considered as positive.

2) Results

[0511] The sera of patients classified as probable cases of SARS from the French hospital of Hanoi, Vietnam or in relation with the French hospital of Hanoi (JYK) were analyzed using the indirect IgG-N test and the double epitope N test.

[0512] The results of the indirect IgG-N test (FIGS. 14 and 15) and double epitope N test (FIGS. 16 and 17) show an excellent correlation between them and with an indirect ELISA test comparing the reactivity of the sera toward a lysate of VeroE6 cells infected or not infected with SARS-CoV (ELISA-SARS-CoV lysate; see table V below). All the sera collected 12 days or more after the onset of the symptoms were found to be positive, including in patients for whom it had not been possible to document the SARS-CoV virus infection by analyzing respiratory samples by RT-PCR, probably because of a sample being collected too late during the infection ($\geq$ D12). In the case of the patient TTH for whom a nasal sample collected on D7 was found to be negative by RT-PCR, the quality of the sample may be in question.

[0513] Some sera were found to be negative whereas the presence of SARS-CoV was detected by RT-PCR. They are in all cases early sera collected less than 10 days after the onset of the symptoms (e.g.: serum # 032637). In the case of a patient PTTH (serum # 032673), only a suspicion of SARS was raised at the, time the samples were collected.

[0514] In conclusion, the indirect IgG-N and N-double epitope serological tests make it possible to document the SARS-CoV infection in all the patients for the sera collected 12 days or more after the infection.

TABLE V

Results of the ELISA tests						
Sample Num	Patient	Day	PCR-SARS (1)	ELISA SARS-CoV lysate (2)	IgG-N (2nd series)	2Xepitope (2nd series)
033168	JYK	38	POS	+++	>5000	NT
033597	JYK	74	POS	NT	≈5000	NT
032552	VTT	8	NEG-D3&D8&D12	NEG	<200	<5
032544	CTP	16	NEG-D16&D20	++	>5000	>>20

TABLE V-continued

Results of the ELISA tests						
Sample Num	Patient	Day	PCR-SARS (1)	ELISA SARS-CoV lysate (2)	IgG-N (2nd series)	2Xepitope (2nd series)
032546	CJF	15	NEG	++	>5000	>>20
			D15&D19			
032548	PTL	17	NEG	++	>5000	>>20
			D17&D21			
032550	NTH	17	NEG-D17&D21	++	>5000	>>20
032553	VTT	8	NEG-	NEG	<200	<5
			D3&D8&D12			
032554	NTBV	4	POS	NEG	<200	<5
032555	NTBV	4	POS	NEG	<200	<5
032564	NTP	15	POS	++	>5000	>>20
032629	NVH	4	POS	NEG	<200	<5
032631	BTTX	9	POS	NEG	<200	<5
032635	NHH	4	POS	NEG	<200	<5
032637	NHB	10	POS	NEG	<200	<5
032642	BTTX	9	POS	NEG	<200	<5
032643	LTDH	1	POS	NEG	<200	<5
032644	NTBV	4	POS	NEG	<200	<5
032646	TTH	12	NEG	++	>5000	>>20
			D7&D12&D16			
032647	DTH	17	NEG	++	>5000	>>20
			D17&D21			
032648	NNT	15	NEG	++	>5000	>>20
			D15&D19			
032649	PTH	17	NEG	++	>5000	>>20
			D17&D21			
032672	LVV	16	NEG	+	>5000	>>20
			D16&D20			
032673	PTTH	NA	NEG	NEG	<200	<5
032674	PNB	17	NEG	++	>5000	>>20
			D17&D21			
032682	VTH	12	NEG	++	>5000	>>20
			D12&D16			
032683	DTV	17	NEG	+	>1000	>>20
			D17&D21			

Remarks:

(1): The RT-PCR analyses were carried out by nested RT-PCR BNI, LC Artus and LC-N on nasal or pharyngeal swabs; POS means that at least one sample was found to be positive in this patient.

(2): The reactivity of the sera in the ELISA test using a lysate of cells infected with SARS-CoV was classified as very highly reactive (+++), highly reactive (++), reactive (+) and negative according to the OD value obtained at the dilutions tested.

EXAMPLE 8

Detection of SARS-associated Coronavirus (SARS-CoV) by RT-PCR

1) Real Time Development of RT-PCR Conditions with the Aid of Primers Specific for the Gene for the Nucleocapsid Protein—"Light Cyclers N" Test

a) Design of the Primers and Probes

[0515] The primers and probes were designed from the sequence of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589, with the aid of the programme "Light Cyclers Probe Design (Roche)". Thus, the following two series of primers and probes were selected:

series 1 (SEQ ID NO: 60, 61, 64, 65):

sense primer:

N/+ /28507:

5'-GGC ATC GTA TGG GTT G-3'

[28507-28522]

-continued

antisense primer:

N/- /28774:

5'-CAG TTT CAC CAC CTC C-3'

[28774-28759]

probe 1:

5'-GGC ACC CGC AAT CCT AAT AAC AAT

[28561-28586]

GC-fluorescein 3'

probe 2:

5' Red705-GCC ACC GTG CTA CAA CTT

[28588-28608]

CCT-phosphate

series 2 (SEQ ID NO: 62, 63, 66, 67)

sense primer:

N/+ /28375:

5'-GGC TAC TAC CGA AGA G-3'

[28375-28390]

antisense primer:

N/- /28702:

5'-AAT TAC CGC GAC TAC G-3'

[28702-28687]

-continued

probe 1:
 SARS/N/FL:
 5'-ATA CAC CCA AAG ACC ACA TTG GC- [28541-28563]
 fluorescein 3'

probe 2:
 SARS/N/LC705:
 5' Red705-CCC GCA ATC CTA ATA ACA ATG [28565-28589]
 CTG C-phosphate 3'

b) Analysis of the Efficacy of the Two Primer Pairs

[0516] In order to test the respective efficacy of the two pairs of primers, an RT-PCR amplification was carried out on a synthetic RNA corresponding to nucleotides 28054-29430 of the genome of the SARS-CoV strain derived from the sample recorded under the number 031589 and containing the sequence of the N gene.

[0517] More specifically:

[0518] This synthetic RNA was prepared by in vitro transcription with the aid of the T7 phage RNA polymerase, of a DNA template obtained by linearization of the plasmid SRAS-N with the enzyme Bam HI. After eliminating the DNA template by digestion with the aid of DNase 1, the synthetic RNAs are purified by a phenol-chloroform extraction, followed by two successive precipitations in ammonium acetate and isopropanol. They are then quantified by measuring the absorbance at 260 nm and their quality is checked by the ratio of the absorbances at 260 and 280 nm and by agarose gel electrophoresis. Thus, the concentration of the synthetic RNA preparation used for these studies is 1.6 mg/ml, which corresponds to 2.1×10^{15} copies/ml of RNA.

[0519] Decreasing quantities of synthetic RNA were amplified by RT-PCR with the aid of the "Superscript™ One-Step RT-PCR with Platinum® Taq" kit and the pairs of primers No. 1 (N+/28507, N-/28774) (FIG. 1A) and No. 2 (N+/28375, N-/28702) (FIG. 1B), according to the supplier's instructions. The amplification conditions used are the following: the cDNA was synthesized by incubation for 30 min at 45° C., 15 min at 55° C. and then 2 min at 94° C. and it was then amplified by 5 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 45° C. for 30 sec and then a step of extension at 72° C. for 30 sec, followed by 35 cycles comprising: a step of denaturation at 94° C. for 15 sec, a step of annealing at 55° C. for 30 sec and then a step of extension at 72° C. for 30 sec, with 2 sec of additional extension at each cycle, and a final step of extension at 72° C. for 5 min. The amplification products obtained were then kept at 10° C.

[0520] The results presented in FIG. 11 show that the pair of primers No. 2 (N+/28375, N-/28702) makes it possible to detect up to 10 copies of RNA (band of weak intensity) or 10^2 copies (band of good intensity) against 10^4 copies for the pair of primers No. 1 (N+/28507, N-/28774). The amplicons are respectively 268 bp (pair 1) and 328 bp (pair 2).

c) Development of Real Time RT-PCR

[0521] A real time RT-PCR was developed with the aid of the pair of primers No. 2 and of the pair of probes consisting of SRAS/N/FL and SRAS/N/LC705 (FIG. 2).

[0522] The amplification was carried out on a LightCycler™ (Roche) with the aid of the "Light Cycler RNA Amplification Kit Hybridization Probes" kit (reference 2 015 145, Roche) under the following optimized conditions. A reaction mixture containing: H₂O (6.8 µl), 25 mM MgCl₂ (0.8 µl, 4 µM Mg²⁺ final), 5× reaction mixture (4 µl), 3 µM probe SRAS/N/FL (0.5 µl, 0.075 µM final), 3 µM probe SRAS/N/LC705 (0.5 µl, 0.075 µM final), 10 µM primer N+/28375 (1 µl, 0.5 µM final), 10 µM primer N-/28702 (1 µl, 0.5 µM final), enzyme mixture (0.4 µl) and sample (viral RNA, 5 µl) was amplified according to the following program:

<u>Reverse transcription:</u>			}	x45
50° C.	10:00 min	analysis mode: none		
<u>Denaturation:</u>				
95° C.	30 sec × 1	analysis mode: none		
<u>Amplification:</u>				
95° C.	2 sec	analysis mode: quantification* thermal ramp 2.0° C./sec	}	
50° C.	15 sec			
72° C.	13 sec			
<u>Annealing:</u>				
40° C.	30 sec × 1	analysis mode: none		

*The fluorescence is measured at the end of the annealing and at each cycle (in SINGLE mode).

[0523] The results presented in FIG. 12 show that this real time RT-PCR is very sensitive since it makes it possible to detect 10^2 copies of synthetic RNA in 100% of the 5 samples analyzed (29/29 samples in 8 experiments) and up to 10 copies of RNA in 100% of the 5 samples analyzed (40/45 samples in 8 experiments). It also shows that this RT-PCR makes it possible to detect the presence of the SARS-CoV genome in a sample and to quantify the number of genomes present. By way of example, the viral RNA of a SARS-CoV stock cultured on Vero E6 cells was extracted with the aid of the "Qiaamp viral RNA extraction" kit (Qiagen), diluted to 0.05×10^{-14} and analyzed by real time RT-PCR according to the protocol described above; the analysis presented in FIG. 12 shows that this virus stock contains 6.5×10^9 genome-equivalents/ml (geq/ml), which is entirely similar to the 1.0×10^{10} geq/ml value measured with the aid of the "RealArt™ HPA-Coronavirus LC RT PCR Reagents" kit marketed by Artus.

2) Development of Nested RT-PCR Conditions Targeting the Gene for RNA Polymerase—"CDC (Centers for Disease Control and Prevention)/IP Nested RT-PCR" Test

a) Extraction of the Viral RNA

[0524] Clinical sample: QIAamp viral RNA Mini Kit (QIAGEN) according to the manufacturer's instructions, or an equivalent technique. The RNA is eluted in a volume of 60 µl.

b) "SNE/SAR" Nested RT-PCR

First Step: "SNE" Coupled RT-PCR

[0525] The Invitrogen "Superscript™ One-Step RT-PCR with Platinum® Taq" kit was used, but the "Titan" kit from Roche Boehringer can be used in its place with similar results.

[0526] Oligonucleotides:

SNE-S1
5' GGT TGG GAT TAT CCA AAA TGT GA 3'

SNE-AS1
5' GCA TCA TCA GAA AGA ATC ATC ATG 3'

→ Expected size: 440 bp

[0527] 1. Prepare a mix:

H2O	6.5 µl
Reaction mix 2X	12.5 µl
Oligo SNE-S1 50 µM	0.2 µl
Oligo SNE-AS1 50 µM	0.2 µl
RNAasin 40 U/µl	0.12 µl
RT/Platinum Taq mix	0.5 µl

[0528] 2. To 20 µl of the mix, add 5 µl of RNA and carry out the amplification on a thermocycler (ABI 9600 conditions):

2.1	45° C.	30 min.		
	55° C.	15 min.		
	94° C.	2 min.		
2.2.	94° C.	15 sec.	}	x5 cycles
	45° C.	30 sec.		
	72° C.	30 sec.		
2.3.	94° C.	15 sec.	}	x35 cycles
	55° C.	30 sec.		
	72° C.	30 sec. + 2 sec./cycle		
2.4.	72° C.	5 min.		
2.5	10° C.	∞		

Storage at +4° C.

[0529] The RNAasin (N2511/N2515) from Promega was used as RNase inhibitors.

[0530] Synthetic RNAs served as positive control. As the control, 10³, 10² and 10 copies of synthetic RNA R_{SNE} were amplified in each experiment.

Second Step: "SAR" Nested PCR

[0531] Oligonucleotides:

SAR1-S
5' CCT CTC TTG TTC TTG CTC GCA 3'

SAR1-AS
5' TAT AGT GAG CCG CCA CAC ATG 3'

→ Expected size: 121 bp

[0532] 1. Prepare a mix:

H2O	35.8 µl
Taq buffer 10X	5 µl
MgCl ₂ 25 mM	4 µl
Mix dNTPs 5 mM	2 µl
Oligo SAR1-S 50 µM	0.5 µl
Oligo SAR1-AS 50 µM	0.5 µl
Taq DNA pol 5 U/µl	0.25 µl

[0533] AmpliTaq DNA Pol from Applied Biosystems was used (10× buffer without MgCl₂, ref 27216601).

[0534] 2. To 48 µl of the mix, add 2 µl of the product from the first PCR and carry out the amplification (ABI 9600 conditions):

2.1.	94° C.	2 min.		
2.2.	94° C.	30 sec.	}	x5 cycles
	45° C.	45 sec.		
	72° C.	30 sec.		
2.3.	94° C.	30 sec.	}	x35 cycles
	55° C.	30 sec.		
	72° C.	30 sec. + 1 sec./cycle		
2.4.	72° C.	5 min.		
2.5	10° C.	∞		

[0535] 3. Analyze 10 µl of the reaction product on "low-melting" gel (Seakem GTG type) containing 3% agarose.

[0536] The sensitivity of the nested test is routinely, under the conditions described, 10 copies of RNA.

[0537] 4. The fragments can then be purified on QIAquick PCR kit (QIAGEN) and sequenced with the oligos SAR1-S and SAR1-AS.

3) Detection of the SARS-CoV RNA by PCR from Respiratory Samples

a) First Comparative Study

[0538] A comparative study was carried out on a series of respiratory samples received by the National Reference Center for the Influenza Virus (Northern region) and likely to contain SARS-CoV. To do this, the RNA was extracted from the samples with the aid of the "Qiaamp viral RNA extraction" kit (Qiagen) and analyzed by real time RT-PCR, on the one hand with the aid of the pairs of primers and probes of the No. 2 series under the conditions described above on the one hand, and on the other hand with the aid of the kit "LightCycler SARS-CoV quantification kit" marketed by Roche (reference 03 604 438). The results are summarized in table VI below. They show that 18 of the 26 samples are negative and 5 of the 26 samples are positive for the two kits, while one sample is positive for the Roche kit alone and two for the "series 2" N reagents alone. Additionally, for 3 samples (20032701, 20032712, 20032714) the quantities of RNA detected are markedly higher with the reagents (probes and primers) of the No. 2 series. These results indicate that the "series 2" N primers and probes are more sensitive for the detection of the SARS-CoV genome in biological samples than those of the kit currently available.

TABLE VI

Real time RT-PCR analysis of the RNAs extracted from a series of samples from 5 patients with the aid of the pairs of primers and probes of the No. 2 series ("series 2" N) or of the kit "Lightcycler SARS-CoV quantification kit" (Roche). The type of sample is indicated as well as the number of copies of viral genome measured in each of the two tests. NEG: negative RT-PCR.				
Sample No.	Patient	Type of sample	ROCHE KIT	"Series 2" N
20033082	K	nasal	NEG	NEG
20033083	K	pharyngeal	NEG	NEG
20033086	K	nasal	NEG	NEG
20033087	K	pharyngeal	NEG	NEG
20032802	M	nasal	NEG	NEG
20032803	M	expectoration	NEG	NEG
20032806	M	nasal or pharyngeal	NEG	NEG
20031746ARN2	C	pharyngeal	NEG	NEG
20032711	C	nasal or pharyngeal	39	NEG
20032910	B	nasal	NEG	NEG
20032911	B	pharyngeal	NEG	NEG
20033356	V	expectoration	NEG	NEG
20033357	V	expectoration	NEG	NEG
20031725	K	endotracheal asp.	NEG	150
20032657	K	endotracheal asp.	NEG	NEG
20032698	K	endotracheal asp.	NEG	NEG
20032720	K	endotracheal asp.	3	5
20033074	K	stools	115	257
20032701	M	pharyngeal	443	1676
20032702	M	expectoration	NEG	249
20031747ARN2	C	pharyngeal	NEG	NEG
20032712	C	unknown	634	6914
20032714	C	pharyngeal	17	223
20032800	B	nasal	NEG	NEG
20033353	V	nasal	NEG	NEG
20033384	V	nasal	NEG	NEG

b) Second Comparative Study

[0539] The performance of various nested RT-PCR and real time RT-PCR methods were then compared for 121 respiratory samples from possible cases of SARS at the French hospital in Hanoi, Vietnam, taken between the 4th and the 17th day after the onset of the symptoms. Among these samples, 14 were found to be positive during a first test using the nested RT-PCR method targeting ORF1b (encoding replicase) as described initially by Bernhard Nocht Institute (BNI nested RT-PCR).

[0540] Information relating to this test is available on the internet, at the address <http://www15.bni-hamburg.de/bni2/neu2/getfile.acgi?area=eng1=diagnostics&pid=4112>.

[0541] The various tests compared in this study are:

[0542] the quantitative RT-PCR method according to the invention, with the "series 2" N primers and probes described above (LightCycler N column),

[0543] the nested RT-PCR test targeting the RNA polymerase gene described above, developed by the CDC, BNI and Institut Pasteur <CDC/IP nested RT-PCR>),

[0544] the ARTUS kit with the reference "HPA Corona LC RT-PCR Kit # 5601-02", which is a real time RT-PCR test targeting the ORF1b gene,

[0545] the BNI nested RT-PCR test, also targeting the RNA polymerase gene mentioned above.

[0546] The inventors observed:

[0547] 1) an inter-test variability for the same technique, linked to the degradation of the RNA preparation during repeated thawing, in particular for the samples containing the lowest quantities of RNA,

[0548] 2) a reduced sensitivity of the CDC/IP nested RT-PCR compared with the BNI nested RT-PCR, and

[0549] 3) a comparable sensitivity of the quantitative RT-PCR test according to the invention (Lightcycler N) compared with the Artus LightCycler (LC) test.

[0550] These results, which are presented in table VII below, show that the quantitative RT-PCR test according to the invention constitutes an excellent addition—or an alternative—to the tests currently available. Indeed, the SARS-linked coronavirus is an emergent virus which is capable of changing rapidly. In particular, the gene for the RNA polymerase of the SARS-linked coronavirus, which is targeted in most of the tests currently available, can recombine with that of other coronaviruses not linked to SARS. The use of a test targeting this gene exclusively could then lead to the production of false-negatives.

[0551] The quantitative RT-PCR test according to the invention does not target the same genomic region as the ARTUS kit since it targets the gene encoding the N protein. By carrying out a diagnostic test targeting two different genes of the SARS-linked coronavirus, it can therefore be hoped to avoid false-negative type results which could be due to the genetic evolution of the virus.

[0552] Furthermore, it appears particularly advantageous to target the gene for the nucleocapsid protein because it is very stable because of the high selection pressure linked to the high structural constraints regarding this protein.

TABLE VII

Comparison of various methods of analysis by gene amplification, from 121 samples of probable cases of SARS at the French hospital in Hanoi, Vietnam (epidemic 2003)							
NRC No.	Sample type (1)	Sample collection day	Patient	CDC/IP nested RT-PCR	BNI nested RT-PCR	Artus LightCycler kit	LightCycler N (IP)
107 samples	N and P			Negative	Negative	Negative	Negative
032529	P	10	NHB	Negative	Positive	Negative	Negative
032530	N	10	NHB	Positive	Positive	3.10E+01	4.20E+01
032531	P	7	LP	Positive	Positive	7.70E+00	3.10E+00

TABLE VII-continued

Comparison of various methods of analysis by gene amplification, from 121 samples of probable cases of SARS at the French hospital in Hanoi, Vietnam (epidemic 2003)							
NRC No.	Sample type (1)	Sample collection day	Patient	CDC/IP nested RT-PCR	BNI nested RT-PCR	Artus Light Cycler kit	Light Cycler N (IP)
032534	N	15	BND	Positive	Positive	1.60E+00	Negative
032600	P	4	NHH	Negative	Positive	Negative	1.30E+02
032612	P	17	NTS	Negative	Positive	Negative	Negative
032688	P	9	BTX	Positive	Positive	Negative	Negative
032689	N	4	NVH	Positive	Positive	1.20E+01	2.30E+02
032690	P	4	NVH	Negative	Positive	1.60E+00	Negative
032727	P	8	NVH	Positive	Positive	2.30E+02	4.00E+02
032728	N	8	NVH	Positive	Positive	1.10E+03	1.60E+04
032729	P	14	NHB	Positive	Positive	5.90E+00	3.40E+01
032730	N	14	NHB	Positive	Positive	1.30E+02	4.80E+02
032741	P	8	NHH	Positive	Positive	2.10E+02	1.30E+02
positives				10	14	10	9
fraction detected from the 14 positives				71.4%	100.0%	71.4%	64.3%

(1) P = pharyngeal swab N = nasal swab

EXAMPLE 9

Production and Characterization of Monoclonal
Antibodies Directed Against the N Protein

[0553] Balb C mice were immunized with the purified recombinant N protein and their spleen cells fused with an appropriate murine myeloma according to the Köhler and Milstein techniques.

[0554] Nineteen anti-N antibody secreting hybridomas were preselected and their immunoreactivities determined. These antibodies do indeed recognize the recombinant N protein (in ELISA) with variable intensities, and the natural viral N protein in ELISA and/or in Western blotting. FIGS. 18 to 20 show the results of these tests for 15 of these 19 monoclonal antibodies.

[0555] The highly reactive clones 12, 17, 28, 57, 72, 76, 86, 87, 98, 103, 146, 156, 166, 170, 199, 212, 218, 219 and 222 were subcloned. Specificity studies were carried out with the appropriate tools in order to determine the epitopes recognized and verify the absence of reactivity toward other human coronaviruses and certain respiratory viruses.

[0556] Epitope mapping studies (performed on spot membrane with the aid of overlapping peptides of 15 aa) and additional studies performed on the natural N protein in Western blotting revealed the existence of 4 groups of monoclonal antibodies:

[0557] 1. Monoclonal antibodies specific for a major linear epitope at the N-ter position (75-81, sequence: INTNSVP).

[0558] The representative of this group is antibody 156. The hybridoma producing this antibody was deposited at the Collection Nationale de Cultures de Microorganismes (CNCM) of the Institut Pasteur (Paris, France) on Dec. 1, 2004, under the number I-3331. This same epitope is also recognized by a rabbit serum (anti-N polyclonal) obtained by conventional immunization with the aid of this same N protein.

[0559] 2. Monoclonal antibodies specific for a major linear epitope located in a central position (position 217-224, sequence: ETALALL); the representatives of this group are the monoclonal antibodies 87 and 166. The hybridoma producing antibody 87 was deposited at the CNCM on Dec. 1, 2004, under the number I-3328.

[0560] 3. Monoclonal antibodies specific for a major linear epitope located at the C-terminal position (position 403-408, sequence: DFFRQL), the representatives of this group are the antibodies 28, 57 and 143. The hybridoma producing antibody 57 was deposited at the CNCM on Dec. 1, 2004, under the number I-3330.

[0561] 4. Monoclonal antibodies specific for a discontinuous conformational epitope. This group of antibodies does not recognize any of the peptides spanning the sequence of the N protein, but react strongly on the non-denatured natural protein. The representative of this final group is the antibody 86. The hybridoma producing this antibody was deposited at the CNCM on Dec. 1, 2004, under the number I-3329.

[0562] Table VIII below summarizes the epitope mapping results obtained:

TABLE VIII

Epitope mapping of the monoclonal antibodies			
Antibody	Epitope	Position	Region
28	DFSRQL Q	403 . . . 408	C-Ter.
143	DFSRQL Q		
76	DFSRQL Q		
57	DFSRQL Q		
	FFGMS RI	315 . . . 319	
146	LPQRQ	383 . . . 387	
166	ETALALLLL	217 . . . 224	central
87	ETALALL	217 . . . 224	
156	INTNSGP	75 . . . 81	N-Ter.
86	Conformational		
212	Conformational		
170	Conformational		

EXAMPLE 10

Combinations of the Monoclonal Antibodies for the Development of a Sensitive Immunocapture Test Specific for the Viral N Antigen in the Serum or Biological Fluids of Patients Infected with the SARS-CoV Virus

[0563] The antibodies listed below were selected because of their very specific properties for an additional capture and detection study of the viral N protein, in the serum of the subjects or patients.

[0564] These antibodies were produced in ascites on mice, purified by affinity chromatography and used alone or in combination, as capture antibodies and as signal antibodies.

[0565] List of the antibodies selected:

[0566] Ab anti-C-ter region (No. 28, 57, 143)

[0567] Ab anti-central region (No. 87, 166)

[0568] Ab anti-N-ter region (No. 15-6)

[0569] Ab anti-discontinuous conformational epitope 486)

1) Preparation of the Reagents:

a) Immunocapture ELISA plates

[0570] The plates are sensitized with the antibody solutions at 5 µg/ml in 0.1 M carbonate buffer, pH 9.6. The (monovalent or plurivalent) solutions are deposited in a volume of 100 µl in the wells and incubated overnight at room temperature. These plates are then washed with PBS buffer (10 mM pH 7.4 supplemented with 0.1% Tween 20) and then saturated with a PBS solution supplemented with 0.3% BSA and 5% sucrose). The plates are then dried and then packaged in a bag in the presence of a desiccant. They are ready to use.

b) Conjugates

[0571] The purified antibodies were coupled with peroxidase according to the Nakane protocol (Nakane et al.—1974, J. of Histo and cytochemistry, vol. 22, pp. 1084-1091) in a ratio of one molecule of IgG per 3 molecules of peroxidase. These conjugates were purified by exclusion chromatography and stored concentrated (concentration between 1 and 2 mg/ml) in the presence of 50% glycerol and at -20° C. They are diluted for their use in the assays at the final concentration of 1 or 2 µg/ml in PBS buffer (pH 7.4) supplemented with 1% BSA.

c) Other Reagents

[0572] Human sera negative for all the serum markers for the HIV, HBV, HCV and THLV viruses

[0573] Pool of negative human sera supplemented with 0.5% Triton X 100

[0574] Inactivated viral Ag: viral culture supernatant inactivated by irradiation and inactivation verified after placing in culture on sensitive cells—titer of the suspension before inactivation about 10⁷ infectious particles per ml or alternatively about 5×10⁹ physical viral particles per ml of antigen

[0575] The Ag samples diluted in negative human serum: these samples were prepared by diluting 1:100 and then by 5-fold serial dilution.

[0576] These noninfectious samples mimic human samples thought to contain low to very low concentrations of viral nucleoprotein N. Such samples are not available for routine work.

[0577] Washing solution R2, solution for visualization TMB R8, chromogen R9 and stop solution R10, are the generic reagents marketed by Bio-Rad in its ELISA kits (e.g.: *Platelia pylori* kit ref. 72778).

2) Procedure

[0578] The samples of human sera overloaded with inactivated viral Ag are distributed in an amount of 100 µl per well, directly in the ready-to-use sensitized plates, and then incubated for 1 hour at 37° C. (Bio-Rad IPS incubation).

[0579] The material not bound to the solid phase is removed by 3 washings (washing with dilute R2 solution, automatic LP 35 washer).

[0580] The appropriate conjugates, diluted to the final concentration of 1 or 2 µg/ml, are distributed in an amount of 100 µl per well and the plates are again incubated for one hour at 37° C. (IPS incubation).

[0581] The excess conjugate is removed by 4 successive washings (dilute R2 solution—LP 35 washer).

[0582] The presence of conjugate attached to the plates is visualized after adding 100 µl of visualization solution prepared before use (1 ml of R9 and 10 ml of R8) and after incubation for 30 minutes, at room temperature and protected from light.

[0583] The enzymatic reaction is finally blocked by adding 100 µl of R10 reagent (1 N H₂SO₄) to all the wells.

[0584] The reading is carried out with the aid of an appropriate microplate reader at double wavelength (450/620 nm).

[0585] The results can be interpreted by using, as provisional threshold value, the mean of at least two negative controls multiplied by a factor of 2 or alternatively the mean of 100 negative sera supplemented with an increment corresponding to 6 SD (standard deviation calculated on the 100 individual measurements).

3) Results

[0586] Various capture antibody and signal antibody combinations were tested based on the properties of the antibodies selected, and avoiding the combinations of antibodies specific for the same epitopes in solid phase and as conjugates.

[0587] The best results were obtained with the 4 combinations listed below. These results are reproduced in table IX below.

1. Combination F/28

[0588] Solid phase (Ab 166+87 central region): conjugate antibody 28 (C-ter)

2. Combination G/28

[0589] Solid phase (Ab 86—conformational epitope): conjugate antibody 28 (C-ter)

3. Combination H/28

[0590] Solid phase (Ab 86, 166 and 87 central region and conformational epitope): conjugate antibody 28 (C-ter)

4. Combination H/28+87

[0591] Solid phase (Ab 86, 166 and 87 central region and conformational epitope): mixed conjugate antibodies 28 (C-ter) and 87 (central)

5. Combination G/87

[0592] Solid phase (Ab 86—conformational epitope): conjugate antibody 87 (central region)

[0593] The first 4 combinations exhibit equivalent and reproduced performance levels, greater than the other combinations used (such as for example the combination G/87). Of course, in these combinations, a monoclonal antibody may be replaced with another antibody recognizing the same epitope. Thus, the following variants may be mentioned:

6. Variant of the combination F/28

[0594] Solid phase (Ab 87 only): conjugate antibody 57 (C-ter)

7. Variant of the combination G/28

[0595] Solid phase (Ab 86—conformational epitope): conjugate antibody 57 (C-ter)

8. Variant of the combination H/28

[0596] Solid phase (Ab 86 and 87 central region and conformational epitope): conjugate antibody 57 (C-ter)

9. Variant of the combination H/28+87

[0597] Solid phase (Ab 86 and 87 central region and conformational epitope): mixed conjugate antibodies 57 (C-ter) and 87 (central)

TABLE IX

Test of immunoreactivity of the anti-SARS-CoV nucleoprotein Abs: optical densities measured with each combination of antibodies according to the dilutions of the inactivated viral antigen.

No.	Dilution	F/28	G/28	G/87	H/28	H/28 + 87
0	1/100	5	5	3.495	3.900	5
1	1/500	3.795	3.814	1.379	3.702	3.804
2	1/2 500	2.815	2.950	0.275	3.268	2.680
3	1/12 500	0.987	1.038	0.135	1.374	0.865
4	1/62 500	0.404	0.348	0.125	0.480	0.328
5	1/312 500	0.285	0.211	0.123	0.240	0.215
6	Control	0.210	0.200	0.098	0.186	0.156
7	Control	0.269	0.153	0.104	0.193	0.202

[0598] The detection limit for these 4 experimental trials corresponds to the antigen dilution in negative serum 1:62 500. A rapid extrapolation suggests the detection of less than 10^3 infectious particles per ml of sera.

[0599] From this study, it is evident that the most appropriate antibodies for the capture of the native viral nucleoprotein are the antibodies specific for the central region and/or for a conformational epitope, both being antibodies also selected for their high affinity for the native antigen.

[0600] Having determined the best antibodies for the composition of the solid phase, the antibodies to be selected as a priority for the detection of the antigens attached to the solid phase are the complementary antibodies specific for a dominant epitope in the C-ter region. The use of any other complementary antibody specific for epitopes located in the N-ter region of the protein leads to average or poor results.

EXAMPLE 11

Eukaryotic Expression Systems for the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein

1) Optimization of the Conditions for Expression of the SARS-CoV S in Mammalian Cells

[0601] The conditions for transient expression of the SARS-CoV spicule (S) protein were optimized in mammalian cells (293T, VeroE6).

[0602] For that, a DNA fragment containing the cDNA for SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ATAGGATCCA CCATGTTTAT TTCTTATTA TTTCTTACTC TCACT-3' and 5'-ATACTC-GAGTT ATGTGTAATG TAATTTGACA CCCTTG-3' from the plasmid pSARS-S (C.N.C.M. No. I-3059) and then inserted between the BamH1 and Xho1 sites of the plasmid pTRIPAU3-CMV containing a lentiviral vector TRIP (Sirven, 2001, Mol. Ther., 3, 438-448) in order to obtain the plasmid pTRIP-S. The BamH1 and Xho1 fragment containing the cDNA for S was then subcloned between BamH1 and Xho1 of the eukaryotic expression plasmid pcDNA3.1(+) (Clontech) in order to obtain the plasmid pcDNA-S. The Nhe1 and Xho1 fragment containing the cDNA for S was then subcloned between the corresponding sites of the expression plasmid pCI (Promega) in order to obtain the plasmid pCI-S. The WPRE sequences of the woodchuck hepatitis virus ("Woodchuck Hepatitis Virus posttranscriptional regulatory element") and the CTE sequences ("constitutive transport element") of the simian retrovirus from Mason-Pfizer were inserted into each of the two plasmids pcDNA-S and pCI-S between the Xho1 and Xba1 sites in order to obtain respectively the plasmids pcDNA-S-CTE, pcDNA-S-WPRE, pCI-S-CTE and pCI-S-WPRE (FIG. 21). The plasmid pCI-S-WPRE was deposited at the CNCM, on Nov. 22, 2004, under the number I-3323. All the inserts were sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377.

[0603] The capacity of the plasmid constructs to direct the expression of SARS-CoV S in mammalian cells was assessed after transfection of VeroE6 cells (FIG. 22). In this experiment, monolayers of 5×10^5 VeroE6 cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pcDNA (as control), pcDNA-S, pCI and pCI-S and 6 μ l of Eugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, cellular extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel, and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum from the rabbit P11135: cf. example 4 above) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL+ kit (Amersham) and autoradiography films Hyperfilm MP (Amersham).

[0604] This experiment (FIG. 22) shows that the plasmid pcDNA-S does not make it possible to direct the expression of SARS-CoV S at detectable levels whereas the plasmid pCI-S allows a weak expression, close to the limit of detection, which may be detected when the film is overexposed. Similar results were obtained when the expression of S was sought by immunofluorescence (data not shown). This impossibility to detect effective expression of S cannot be

attributed to the detection techniques used since the S protein can be detected at the expected size (180 kDa) in an extract of cells infected with SARS-CoV or in an extract of VeroE6 cells infected with the recombinant vaccinia virus VV-TF7.3 and transfected with the plasmid pcDNA-S. In this latter experiment, the virus VV-TF7.3 expresses the RNA polymerase of the T7 phage and allows the cytoplasmic transcription of an uncapped RNA capable of being efficiently translated. This experiment suggests that the expression defects described above are due to an intrinsic inability of the cDNA for S to be efficiently expressed when the step for transcription to messenger RNA is carried out at the nuclear level.

[0605] In a second experiment, the effect of the CTE and WPRE signals on the expression of S was assessed after transfection of VeroE6 (FIG. 23A) and 293T (FIG. 23B) cells and according to a protocol similar to that described above. Whereas the expression of S cannot be detected after transfection of the plasmids pcDNA-S-CTE and pcDNA-S-WPRE derived from pcDNA-S, the insertion of the WPRE and CTE signals greatly improves the expression of S in the context of the expression plasmid pCI-S.

[0606] To specify this result, a second series of experiments were carried out where the immunoblot is quantitatively visualized by luminescence and acquisition on a digital imaging device (FluorS, BioRad). The analysis of the results obtained with the QuantityOne v4.2.3 software (BioRad) shows that the WPRE and CTE sequences increase respectively the expression of S by a factor of 20 to 42 and 10 to 26 in Vero E6 cells (table X). In 293T cells (table X), the effect of the CTE sequence is more moderate (4 to 5 times) whereas that of the WPRE sequence remains high (13 to 28 times).

TABLE X

Quantitative analysis of the effect of the CTE and WPRE signals on the expression of SARS-CoV S: Cellular extracts were prepared 48 hours after transfection of VeroE6 or 293T cells with the plasmid pCI, pCI-S, pCI-S-CTE and pCI-S-WPRE and analyzed by Western blotting as described in the legend to FIG. 22. The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The expression levels are indicated according to an arbitrary scale where the value of 1 represents the level measured after transfection of the plasmid pCI-S. Two independent experiments were carried out for each of the two cell types. In experiment 1 on VeroE6 cells, the transfections were carried out in duplicate and the results are indicated in the form of the mean and standard deviation values for the expression levels measured.			
Plasmid	cell	exp. 1	exp. 2
PCI	VeroE6	0.0	0.0
pCI-S	VeroE6	1.0 ± 0.1	1.0
pCI-S-CTE	VeroE6	9.8 ± 0.9	26.4
pCI-S-WPRE	VeroE6	20.1 ± 2.0	42.3
PCI	293T	0.0	0.0
pCI-S	293T	1.0	1.0
pCI-S-CTE	293T	4.6	4.0
pCI-S-WPRE	293T	27.6	12.8

[0607] In summary, all these results show that the expression, in mammalian cells, of the cDNA for the SARS-CoV S under the control of the RNA polymerase II promoter

sequences requires, to be efficient, the expression of a splice signal and of either of the sequences WPRE and CTE.

2) Production of Stable Lines Allowing the Expression of SARS-CoV S

[0608] The cDNA for the SARS-CoV S protein was cloned in the form of a BamHI-XhoI fragment into the plasmid pTRIPAU3-CMV containing a defective lentiviral vector TRIP with central DNA flap (Sirven et al., 2001, Mol. Ther., 3: 438-448) in order to obtain the plasmid pTRIP-S (FIG. 24). Transient cotransfection according to Zennou et al. (2000, Cell, 101: 173-185) of this plasmid, of an encapsidation plasmid (p8.2) and of a plasmid for expression of the VSV envelope glycoprotein G (pHCMV-G) in 293T cells allowed the preparation of retroviral pseudoparticles containing the vector TRIP-S and pseudotyped with the envelope protein G. These pseudotyped TRIP-S vectors were used to transduce 293T and FRhK-4 cells: no expression of the S protein could be detected by Western blotting and immunofluorescence in the transduced cells (data not presented).

[0609] The optimum expression cassettes consisting of the CMV virus immediate/early promoter, a splice signal, cDNA for S and either of the posttranscriptional signals WPRE or CTE described above were then substituted for the EF1 α -EGFP cassette of the defective lentiviral expression vector with central DNA flap TRIPAU3-EF1 α (Sirven et al., 2001, Mol. Ther., 3: 438-448) (FIG. 25). These substitutions were carried out by a series of successive subclonings of the S expression cassettes which were excised from the plasmids PCT-S-CTE (BglII-ApaI) or respectively pCI-S-WPRE (BglII-SalI) and then inserted between the MluI and KpnI sites or respectively MluI or XhoI sites of the plasmid TRIPAU3-EF1 α in order to obtain the plasmids pTRIP-SD/SA-S-CTE and pTRIP-SD/SA-S-WPRE, deposited at the CNCM, on Dec. 1, 2004, under the numbers I-3336 and I-3334, respectively. Pseudotyped vectors were produced according to Zennou et al. (2000, Cell, 101: 173-185) and used to transduce 293T cells (10 000 cells) and FRhK-4 cells (15 000 cells) according to a series of 5 successive transduction cycles with a quantity of vectors corresponding to 25 ng (TRIP-SD/SA-S-CTE) or 22 ng TRIP-SD/SA-S-WPRE) of p24 per cycle.

[0610] The transduced cells were cloned by limiting dilution and a series of clones were qualitatively analyzed for the expression of SARS-CoV S by immunofluorescence (data not shown), and then quantitatively by Western blotting (FIG. 25) with the aid of an anti-S rabbit polyclonal serum. The results presented in FIG. 25 show that clones 2 and 15 of FRhK4-s-CTE cells transduced with TRIP-SD/SA-S-CTE and clones 4, 9 and 12 of FRhK4-S-WPRE cells transduced with TRIP-SD/SA-S-WPRE allow the expression of the SARS-CoV S at respectively low or moderate levels if they are compared to those which can be observed during infection with SARS-CoV.

[0611] In summary, the vectors TRIP-SD/SA-S-CTE and TRIP-SD/SA-S-WPRE allow the production of stable clones of FRhK-4 cells and similarly 293T cells expressing SARS-CoV S, whereas the assays carried out with the "parent" vector TRIP-S remained unsuccessful, which demonstrates the need for a splice signal and for either of the sequences CTE and WPRE for the production of stable cell clones expressing the S protein.

[0612] In addition, these modifications of the vector TRIP (insertion of a splice signal and of a post-transcriptional signal like CTE and WPRE) could prove advantageous for improving the expression of other cDNAs than that for S.

3) Production of Stable Lines Allowing the Expression of a Soluble Form of SARS-CoV S. Purification of this Recombinant Antigen.

[0613] A cDNA encoding a soluble form of the S protein (Ssol) was obtained by fusing the sequences encoding the ectodomain of the protein (amino acids 1 to 1193) with those of a tag (FLAG: DYKDDDDK) via a BspE1 linker encoding the SG dipeptide. Practically, in order to obtain the plasmid pcDNA-Ssol, a DNA fragment encoding the ectodomain of SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ATAGGATCCA CCATGTTTAT TTTCTTATTA TTTCTTACTC TCACT-3' and 5'-ACCTC-CGGAT TTAATATATT GCTCATATTT TCCCAA-3' from the plasmid pcDNA-S, and then inserted between the unique BamH1 and BspE1 sites of a modified eukaryotic expression plasmid pcDNA3.1(+) (Clontech) containing the tag sequence FLAG between its BamH1 and Xho1 sites:

```
// GGATCC . . . nnn . . . TCC GGA GAT TAT AAA GAT
   BamH1          S G D Y K D
```

```
GAC GAC GAT AAA TAA CTCGAG //
   D D D K   ter Xho1
```

[0614] The Nhe1-Xho1 and BamH1-Xho1 fragments, containing the cDNA for S, were then excised from the plasmid pcDNA-Ssol, and subcloned between the corresponding sites of the plasmid pTRIP-SD/SA-S-CTE and of the plasmid pTRIP-SD/SA-S-WPRE, respectively, in order to obtain the plasmids pTRIP-SD/SA-Ssol-CTE and pTRIP-SD/SA-Ssol-WPRE, deposited at the CNCM, on Dec. 1, 2004, under the numbers I-3337 and I-3335, respectively.

[0615] Pseudotyped vectors were produced according to Zennou et al. (2000, Cell, 101:173-185) and used to transduce FRhK-4 cells (15 000 cells) according to a series of 5 successive transduction cycles (15 000 cells) with a quantity of vector corresponding to 24 ng (TRIP-SD/SA-Ssol-CTE) or 40 ng (TRIP-SD/SA-Ssol-WPRE) of p24 per cycle. The transduced cells were cloned by limiting dilution and a series of 16 clones transduced with TRIP-SD/SA-Ssol-CTE and of 15 clones with TRIP-SD/SA-Ssol-WPRE were analyzed for the expression of the Ssol polypeptide by Western blotting visualized with an anti-FLAG monoclonal antibody (FIG. 26 and data not presented), and by capture ELISA specific for the Ssol polypeptide which was developed for this purpose (table XI and data not presented). Part of the process for selecting the best secretory clones is shown in FIG. 26. Capture ELISA is based on the use of solid phases coated with polyclonal antibodies of rabbits immunized with purified and inactivated SARS-CoV. These solid phases allow the capture of the Ssol polypeptide secreted into the cellular supernatants, whose presence is then visualized with a series of steps successively involving the attachment of an anti-FLAG monoclonal antibody (M2, SIGMA), of anti-mouse IgG(H+L) biotinylated rabbit polyclonal antibodies (Jackson) and of a streptavidin-peroxidase conjugate (Amersham) and then the addition of chromogen and substrate (TMB+H₂O₂, KPL)

TABLE XI

Analysis of the expression of the Ssol polypeptide by cell lines transduced with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. The secretion of the Ssol polypeptide was assessed in the supernatant of a series of cell clones isolated after transduction of FRhK-4 cells with the lentiviral vectors TRIP-SD/SA-Ssol-WPRE and TRIP-SD/SA-Ssol-CTE. The supernatants diluted 1/50 were analyzed by a capture ELISA test specific for SARS-CoV S.		
Vector	Clone	OD (450 nm)
Control	—	0.031
TRIP-SD/SA-Ssol-CTE	CTE2	0.547
	CTE3	0.668
	CTE9	0.171
	CTE12	0.208
	CTE13	0.133
TRIP-SD/SA-Ssol-WPRE	WPPE1	0.061
	WPPE10	0.134

[0616] The cell line secreting the highest quantities of Ssol polypeptide in the culture supernatant is the FRhK4-Ssol-CTE3 line. It was subjected to a second series of 5 cycles of transduction with the vector TRIP-SD/SA-Ssol-CTE under conditions similar to those described above and then cloned. The subclone secreting the highest quantities of Ssol was selected by a combination of Western blot and capture ELISA analysis: it is the subclone FRhK4-Ssol-30, which was deposited at the CNCM, on Nov. 22, 2004, under the name I-3325.

[0617] The FRhK4-Ssol-30 line allows the quantitative production and purification of the recombinant Ssol polypeptide. In a typical experiment where the experimental conditions for growth, production and purification were optimized, the cells of the FRhK4-Ssol-30 line are inoculated in standard culture medium (pyruvate-free DMEM containing 4.5 g/l of glucose and supplemented with 5% FCS, 100 U/ml of penicillin and 100 µg/ml of streptomycin) in the form of a subconfluent monolayer (1 million cells per each 100 cm² in 20 ml of medium). At confluence, the standard medium is replaced with the secretion medium where the quantity of FCS is reduced to 0.5% and the quantity of medium reduced to 16 ml per each 100 cm². The culture supernatant is removed after 4 to 5 days of incubation at 35° C. and under 5% CO₂. The recombinant polypeptide Ssol is purified from the supernatant by the succession of steps of filtration on 0.1 µm polyethersulfone (PES) membrane, concentration by ultrafiltration on a PES membrane with a 50 kD cut-off, affinity chromatography on anti-FLAG matrix with elution with a solution of FLAG peptide (DYKDDDDK) at 100 µg/ml in TBS (50 mM tris, pH 7.4, 150 mM NaCl) and then gel filtration chromatography in TBS on sephadex G-75 beads (Pharmacia). The concentration of the purified recombinant Ssol polypeptide was determined by micro-BCA test (Pierce) and then its biochemical characteristics analyzed.

[0618] Analysis by 8% SDS acrylamide gel stained with silver nitrate demonstrates a predominant polypeptide whose molecular mass is about 180 kD and whose degree of purity may be evaluated at 98% (FIG. 27A). Two main peaks are detected by SELDI-TOF mass spectrometry (Cypher-ger): they correspond to single and double charged forms of a predominant polypeptide whose molecular mass is thus determined at 182.6±3.7 kD (FIGS. 27B and C). After transfer onto Prosorb membrane and rinsing in 0.1% TFA,

the N-terminal end of the Ssol polypeptide was sequenced in liquid phase by Edman degradation on 5 residues (ABI494, Applied Biosystems) and determined as being SDLDR (FIG. 27D). This demonstrates that the signal peptide located at the N-terminal end of the SARS-CoV S protein, composed of aa 1 to 13 (MFIFLLFLTLTSG) according to an analysis carried out with the software signalP v2.0 (Nielsen et al., 1997, *Protein Engineering*, 10:1-6), is cleaved from the mature Ssol polypeptide. The recombinant Ssol polypeptide therefore consists of amino acids 14 to 1193 of the SARS-CoV S protein fused at the C-terminals with a sequence SGDYKDDDDK containing the sequence of the FLAG tag (underlined). The difference between the theoretical molar mass of the naked Ssol polypeptide (132.0 kD) and the real molar mass of the mature polypeptide (182.6 kD) suggests that the Ssol polypeptide is glycosylated.

[0619] A preparation of purified Ssol polypeptide, whose protein concentration was determined by micro-BCA test, makes it possible to prepare a calibration series in order to measure, with the aid of the capture ELISA test described above, the concentrations of Ssol present in the culture supernatants and to review the characteristics of the secretory lines. According to this test, the FRhK4-Ssol-CT3 line secretes 4 to 6 µg/ml of polypeptide Ssol while the FRhK4-Ssol-30 line secretes 9 to 13 µg/ml of Ssol after 4 to 5 days of culture at confluence. In addition, the purification scheme presented above makes it possible routinely to purify from 1 to 2 mg of Ssol polypeptide per liter of culture supernatant.

EXAMPLE 12

Gene Immunization Involving the SARS-associated Corona Virus (SARS-CoV) Spicule (S) Protein

[0620] The effect of a splice signal and of the posttranscriptional signals WPRE and CTE was analyzed after gene immunization of BALB/c mice (FIG. 28).

[0621] For that, BALB/c mice were immunized at intervals of 4 weeks by injecting into the tibialis anterior a saline solution of 50 µg of plasmid DNA of pcDNA-S and pCI-S and, as a control, 50 µg of plasmid DNA of pcDNA-N (directing the expression of SARS-CoV N) or of pCI-HA (directing the expression of the HA of the influenza virus A/PR/8/34) and the immune sera collected 3 weeks after the 2nd injection. The presence of antibodies directed against the SARS-CoV S was assessed by indirect ELISA using as antigen a lysate of VeroE6 cells infected with SARS-CoV and, as a control, a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers (TI) are calculated as the reciprocal of the dilution producing a specific OD of 0.5 (difference between OD measured on a lysate of infected cells and OD measured on a lysate of noninfected cells) after visualization with an anti-mouse IgG polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL) (FIG. 28A).

[0622] Under these conditions, the expression plasmid pcDNA-S only allows the induction of low antibody titers directed against SARS-CoV S in 3 mice out of 6 ($\text{LOG}_{10}(\text{TI})=1.9\pm0.6$) whereas the plasmid pcDNA-N allows the induction of anti-N antibodies at high titers ($\text{LOG}_{10}(\text{TI})=3.9\pm0.3$) in all the animals, and the control plasmids (pCI, pCI-HA) do not result in any detectable antibody ($\text{LOG}_{10}(\text{TI})<1.7$). The plasmid pCI-S equipped with a splice signal allows the induction of antibodies at high titers ($\text{LOG}_{10}(\text{TI})=3.7\pm0.2$), which are approximately 60 times higher than those observed after injection of the plasmid pcDNA-S ($p<10^{-5}$).

[0623] The efficiency of the posttranscriptional signals was studied by carrying out a dose-response study of the anti-S antibody titers induced in the BALB/c mouse as a function of the quantity of plasmid DNA used as immunogen (2 µg, 10 µg and 50 µg). This study (FIG. 28B) demonstrates that the posttranscriptional signal WPRE greatly improves the efficiency of gene immunization when small doses of DNA are used ($p<10^{-5}$ for a dose of 2 µg of DNA and $p<10^{-2}$ for a dose of 10 µg), whereas the effect of the CTE signal remains marginal ($p=0.34$ for a dose of 2 µg of DNA).

[0624] Finally, the antibodies induced in mice after gene immunization neutralize the infectivity of SARS-CoV in vitro (FIGS. 29A and 29B) at titers which are consistent with the titers measured by ELISA.

[0625] In summary, the use of a splice signal and of the posttranscriptional signal WPRE of the woodchuck hepatitis virus considerably improves the induction of neutralizing antibodies directed against SARS-CoV after gene immunization with the aid of plasmid DNA directing the expression of the cDNA for SARS-CoV S.

EXAMPLE 13

Diagnostic Applications of the S Protein

[0626] The ELISA reactivity of the recombinant Ssol polypeptide was analyzed with respect to sera from patients suffering from SARS.

[0627] The sera from probable cases of SARS tested were chosen on the basis of the results (positive or negative) of analysis of their specific reactivity toward the native antigens of SARS-CoV by immunofluorescence test on VeroE6 cells infected with SARS-CoV and/or by indirect ELISA test using as antigen a lysate of VeroE6 cells infected with SARS-CoV. The sera of these patients are identified by a serial number of the National Reference Center for Influenza Viruses and by the initials of the patient and the number of days elapsed since the onset of the symptoms. All the sera of probable cases (cf. Table XII) recognize the native antigens of SARS-CoV, with the exception of the serum 032552 of the patient VTT for whom infection with SARS-CoV could not be confirmed by RT-PCR performed on respiratory samples of days 3, 8 and 12. A panel of control sera was used as control (TV sera): they are sera collected in France before the SARS epidemic that occurred in 2003.

TABLE XII

Sera of probable cases of SARS		
Serum	Patient	Sample collection day
031724	JYK	7
033168	JYK	38
033597	JYK	74
032632	NTM	17
032634	THA	15
032541	PHV	10
032542	NIH	17
032552	VTT	8
032633	PTU	16
032791	JLB	3
033258	JLB	27
032703	JCM	8
033153	JCM	29

[0628] Solid phases sensitized with the recombinant Ssol polypeptide were prepared by adsorption of a solution of purified Ssol polypeptide at 2 µg/ml in PBS in the wells of an ELISA plate, and then the plates are incubated overnight at 4° C. and washed with PBS-Tween buffer (PBS, 0.1% Tween 20). After saturating the ELISA plates with a solution of PBS-10% skimmed milk (weight/volume) and washing in PBS-Tween, the sera to be tested (100 µl) are diluted 1/400 in PBS skimmed milk-Tween buffer (PBS, 3% skimmed milk, 0.1% Tween) and then added to the wells of the sensitized ELISA plate. The plates are incubated for 1 h at 37° C. After 3 washings with PBS-Tween buffer, the anti-human IgG conjugate labeled with peroxidase (ref. NA933V, Amersham) diluted 1/4000 in PBS-skimmed milk-Tween buffer is added, and then the plates are incubated for 1 hour at 37° C. After 6 washings with PBS-Tween buffer, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 10 minutes protected from light. The reaction is stopped by adding a 1 N H₃PO₄ solution, and then the absorbance is measured at 450 nm with a reference at 620 nm.

[0629] The ELISA tests (FIG. 30) demonstrate that the recombinant Ssol polypeptide is specifically recognized by the serum antibodies of patients suffering from SARS collected at the medium or late phase of infection (≥10 days after the onset of the symptoms) whereas it is not significantly recognized by the serum antibodies of 2 patients (JLB and JCM) collected in the early phase of infection (3 to 8 days after the onset of the symptoms) or by control sera of subjects not suffering from SARS. The serum antibodies of patients JLB and JCM show a seroconversion between days 3 and 27 for the first and 8 and 29 for the second after the onset of the symptoms, which confirms the specificity of the reactivity of these sera toward the Ssol polypeptide.

[0630] In conclusion, these results demonstrate that the recombinant Ssol polypeptide may be used as an antigen for the development of an ELISA test for serological diagnosis of infection with SARS-CoV.

EXAMPLE 14

Vaccine Applications of the Recombinant Soluble S Protein

[0631] The immunogenicity of the recombinant Ssol polypeptide was studied in mice.

[0632] For that, a group of 6 mice was immunized at 3 weeks' interval with 10 µg of recombinant Ssol polypeptide adjuvanted with 1 mg of aluminum hydroxide (Alu-gel-S, Serva) diluted in PBS. Three successive immunizations were performed and the immune sera were collected 3 weeks after each of the immunizations (IS1, IS2, IS3). As a control, a group of mice (mock group) received aluminum hydroxide alone according to the same protocol.

[0633] The immune sera were analyzed per pool for each of the 2 groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen and as a control a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG (H+L) polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL). This analysis (FIG. 31) shows that the

immunization with the Ssol polypeptide induces in mice, from the first immunization, antibodies directed against the native form of the SARS-CoV spicule protein present in the lysate of infected VeroE6 cells. After 2 then 3 immunizations, the anti-S antibody titers become very high.

[0634] The immune sera were analyzed per pool for each of the two groups for their capacity to seroneutralize the infectivity of SARS-CoV. 4 points of seroneutralization on FRhK-4 cells (100 TCID₅₀ of SARS-CoV) are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4. This analysis shows that the antibodies induced in mice by the Ssol polypeptide are neutralizing; the titers observed are very high after 2 and then 3 immunizations (greater than 2560 and 5120 respectively, table XIII).

TABLE XIII

Induction of antibodies directed against SARS-CoV after immunization with the recombinant Ssol polypeptide. The immune sera were analyzed per pool for each of the two groups for their capacity to seroneutralize the infectivity of 100 TCID ₅₀ of SARS-CoV on FRhK-4 cells. 4 points are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4.		
Group	Sera	Neutralizing Ab
Mock	pi	<20
	IS1	<20
	IS2	<20
	IS3	<20
Ssol	pi	<20
	IS1	57
	IS2	>2560
	IS3	>5120

[0635] The neutralizing titers observed in mice immunized with the Ssol polypeptide reach levels far greater than the titers observed by Yang et al. in mice (2004, Nature, 428:561-564) and those observed by Buchholz in the hamster (2004, PNAS 101:9804-9809) which protect respectively mice and hamsters from infection with SARS-CoV. It is therefore probable that the neutralizing antibodies induced in mice after immunization with the Ssol polypeptide protect these animals against infection with SARS-CoV.

EXAMPLE 15

Optimized Synthetic Gene for the Expression in Mammalian Cells of the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein

1) Design of the Synthetic Gene

[0636] A synthetic gene encoding the SARS-CoV spicule protein was designed from the gene of the isolate 031589 (plasmid pSARS-S, C.N.C.M. No. I-3059) so as to allow high levels of expression in mammalian cells and in particular in cells of human origin.

[0637] For that:

[0638] the use of codons of the wild-type gene of the isolate 031589 was modified so as to become close to

the bias observed in humans and to improve the efficiency of translation of the corresponding mRNA

[0639] the overall GC content of the gene was increased so as to extend the half-life of the corresponding mRNA

[0640] the optionally cryptic motifs capable of interfering with an efficient expression of the gene were deleted (splice donor and acceptor sites, polyadenylation signals, sequences very rich (>80%) or very low (<30%) in GC, repeat sequences, sequences involved in the formation of secondary RNA structures, TATA boxes)

[0641] a second STOP codon was added to allow efficient termination of translation.

[0642] In addition, CpG motifs were introduced into the gene so as to increase its immunogenicity as DNA vaccine. In order to facilitate the manipulation of the synthetic gene, two BamH1 and Xho1 restriction sites were placed on either side of the open reading frame of the S protein, and the BamH1, Xho1, Nhe1, Kpn1, BspE1 and Sal1 restriction sites were avoided in the synthetic gene.

[0643] The sequence of the synthetic gene designed (gene 040530) is given in SEQ ID No: 140.

[0644] An alignment of the synthetic gene 040530 with the sequence of the wild-type gene of the isolate 031589 of SARS-CoV deposited at the C.N.C.M. under the number I-3059 (SEQ ID No: 4, plasmid pSRAS-S) is presented in FIG. 32.

2) Plasmid Constructs

[0645] The synthetic gene SEQ ID No: 140 was assembled from synthetic oligonucleotides and cloned between the Kpn1 and Sac1 sites of the plasmid pUC-Kana in order to give the plasmid 040530pUC-Kana. The nucleotide sequence of the insert of the plasmid 040530pUC-Kana was verified by automated sequencing (Applied).

[0646] A Kpn1-Xho1 fragment containing the synthetic gene 040530 was excised from the plasmid 040530pUC-Kana and subcloned between the Nhe1 and Xho1 sites of the expression plasmic pCI (Promega) in order to obtain the plasmid pCI-SSYNTH, deposited at the CNCM on Dec. 1, 2004, under the number I-3333.

[0647] A synthetic gene encoding the soluble form of the S protein was then obtained by fusing the synthetic sequences encoding the ectodomain of the S protein (amino acids 1 to 1193) with those of the tag (FLAG: DYKDDDDK) via a linker BspE1 encoding the dipeptide SG. Practically, a DNA fragment encoding the ectodomain of the SARS-CoV S was amplified by PCR with the aid of the oligonucleotides 5'-ACTAGCTAGCGGATCCACCATGTTTCATCTT CCTG-3' and 5'-AGTATCCGGAC TTG ATGTACT GCTCG-TACTTGC-3' from the plasmid 04053-0pUC-Kana, digested with Nhe1 and BspE1 and then inserted between the unique Nhe1 and BspE1 sites of the plasmid pCI-Ssol, to give the plasmid pCI-SCUBE, deposited at the CNCM on Dec. 1, 2004, under the number I-3332. The plasmids pCI-Ssol, pCI-Ssol-CTE, and pCI-Ssol-WPRE (deposited at the CNCM, on Nov. 22, 2004, under the number I-3324) had been previously obtained by subcloning the Kpn1-Xho1 fragment excised from the plasmid pcDNA-Ssol (see technical note of DI 2004-106) between the Nhe1 and Xho1 sites of the plasmids pCI, pCI-S-CTE and pCI-S-WPRE respectively.)

[0648] The plasmids pCI-Scube and pCI-Ssol encode the same recombinant Ssol polypeptide.

3) Results

[0649] The capacity of the synthetic gene encoding the S protein to efficiently direct the expression of the SARS-CoV S in mammalian cells was compared with that of the wild-type gene after transient transfection of primate cells (VeroE6) and of human cells (293T).

[0650] In the experiment presented in FIG. 33 and in table XIV, monolayers of 5×10^5 VeroE6 cells or 7×10^5 293T cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pCI (as control), pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-S-Synth and 6 μ l of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, cell extracts were prepared in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and then transferred onto a PVDF membrane (BioRad). The detection of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum of the rabbit P11135: cf example 4 above) and of donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The immunoblot was quantitatively visualized by luminescence with the aid of the ECL+ kit (Amersham) and acquisition on a digital imaging device (FluorS, BioRad).

[0651] The analysis of the results obtained with the software QuantityOne v4.2.3 (BioRad) shows that in this experiment, the plasmid pCI-Synth allows the transient expression of the S protein at high levels in the VeroE6 and 293T cells, whereas the plasmid pCI-S does not make it possible to induce expression at sufficient levels to be detected. The expression levels observed are of the order of twice as high as those observed with the plasmid pCI-S-WPRE.

TABLE XIV

Use of a synthetic gene for the expression of the SARS-CoV S. Cell extracts prepared 48 hours after transfection of VeroE6 or 293T cells with the plasmids pCI, pCI-S, pCI-S-CTE, pCI-S-WPRE and pCI-S-Synth were separated on 8% SDS acrylamide gel and analyzed by Western blotting with the aid of an anti-S rabbit polyclonal antibody and an anti-rabbit IgG(H + L) polyclonal antibody coupled with peroxidase (NA934V, Amersham). The Western blot is visualized by luminescence (ECL+, Amersham) and acquisition on a digital imaging device (FluorS, BioRad). The expression levels of the S protein were measured by quantifying the two predominant bands identified on the image (see FIG. 33) and are indicated according to an arbitrary scale where the value 1 represents the level measured after transfection of the plasmid pCI-S-WPRE.		
Plasmid	VeroE6	293T
pCI	0.0	0.0
pCI-S	≤0.1	≤0.1
pCI-S-CTE	0.5	≤0.1
pCI-S-WPRE	1.0	1.0
pCI-Ssynth	1.8	1.9

[0652] In a second instance, the capacity of the synthetic gene Scube to efficiently direct the synthesis and the secretion of the Ssol polypeptide by mammalian cells was compared with that of the wild-type gene after transient transfection of hamster cells (BHK-21) and of human cells (293T).

[0653] In the experiment presented in table XV, monolayers of 6×10^5 BHK-21 cells and 7×10^5 293T cells in 35 mm Petri dishes were transfected with 2 μ g of plasmids pCI (as control), pCI-Ssol, pCI-Ssol-CTE, pCI-Ssol-WPRE and pCI-Scube and 6 μ l of Fugene6 reagent according to the manufacturer's instructions (Roche). After 48 hours of incubation at 37° C. and under 5% CO₂, the cellular supernatants were collected and quantitatively analyzed for the secretion of the Ssol polypeptide by a capture ELISA test specific for the Ssol polypeptide.

[0654] Analysis of the results shows that, in this experiment, the plasmid pCI-Scube allows the expression of the Ssol polypeptide at levels 8 times (BHK-21 cells) to 20 times (293T cells) higher than the plasmid pCI-Ssol. The levels of expression observed are of the order of twice (293T cells) to 5 times (BHK-21 cells) as high as those observed with the plasmid pCI-Ssol-WPRE.

TABLE XV

Use of a synthetic gene for the expression of the Ssol polypeptide. The supernatants were harvested 48 hours after transfection of BHK or 293T cells with the plasmids pCI, pCI-Ssol, pCI-Ssol-CTE, pCI-Ssol-WPRE and pCI-Scube and quantitatively analyzed for the secretion of the Ssol polypeptide by an ELISA test specific for the Ssol polypeptide. The transfections were carried out in duplicate and the results are presented in the form of means and standard deviations of the concentrations of Ssol polypeptide (ng/ml) measured in the supernatants.		
Plasmid	BHK	293T
pCI	<20	<20
pCI-Ssol	<20	56 $\pm$ 10
pCI-Ssol-CTE	<20	63 $\pm$ 8
pCI-Ssol-WPRE	28 $\pm$ 1	531 $\pm$ 15
pCI-Scube	152 $\pm$ 6	1140 $\pm$ 20

[0655] In summary, these results show that the expression, in mammalian cells, of the synthetic gene 040530 encoding SARS-CoV S under the control of RNA polymerase II promoter sequences is much more efficient than that of the wild-type gene of the 031589 isolate. This expression is even more efficient than that directed by the wild-type gene in the presence of the WPRE sequences of the woodchuck hepatitis virus.

4) Applications

[0656] The use of the synthetic gene 040530 encoding SARS-CoV S or its Scube variant encoding the polypeptide Ssol is capable of advantageously replacing the wild-type gene in numerous applications where the expression of S is necessary at high levels. In particular in order to:

[0657] improve the efficiency of gene immunization with plasmids of the pCI-Ssynth or even pCI-Ssynth-CTE or pCI-Ssynth-WPRE type

[0658] establish novel cell lines expressing higher quantities of the S protein or of the Ssol polypeptide with the aid of recombinant lentiviral vectors carrying the Ssynth gene or the Scube gene respectively

[0659] improve the immunogenicity of the recombinant lentiviral vectors allowing the expression of the S protein or of the Ssol polypeptide

[0660] improve the immunogenicity of live vectors allowing the expression of the S protein or of the Ssol polypeptide like recombinant vaccinia viruses or recombinant measles viruses (see examples 16 and 17 below)

EXAMPLE 16

Expression of the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein with the Aid of Recombinant Vaccinia Viruses

Vaccine Application

Application to the Production of a Soluble Form of the Spicule (S) Protein and Design of a Serological Test for SARS

1) Introduction

[0661] The aim of this example is to evaluate the capacity of recombinant vaccinia viruses (VV) expressing various SARS-associated coronavirus (SARS-CoV) antigens to constitute novel vaccine candidates against SARS and a means of producing recombinant antigens in mammalian cells.

[0662] For that, the inventors focused on the SARS-CoV spicule (S) protein which makes it possible to induce, after gene immunization in animals, antibodies neutralizing the infectivity of SARS-CoV, and a soluble and secreted form of this protein, the Ssol polypeptide, which is composed of the ectodomain (aa 1-1193) of S fused at its C-ter end with a tag FLAG (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide. This Ssol polypeptide exhibits an antigenicity similar to that of the S protein and allows, after injection into mice in the form of a purified protein adjuvanted with aluminum hydroxide, the induction of high neutralizing antibody titers against SARS-CoV.

[0663] The various forms of the S gene were placed under the control of the promoter of the 7.5K gene and then introduced into the thymidine kinase (TK) locus of the Copenhagen strain of the vaccinia virus by double homologous recombination *in vivo*. In order to improve the immunogenicity of the recombinant vaccinia viruses, a synthetic late promoter was chosen in place of the 7.5K promoter, in order to increase the production of S and Ssol during the late phases of the viral cycle.

[0664] After having isolated the recombinant vaccinia viruses and verified their capacity to express the SARS-CoV S antigen, their capacity to induce in mice an immune response against SARS was tested. After having purified the Ssol antigen from the supernatant of infected cells, an ELISA test for serodiagnosis of SARS was designed, and its efficiency was evaluated with the aid of sera from probable cases of SARS.

2) Construction of the Recombinant Viruses

[0665] Recombinant vaccinia viruses directing the expression of the S glycoprotein of the 031589 isolate of SARS-CoV and of a soluble and secreted form of this protein, the Ssol polypeptide, under the control of the 7.5K promoter were obtained. With the aim of increasing the levels of expression of S and Ssol, recombinant viruses in which the cDNAs for S and for Ssol are placed under the control of a late synthetic promoter were also obtained.

where the quantity of FCS is reduced to 0.5% and the TPB eliminated. The culture supernatant is removed after 2.5 days of incubation at 35° C. and under 5% CO₂ and the vaccinia virus inactivated by addition of Triton X-100 (0.1%). After filtration on 0.1 µm polyethersulfone (PES) membrane, the recombinant Ssol polypeptide is purified by affinity chromatography on an anti-FLAG matrix with elution with a solution of FLAG peptide (DYKDDDDK) at 100 µg/ml in TBS (50 mM Tris, pH 7.4, 150 mM NaCl).

[0674] The analysis by 8% SDS acrylamide gel stained with silver nitrate identified a predominant polypeptide whose molecular mass is about 180 kD and whose degree of purity is greater than 90% (FIG. 37). The concentration of the purified Ssol recombinant polypeptide was determined by comparison with molecular mass markers and estimated at 24 ng/µl.

[0675] This purified Ssol polypeptide preparation makes it possible to produce a calibration series in order to measure, with the aid of a capture ELISA test, the Ssol concentrations present in the culture supernatants. According to this test, the BHK-21 line secretes about 1 µg/ml of Ssol polypeptide under the production conditions described above. In addition, the purification scheme presented makes it possible to purify of the order of 160 µg of Ssol polypeptide per liter of culture supernatant.

[0676] The ELISA reactivity of the recombinant Ssol polypeptide was analyzed toward sera from patients suffering from SARS.

[0677] The sera of probable cases of SARS tested were chosen on the basis of the results (positive or negative) of analysis of their specific reactivity toward the native antigens of SARS-CoV by immunofluorescence test on VeroE6 cells infected with SARS-CoV and/or by indirect ELISA test using, as antigen, a lysate of VeroE6 cells infected with SARS-CoV. The sera of these patients are identified by a serial number of the National Reference Center for Influenza Viruses and by the patient's initials and the number of days elapsed since the onset of the symptoms. All the sera of probable cases (cf. table XVI) recognize the native antigens of SARS-CoV with the exception of the serum 032552 of the patient VTT, for which infection with SARS-CoV could not be confirmed by RT-PCR performed on respiratory samples of days 3, 8 and 12. A panel of control sera was used as control (TV sera): they are sera collected in France before the SARS epidemic which occurred in 2003.

TABLE XVI

Sera of probable cases of SARS		
Serum	Patient	Sample collection day
033168	JYK	38
033597	JYK	74
032632	NTM	17
032634	THA	15
032541	PHV	10
032542	NIH	17
032552	VTT	8
032633	PTU	16

[0678] Solid phases sensitized with the recombinant Ssol polypeptide were prepared by adsorption of a solution of purified Ssol polypeptide at 4 µg/ml in PBS in the wells of

an ELISA plate. The plates are incubated overnight at 4° C. and then washed with PBS-Tween buffer (PBS, 0.1% Tween 20). After washing with PBS-Tween, the sera to be tested (100 µl) are diluted 1/100 and 1/400 in PBS-skimmed milk-Tween buffer (PBS, 3% skimmed milk, 0.1% Tween) and then added to the wells of the sensitized ELISA plate. The plates are then incubated for 1 h at 37° C. After 3 washings with PBS-Tween buffer, the anti-human IgG conjugate labeled with peroxidase (ref. NA933V, Amersham) diluted 1/4000 in PBS-skimmed milk-Tween buffer is added and then the plates are incubated for one hour at 37° C. After 6 washings with PBS-Tween buffer, the chromogen (TMB) and the substrate (H₂O₂) are added and the plates are incubated for 10 minutes protected from light. The reaction is stopped by adding a 1M solution of H₃PO₄ and then the absorbance is measured at 450 nm with a reference at 620 nm.

[0679] The ELISA tests (FIG. 38) demonstrate that the recombinant Ssol polypeptide is specifically recognized by the serum antibodies of patients suffering from SARS, collected at the middle or late phase of infection (≥ 10 days after the onset of the symptoms), whereas it is not significantly recognized by the serum antibodies of the control sera of subjects not suffering from SARS.

[0680] In conclusion, these results demonstrate that the recombinant Ssol polypeptide can be purified from the supernatant of mammalian cells infected with the recombinant vaccinia virus VV-TN-Ssol and can be used as antigen for developing an ELISA test for serological diagnosis of infection with SARS-CoV.

5. Vaccine Applications

[0681] The immunogenicity of the recombinant vaccinia viruses was studied in mice.

[0682] For that, groups of 7 BALB/c mice were immunized by the i.v. route twice at 4 weeks' interval with 10⁶ p.f.u. of recombinant vaccinia viruses VV-TG, VV-T-G-S, VV-TG-Ssol, VV-TN, VV-TN-S and VV-TN-Ssol and, as a control, VV-TG-HA which directs the expression of hemagglutinin of the A/PR/8/34 strain of the influenza virus. The immune sera were collected 3 weeks after each of the immunizations (IS1, IS2).

[0683] The immune sera were analyzed per pool for each of the groups by indirect ELISA using a lysate of VeroE6 cells infected with SARS-CoV as antigen and, as control, a lysate of noninfected VeroE6 cells. The anti-SARS-CoV antibody titers (TI) are calculated as the reciprocal of the dilution producing a specific OD of 0.5 after visualization with an anti-mouse IgG(H+L) polyclonal antibody coupled with peroxidase (NA931V, Amersham) and TMB supplemented with H₂O₂ (KPL). This analysis (FIG. 39A) shows that immunization with the virus VV-TG-S and VV-TN-S induces in mice, from the first immunization, antibodies directed against the native form of the SARS-CoV spicule protein present in the lysate of infected VeroE6 cells. The responses induced by the VV-TN-S virus are higher than those induced by the VV-TG-S virus after the first (TI=740 and TI=270 respectively) and the second (TI=3230 and TI=600 respectively) immunization. The VV-TN-Ssol virus induces high anti-SARS-CoV antibody titers after two immunizations (TI=640), whereas the virus VV-TG-Ssol induces a response at the detection limit (TI=40).

[0684] The immune sera were analyzed per pool for each of the groups for their capacity to seroneutralize the infectivity of SARS-CoV. 4 seroneutralization points on FRhK-4 cells (100 TCID₅₀ of SARS-CoV) are produced for each of the 2-fold dilutions tested from 1/20. The seroneutralizing titer is calculated according to the Reed and Munsch method as the reciprocal of the dilution neutralizing the infectivity of 2 wells out of 4. This analysis shows that the antibodies induced in mice by the vaccinia viruses expressing the S protein or the Ssol polypeptide are neutralizing and that the viruses with synthetic promoters are more efficient immunogens than the viruses carrying the 7.5K promoter: the highest titers (640) are observed after 2 immunizations with the virus VV-TN-S (FIG. 39B).

[0685] The protective power of the neutralizing antibodies induced in mice after immunization with the recombinant vaccinia viruses is evaluated with the aid of a challenge infection with SARS-CoV.

6) Other Applications

[0686] Third generation recombinant vaccinia viruses are constructed by substituting the wild-type sequences of the S and Ssol genes by synthetic genes optimized for the expression in mammalian cells, described above. These recombinant vaccinia viruses are capable of expressing larger quantities of S and Ssol antigens and therefore of exhibiting increased immunogenicity.

[0687] The recombinant vaccinia virus VV-TN-Ssol can be used for the quantitative production and purification of the Ssol antigen for diagnostic (serology by ELISA) and vaccine (subunit vaccine) applications.

EXAMPLE 17

Recombinant Measles Virus Expressing the SARS-associated Coronavirus (SARS-CoV) Spicule (S) Protein. Vaccine Applications.

1) Introduction

[0688] The measles vaccine (MV) induces a lasting protective immunity in humans after a single injection (Hilleman, 2002, Vaccine, 20: 651-665). The protection conferred is very robust and is based on the induction of an antibody response and of a CD4 and CD8 cell response. The MV genome is very stable and no reversion of the vaccine strains to virulence has ever been observed. The measles virus belongs to the genus *Morbillivirus* of the Paramyxoviridae family; it is an enveloped virus whose genome is a 16 kb single-stranded RNA of negative polarity (FIG. 40A) and whose exclusively cytoplasmic replication cycle excludes any possibility of integration into the genome of the host. The measles vaccine is thus one of the most effective and one of the safest live vaccines used in the human population. Frederic Tangy's team recently developed an expression vector on the basis of the Schwarz strain of the measles virus, which is the safest attenuated strain and the most widely used in humans as vaccine against measles. This vaccine strain may be isolated from an infectious molecular clone while preserving its immunogenicity in primates and in mice that are sensitive to the infection. It constitutes, after insertion of additional transcription units, a vector for the expression of heterologous sequences (Combretet, 2003, J. Virol. 77: 11546-11554). In addition, a recombinant MV Schwarz expressing the envelope glycoprotein of the West Nile virus (WNV) induces an effective and lasting antibody response which protects mice from a lethal challenge infec-

tion with WNV (Despres et al., 2004, J. Infect. Dis., in press). All these characteristics make the attenuated Schwarz strain of the measles virus an extremely promising candidate vector for the construction of novel recombinant live vaccines.

[0689] The aim of this example is to evaluate the capacity of recombinant measles viruses (MV) expressing various SARS-associated coronavirus (SARS-CoV) antigens to constitute novel candidate vaccines against SARS.

[0690] The inventors focused on the SARS-CoV spicule (S) protein, which makes it possible to induce, after gene immunization in animals, antibodies neutralizing the infectivity of SARS-CoV, and on a soluble and secreted form of this protein, the Ssol polypeptide, which is composed of the ectodomain (aa 1-1193) of S fused at its C-ter end with a FLAG tag (DYKDDDDK) via a BspEI linker encoding the SG dipeptide. This Ssol polypeptide exhibits a similar antigenicity to that of the S protein and allows, after injection into mice in the form of a purified protein adjuvanted with aluminum hydroxide, the induction of high neutralizing antibody titers against SARS-CoV.

[0691] The various forms of the S gene were introduced in the form of an additional transcription unit between the P (phosphoprotein) and M (matrix) genes into the cDNA of the Schwarz strain of MV previously described (Combretet, 2003, J. Virol. 77: 11546-11554; EP application No. 02291551.6 of Jun. 20, 2002, and EP application No. 02291550.8 of Jun. 20, 2002). After having isolated the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and checked their capacity to express the SARS-CoV S antigen, their capacity to induce a protective immune response against SARS in mice and then in monkeys was tested.

2) Construction of the Recombinant Viruses

[0692] The plasmid pTM-MVSchw-ATU2 (FIG. 40B) contains an infectious cDNA corresponding to the antigenome of the Schwarz vaccine strain of the measles virus (MV) into which an additional transcription unit (ATU) has been introduced between the P (phosphoprotein) and M (matrix) genes (Combretet, 2003, Journal of Virology, 77: 11546-11554). Recombinant genomes MVSchw2-SARS-S and MVSchw2-SARS-Ssol of the measles virus were constructed by inserting ORFs of the S protein and of the Ssol polypeptide into the additional transcription unit of the MVSchw-ATU2 vector.

[0693] For that, a DNA fragment containing the SARS-CoV S cDNA was amplified by PCR with the aid of the oligonucleotides 5'-ATACGTACGA CCATGTTTAT TTTCTTATTA TTTCTTACTC TCACT-3' and 5'-AT-AGCGCGCT CATTATGTGT AATGTAATTT GACAC-CCTTG-3' using the plasmid pcDNA-S as template and then inserted into the plasmid pCR®2.1-TOPO (Invitrogen) in order to obtain the plasmid pTOPO-S-MV. The two oligonucleotides used contain restriction sites BsiW1 and BssHII, so as to allow subsequent insertion into the measles vector, and were designed so as to generate a sequence of 3774 nt including the codons for initiation and termination, so as to observe the rule of 6 which stipulates that the length of the genome of a measles virus must be divisible by 6 (Calain & Roux, 1993, J. Virol., 67: 4822-4830; Schneider et al., 1997, Virology, 227: 314-322). The insert was sequenced with the aid of a BigDye Terminator v1.1 kit (Applied Biosystems) and an automated sequencer ABI377.

[0694] To express a soluble and secreted form of SARS-CoV S, a plasmid containing the cDNA of the Ssol polypep-

tide corresponding to the ectodomain (aa 1-1193) of SARS-CoV S fused at its C-ter end with the sequence of a FLAG tag (DYKDDDDK) via a BspE1 linker encoding the SG dipeptide was then obtained. For that, a DNA fragment was amplified with the aid of the oligonucleotides 5'-CCATTTC AAC AATTTGGCCG-3' and 5'-ATAGGATC-CGCGCGCTCATT ATTTATCGTC GTCATCTTTA TAATC-3' from the plasmid pcDNA-Ssol and then inserted into the plasmid pTOPO-S-MV between the SalI and BamHI sites in order to obtain the plasmid pTOPO-S-MV-SF. The sequence generated is 3618 nt long between the BsiW1 and BssHII sites and observes the rule of 6. The insert was sequenced as indicated above.

[0695] The BsiW1-BssHII fragments containing the cDNAs for the S protein and the Ssol polypeptide were then excised by digestion of the plasmids pTOPO-S-MV and pTOPO-S-MV-SF and then subcloned between the corresponding sites of the plasmid pTM-MVSchw-ATU2 in order to give the plasmids pTM-MVSchw2-SARS-S and pTM-MVSchw2-SARS-Ssol (FIG. 40B). These two plasmids were deposited at the C.N.C.M. on Dec. 1, 2004, under the numbers I-3326 and I-3327, respectively.

[0696] The recombinant measles viruses corresponding to the plasmids pTM-MVSchw2-SARS-S and pTM-MVSchw2-SARS-Ssol were obtained by reverse genetics according to the system based on the use of a helper cell line, described by Radecke et al. (1995, *Embo J.*, 14: 5773-5784) and modified by Parks et al. (1999, *J. Virol.*, 73: 3560-3566). Briefly, the helper cells 293-3-46 are transfected according to the calcium phosphate method with 5 µg of the plasmids pTM-MVSchw2-SARS-S or pTM-MVSchw2-SARS-Ssol and 0.02 µg of the plasmid pEMC-La directing the expression of the MV L polymerase (gift from M. A. Billeter). After incubating overnight at 37° C., a heat shock is produced for 2 hours at 43° C. and the transfected cells are transferred onto a monolayer of Vero cells. For each of the two plasmids, syncytia appeared after 2 to 3 days of coculture and were transferred successively onto monolayers of Vero cells at 70% confluence in 35 mm Petri dishes and then in 25 and 75 cm² flasks. When the syncytia have reached 80-90% confluence, the cells are recovered with the aid of a scraper and then frozen and thawed once. After low-speed centrifugation, the supernatant containing the virus is stored in aliquots at -80° C. The titers of the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol were determined by limiting dilution on Vero cells and the titer as dose infecting 50% of the wells (TCID₅₀) calculated according to the Kärber method.

3) Characterization of the Recombinant Viruses

[0697] The expression of the transgenes encoding the S protein and the Ssol polypeptide was assessed by Western blotting and immunofluorescence.

[0698] Monolayers of Vero cells in T-25 flasks were infected at a multiplicity of 0.05 by various passages of the two viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and the wild-type virus MWSchw as a control. When the syncytia had reached 80 to 90% confluence, cytoplasmic extracts were prepared in an extraction buffer (150 mM NaCl, 50 mM Tris-HCl, pH 7.2, 1% Triton X-100, 0.1% SDS, 1% DOC) and then diluted in loading buffer according to Laemmli, separated on 8% SDS polyacrylamide gel and transferred onto a PVDF membrane (BioRad). The detection

of this immunoblot (Western blot) was carried out with the aid of an anti-S rabbit polyclonal serum (immune serum of the rabbit P11135: cf. example 4 above) and donkey polyclonal antibodies directed against rabbit IgGs and coupled with peroxidase (NA934V, Amersham). The bound antibodies were visualized by luminescence with the aid of the ECL-kit (Amersham) and Hyperfilm MP autoradiography films (Amersham).

[0699] Vero cells in monolayers on glass slides were infected with the two viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol and the wild-type virus MWSchw as a control at multiplicities of infection of 0.05. When the syncytia had reached 90 to 100% (MVSchw2-SARS-Ssol virus) or 30 to 40% (MVSchw2-SARS-S, MWSchw) confluence, the cells were fixed in a 4% PBS-PFA solution, permeabilized with a PBS solution containing 0.2% Triton and then labeled with rabbit polyclonal antibodies hyperimmunized with purified and inactivated SARS-CoV virions and with an anti-rabbit IgG(H+L) goat antibody conjugate coupled with FITC (Jackson).

[0700] As shown in FIGS. 41 and 42, the recombinant viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol direct the expression of the S protein and the Ssol polypeptide respectively at levels comparable to those which can be observed 8 h after infection with SARS-CoV. The expression of these polypeptides is stable after 3 passages of the recombinant viruses in cell culture. These results demonstrate that the recombinant measles viruses are indeed carriers of the transgenes and allow the expression of the SARS glycoprotein in its membrane form (S) or in a soluble form (Ssol). The Ssol polypeptide is expected to be secreted by cells infected with the MVSchw2-SARS-Ssol virus as is the case when this same polypeptide is expressed in mammalian cells after transient transfection of the corresponding sequences (cf. example 11 above).

4) Applications

[0701] Having shown that the viruses MVSchw2-SARS-S and MVSchw2-SARS-Ssol allow the expression of the SARS-CoV S, their capacity to induce a protective immune response against SARS-CoV in CD46^{+/+} IFN-αβR^{-/-} mice, which is sensitive to infection by MV, is evaluated. The antibody response of the immunized mice is evaluated by ELISA test against the native antigens of SARS-CoV and for their capacity to neutralize the infectivity of SARS-CoV in vitro, using the methodologies described above. The protective power of the response will be evaluated by measuring the reduction in the pulmonary viral load 2 days after a nonlethal challenge infection with SARS-CoV.

[0702] Second generation recombinant measles viruses are constructed by substituting the wild-type sequences of the S and Sol genes by synthetic genes optimized for expression in mammalian cells, described in example 15 above. These recombinant measles viruses are capable of expressing larger quantities of the S and Ssol antigens and therefore of exhibiting increased immunogenicity.

[0703] Alternatively, the wild-type or synthetic genes encoding the S protein or the Ssol polypeptide may be inserted into the measles vector MVSchw-ATU3 in the form of an additional transcription unit located between the H and L genes, and then the recombinant viruses produced and characterized in a similar manner. This insertion is capable

of generating recombinant viruses possessing different characteristics (multiplication of the virus, level of expression of the transgene) and possibly an improved immunogenicity compared with those obtained after insertion of the transgenes between the P and N genes.

[0704] The recombinant measles virus MVSchw2-SARS-Ssol may be used for the quantitative production and the purification of the Ssol antigen for diagnostic and vaccine applications.

EXAMPLE 18

Other Applications Linked to the S Protein

[0705] a) The lentiviral vectors allowing the expression of S or Ssol (or even of fragments of S) can constitute a

recombinant vaccine against SARS-CoV, to be used in human or veterinary prophylaxis. In order to demonstrate the feasibility of such a vaccine, the immunogenicity of the recombinant lentiviral vectors TRIP-SD/SA-S-WPRE and TRIP-SD/SA-Ssol-WPRE is studied in mice.

[0706] b) Monoclonal antibodies are produced with the aid of the recombinant Ssol polypeptide. According to the results presented in example 14 above, these antibodies or at least the majority of them will recognize the native form of the SARS-CoV S and will be capable of diagnostic and/or prophylactic applications.

[0707] c) A serological test for SARS is developed with the Ssol polypeptide used as antigen and the double epitope methodology.

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	410						415				420					
act	agg	aac	att	gat	gct	act	tca	act	ggg	aat	tat	aat	tat	aaa	tat	1408
Thr	Arg	Asn	Ile	Asp	Ala	Thr	Ser	Thr	Gly	Asn	Tyr	Asn	Tyr	Lys	Tyr	
	425				430					435					440	
agg	tat	ctt	aga	cat	ggc	aag	ctt	agg	ccc	ttt	gag	aga	gac	ata	tct	1456
Arg	Tyr	Leu	Arg	His	Gly	Lys	Leu	Arg	Pro	Phe	Glu	Arg	Asp	Ile	Ser	
			445						450					455		
aat	gtg	cct	ttc	tcc	cct	gat	ggc	aaa	cct	tgc	acc	cca	cct	gct	ctt	1504
Asn	Val	Pro	Phe	Ser	Pro	Asp	Gly	Lys	Pro	Cys	Thr	Pro	Pro	Ala	Leu	
		460						465					470			
aat	tgt	tat	tg	cca	tta	aat	gat	tat	ggg	ttt	tac	acc	act	act	ggc	1552
Asn	Cys	Tyr	Trp	Pro	Leu	Asn	Asp	Tyr	Gly	Phe	Tyr	Thr	Thr	Thr	Gly	
	475						480					485				
att	ggc	tac	caa	cct	tac	aga	gtt	gta	gta	ctt	tct	ttt	gaa	ctt	tta	1600
Ile	Gly	Tyr	Gln	Pro	Tyr	Arg	Val	Val	Val	Leu	Ser	Phe	Glu	Leu	Leu	
	490					495					500					
aat	gca	ccg	gcc	acg	gtt	tgt	gga	cca	aaa	tta	tcc	act	gac	ctt	att	1648
Asn	Ala	Pro	Ala	Thr	Val	Cys	Gly	Pro	Lys	Leu	Ser	Thr	Asp	Leu	Ile	
	505				510					515					520	
aag	aac	cag	tgt	gtc	aat	ttt	aat	ttt	aat	gga	ctc	act	ggg	act	ggg	1696
Lys	Asn	Gln	Cys	Val	Asn	Phe	Asn	Phe	Asn	Gly	Leu	Thr	Gly	Thr	Gly	
			525						530					535		
gtg	tta	act	cct	tct	tca	aag	aga	ttt	caa	cca	ttt	caa	caa	ttt	ggc	1744
Val	Leu	Thr	Pro	Ser	Ser	Lys	Arg	Phe	Gln	Pro	Phe	Gln	Gln	Phe	Gly	
			540						545				550			
cgt	gat	gtt	tct	gat	ttc	act	gat	tcc	gtt	cga	gat	cct	aaa	aca	tct	1792
Arg	Asp	Val	Ser	Asp	Phe	Thr	Asp	Ser	Val	Arg	Asp	Pro	Lys	Thr	Ser	
	555						560					565				
gaa	ata	tta	gac	att	tca	cct	tgc	tct	ttt	ggg	ggg	gta	agt	gta	att	1840
Glu	Ile	Leu	Asp	Ile	Ser	Pro	Cys	Ser	Phe	Gly	Gly	Val	Ser	Val	Ile	
	570					575					580					
aca	cct	gga	aca	aat	gct	tca	tct	gaa	gtt	gct	gtt	cta	tat	caa	gat	1888
Thr	Pro	Gly	Thr	Asn	Ala	Ser	Ser	Glu	Val	Ala	Val	Leu	Tyr	Gln	Asp	
	585				590					595					600	
gtt	aac	tgc	act	gat	gtt	tct	aca	gca	att	cat	gca	gat	caa	ctc	aca	1936
Val	Asn	Cys	Thr	Asp	Val	Ser	Thr	Ala	Ile	His	Ala	Asp	Gln	Leu	Thr	
			605						610					615		
cca	gct	tg	cgc	ata	tat	tct	act	gga	aac	aat	gta	ttc	cag	act	caa	1984
Pro	Ala	Trp	Arg	Ile	Tyr	Ser	Thr	Gly	Asn	Asn	Val	Phe	Gln	Thr	Gln	
			620						625				630			
gca	ggc	tgt	ctt	ata	gga	gct	gag	cat	gtc	gac	act	tct	tat	gag	tgc	2032
Ala	Gly	Cys	Leu	Ile	Gly	Ala	Glu	His	Val	Asp	Thr	Ser	Tyr	Glu	Cys	
		635					640					645				
gac	att	cct	att	gga	gct	ggc	att	tgt	gct	agt	tac	cat	aca	gtt	tct	2080
Asp	Ile	Pro	Ile	Gly	Ala	Gly	Ile	Cys	Ala	Ser	Tyr	His	Thr	Val	Ser	
	650					655					660					
tta	tta	cgt	agt	act	agc	caa	aaa	tct	att	gtg	gct	tat	act	atg	tct	2128
Leu	Leu	Arg	Ser	Thr	Ser	Gln	Lys	Ser	Ile	Val	Ala	Tyr	Thr	Met	Ser	
	665					670				675					680	
tta	ggg	gct	gat	agt	tca	att	gct	tac	tct	aat	aac	acc	att	gct	ata	2176
Leu	Gly	Ala	Asp	Ser	Ser	Ile	Ala	Tyr	Ser	Asn	Asn	Thr	Ile	Ala	Ile	
			685						690					695		
cct	act	aac	ttt	tca	att	agc	att	act	aca	gaa	gta	atg	cct	gtt	tct	2224

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Pro	Thr	Asn	Phe	Ser	Ile	Ser	Ile	Thr	Thr	Glu	Val	Met	Pro	Val	Ser	
			700					705					710			
atg	gct	aaa	acc	tcc	gta	gat	tgt	aat	atg	tac	atc	tgc	gga	gat	tct	2272
Met	Ala	Lys	Thr	Ser	Val	Asp	Cys	Asn	Met	Tyr	Ile	Cys	Gly	Asp	Ser	
		715					720					725				
act	gaa	tgt	gct	aat	ttg	ctt	ctc	caa	tat	ggt	agc	ttt	tgc	aca	caa	2320
Thr	Glu	Cys	Ala	Asn	Leu	Leu	Leu	Gln	Tyr	Gly	Ser	Phe	Cys	Thr	Gln	
	730					735				740						
cta	aat	cgt	gca	ctc	tca	ggt	att	gct	gct	gaa	cag	gat	cgc	aac	aca	2368
Leu	Asn	Arg	Ala	Leu	Ser	Gly	Ile	Ala	Ala	Glu	Gln	Asp	Arg	Asn	Thr	
	745				750					755				760		
cgt	gaa	gtg	ttc	gct	caa	gtc	aaa	caa	atg	tac	aaa	acc	cca	act	ttg	2416
Arg	Glu	Val	Phe	Ala	Gln	Val	Lys	Gln	Met	Tyr	Lys	Thr	Pro	Thr	Leu	
			765					770						775		
aaa	tat	ttt	ggt	ggt	ttt	aat	ttt	tca	caa	ata	tta	cct	gac	cct	cta	2464
Lys	Tyr	Phe	Gly	Gly	Phe	Asn	Phe	Ser	Gln	Ile	Leu	Pro	Asp	Pro	Leu	
			780					785					790			
aag	cca	act	aag	agg	tct	ttt	att	gag	gac	ttg	ctc	ttt	aat	aag	gtg	2512
Lys	Pro	Thr	Lys	Arg	Ser	Phe	Ile	Glu	Asp	Leu	Leu	Phe	Asn	Lys	Val	
		795					800					805				
aca	ctc	gct	gat	gct	ggc	ttc	atg	aag	caa	tat	ggc	gaa	tgc	cta	ggt	2560
Thr	Leu	Ala	Asp	Ala	Gly	Phe	Met	Lys	Gln	Tyr	Gly	Glu	Cys	Leu	Gly	
	810					815					820					
gat	att	aat	gct	aga	gat	ctc	att	tgt	gcg	cag	aag	ttc	aat	gga	ctt	2608
Asp	Ile	Asn	Ala	Arg	Asp	Leu	Ile	Cys	Ala	Gln	Lys	Phe	Asn	Gly	Leu	
	825					830				835				840		
aca	gtg	ttg	cca	cct	ctg	ctc	act	gat	gat	atg	att	gct	gcc	tac	act	2656
Thr	Val	Leu	Pro	Pro	Leu	Leu	Thr	Asp	Asp	Met	Ile	Ala	Ala	Tyr	Thr	
				845					850					855		
gct	gct	cta	gtt	agt	ggt	act	gcc	act	gct	gga	tgg	aca	ttt	ggt	gct	2704
Ala	Ala	Leu	Val	Ser	Gly	Thr	Ala	Thr	Ala	Gly	Trp	Thr	Phe	Gly	Ala	
		860						865					870			
ggc	gct	gct	ctt	caa	ata	cct	ttt	gct	atg	caa	atg	gca	tat	agg	ttc	2752
Gly	Ala	Ala	Leu	Gln	Ile	Pro	Phe	Ala	Met	Gln	Met	Ala	Tyr	Arg	Phe	
		875				880						885				
aat	ggc	att	gga	gtt	acc	caa	aat	gtt	ctc	tat	gag	aac	caa	aaa	caa	2800
Asn	Gly	Ile	Gly	Val	Thr	Gln	Asn	Val	Leu	Tyr	Glu	Asn	Gln	Lys	Gln	
	890					895					900					
atc	gcc	aac	caa	ttt	aac	aag	gcg	att	agt	caa	att	caa	gaa	tca	ctt	2848
Ile	Ala	Asn	Gln	Phe	Asn	Lys	Ala	Ile	Ser	Gln	Ile	Gln	Glu	Ser	Leu	
	905				910					915				920		
aca	aca	aca	tca	act	gca	ttg	ggc	aag	ctg	caa	gac	gtt	gtt	aac	cag	2896
Thr	Thr	Thr	Ser	Thr	Ala	Leu	Gly	Lys	Leu	Gln	Asp	Val	Val	Asn	Gln	
				925					930					935		
aat	gct	caa	gca	tta	aac	aca	ctt	gtt	aaa	caa	ctt	agc	tct	aat	ttt	2944
Asn	Ala	Gln	Ala	Leu	Asn	Thr	Leu	Val	Lys	Gln	Leu	Ser	Ser	Asn	Phe	
			940					945					950			
ggt	gca	att	tca	agt	gtg	cta	aat	gat	atc	ctt	tcg	cga	ctt	gat	aaa	2992
Gly	Ala	Ile	Ser	Ser	Val	Leu	Asn	Asp	Ile	Leu	Ser	Arg	Leu	Asp	Lys	
		955				960						965				
gtc	gag	gcg	gag	gta	caa	att	gac	agg	tta	att	aca	ggc	aga	ctt	caa	3040
Val	Glu	Ala	Glu	Val	Gln	Ile	Asp	Arg	Leu	Ile	Thr	Gly	Arg	Leu	Gln	
	970					975						980				
agc	ctt	caa	acc	tat	gta	aca	caa	caa	cta	atc	agg	gct	gct	gaa	atc	3088
Ser	Leu	Gln	Thr	Tyr	Val	Thr	Gln	Gln	Leu	Ile	Arg	Ala	Ala	Glu	Ile	
	985				990					995				1000		
agg	gct	tct	gct	aat	ctt	gct	gct	act	aaa	atg	tct	gag	tgt	gtt		3133

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Arg	Ala	Ser	Ala	Asn	Leu	Ala	Ala	Thr	Lys	Met	Ser	Glu	Cys	Val	
				1005					1010					1015	
ctt	gga	caa	tca	aaa	aga	gtt	gac	ttt	tgt	gga	aag	ggc	tac	cac	3178
Leu	Gly	Gln	Ser	Lys	Arg	Val	Asp	Phe	Cys	Gly	Lys	Gly	Tyr	His	
				1020					1025					1030	
ctt	atg	tcc	ttc	cca	caa	gca	gcc	ccg	cat	ggg	gtt	gtc	ttc	cta	3223
Leu	Met	Ser	Phe	Pro	Gln	Ala	Ala	Pro	His	Gly	Val	Val	Phe	Leu	
				1035					1040					1045	
cat	gtc	acg	tat	gtg	cca	tcc	cag	gag	agg	aac	ttc	acc	aca	gcg	3268
His	Val	Thr	Tyr	Val	Pro	Ser	Gln	Glu	Arg	Asn	Phe	Thr	Thr	Ala	
				1050					1055					1060	
cca	gca	att	tgt	cat	gaa	ggc	aaa	gca	tac	ttc	cct	cgt	gaa	ggg	3313
Pro	Ala	Ile	Cys	His	Glu	Gly	Lys	Ala	Tyr	Phe	Pro	Arg	Glu	Gly	
				1065					1070					1075	
gtt	ttt	gtg	ttt	aat	ggc	act	tct	tgg	ttt	att	aca	cag	agg	aac	3358
Val	Phe	Val	Phe	Asn	Gly	Thr	Ser	Trp	Phe	Ile	Thr	Gln	Arg	Asn	
				1080					1085					1090	
ttc	ttt	tct	cca	caa	ata	att	act	aca	gac	aat	aca	ttt	gtc	tca	3403
Phe	Phe	Ser	Pro	Gln	Ile	Ile	Thr	Thr	Asp	Asn	Thr	Phe	Val	Ser	
				1095					1100					1105	
gga	aat	tgt	gat	gtc	gtt	att	ggc	atc	att	aac	aac	aca	gtt	tat	3448
Gly	Asn	Cys	Asp	Val	Val	Ile	Gly	Ile	Ile	Asn	Asn	Thr	Val	Tyr	
				1110					1115					1120	
gat	cct	ctg	caa	cct	gag	ctt	gac	tca	ttc	aaa	gaa	gag	ctg	gac	3493
Asp	Pro	Leu	Gln	Pro	Glu	Leu	Asp	Ser	Phe	Lys	Glu	Glu	Leu	Asp	
				1125					1130					1135	
aag	tac	ttc	aaa	aat	cat	aca	tca	cca	gat	gtt	gat	ctt	ggc	gac	3538
Lys	Tyr	Phe	Lys	Asn	His	Thr	Ser	Pro	Asp	Val	Asp	Leu	Gly	Asp	
				1140					1145					1150	
att	tca	ggc	att	aac	gct	tct	gtc	gtc	aac	att	caa	aaa	gaa	att	3583
Ile	Ser	Gly	Ile	Asn	Ala	Ser	Val	Val	Asn	Ile	Gln	Lys	Glu	Ile	
				1155					1160					1165	
gac	cgc	ctc	aat	gag	gtc	gct	aaa	aat	tta	aat	gaa	tca	ctc	att	3628
Asp	Arg	Leu	Asn	Glu	Val	Ala	Lys	Asn	Leu	Asn	Glu	Ser	Leu	Ile	
				1170					1175					1180	
gac	ctt	caa	gaa	ttg	gga	aaa	tat	gag	caa	tat	att	aaa	tggt	cct	3673
Asp	Leu	Gln	Glu	Leu	Gly	Lys	Tyr	Glu	Gln	Tyr	Ile	Lys	Trp	Pro	
				1185					1190					1195	
tggt	tat	gtt	tggt	ctc	ggc	ttc	att	gct	gga	cta	att	gcc	atc	gtc	3718
Trp	Tyr	Val	Trp	Leu	Gly	Phe	Ile	Ala	Gly	Leu	Ile	Ala	Ile	Val	
				1200					1205					1210	
atg	gtt	aca	atc	ttg	ctt	tgt	tgc	atg	act	agt	tgt	tgc	agt	tgc	3763
Met	Val	Thr	Ile	Leu	Leu	Cys	Cys	Met	Thr	Ser	Cys	Cys	Ser	Cys	
				1215					1220					1225	
ctc	aag	ggg	gca	tgc	tct	tgt	ggg	tct	tgc	tgc	aag	ttt	gat	gag	3808
Leu	Lys	Gly	Ala	Cys	Ser	Cys	Gly	Ser	Cys	Cys	Lys	Phe	Asp	Glu	
				1230					1235					1240	
gat	gac	tct	gag	cca	gtt	ctc	aag	ggg	gtc	aaa	tta	cat	tac	aca	3853
Asp	Asp	Ser	Glu	Pro	Val	Leu	Lys	Gly	Val	Lys	Leu	His	Tyr	Thr	
				1245					1250					1255	
taaacgaact tatggatttg tttatgagat tttttactct tggatcaatt actgcacagc															3913
cagtaaaaat tgacaatgct tctcctgcaa gt															3945

<210> SEQ ID NO 3

<211> LENGTH: 1255

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 3

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20      25      30
His Thr Ser Ser Met Arg Gly Val Tyr Tyr Pro Asp Glu Ile Phe Arg
35      40      45
Ser Asp Thr Leu Tyr Leu Thr Gln Asp Leu Phe Leu Pro Phe Tyr Ser
50      55      60
Asn Val Thr Gly Phe His Thr Ile Asn His Thr Phe Gly Asn Pro Val
65      70      75      80
Ile Pro Phe Lys Asp Gly Ile Tyr Phe Ala Ala Thr Glu Lys Ser Asn
85      90      95
Val Val Arg Gly Trp Val Phe Gly Ser Thr Met Asn Asn Lys Ser Gln
100     105     110
Ser Val Ile Ile Ile Asn Asn Ser Thr Asn Val Val Ile Arg Ala Cys
115     120     125
Asn Phe Glu Leu Cys Asp Asn Pro Phe Phe Ala Val Ser Lys Pro Met
130     135     140
Gly Thr Gln Thr His Thr Met Ile Phe Asp Asn Ala Phe Asn Cys Thr
145     150     155     160
Phe Glu Tyr Ile Ser Asp Ala Phe Ser Leu Asp Val Ser Glu Lys Ser
165     170     175
Gly Asn Phe Lys His Leu Arg Glu Phe Val Phe Lys Asn Lys Asp Gly
180     185     190
Phe Leu Tyr Val Tyr Lys Gly Tyr Gln Pro Ile Asp Val Val Arg Asp
195     200     205
Leu Pro Ser Gly Phe Asn Thr Leu Lys Pro Ile Phe Lys Leu Pro Leu
210     215     220
Gly Ile Asn Ile Thr Asn Phe Arg Ala Ile Leu Thr Ala Phe Ser Pro
225     230     235     240
Ala Gln Asp Ile Trp Gly Thr Ser Ala Ala Ala Tyr Phe Val Gly Tyr
245     250     255
Leu Lys Pro Thr Thr Phe Met Leu Lys Tyr Asp Glu Asn Gly Thr Ile
260     265     270
Thr Asp Ala Val Asp Cys Ser Gln Asn Pro Leu Ala Glu Leu Lys Cys
275     280     285
Ser Val Lys Ser Phe Glu Ile Asp Lys Gly Ile Tyr Gln Thr Ser Asn
290     295     300
Phe Arg Val Val Pro Ser Gly Asp Val Val Arg Phe Pro Asn Ile Thr
305     310     315     320
Asn Leu Cys Pro Phe Gly Glu Val Phe Asn Ala Thr Lys Phe Pro Ser
325     330     335
Val Tyr Ala Trp Glu Arg Lys Lys Ile Ser Asn Cys Val Ala Asp Tyr
340     345     350
Ser Val Leu Tyr Asn Ser Thr Phe Phe Ser Thr Phe Lys Cys Tyr Gly
355     360     365
Val Ser Ala Thr Lys Leu Asn Asp Leu Cys Phe Ser Asn Val Tyr Ala
370     375     380
Asp Ser Phe Val Val Lys Gly Asp Asp Val Arg Gln Ile Ala Pro Gly

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385	390	395	400
Gln Thr Gly Val Ile Ala Asp Tyr Asn Tyr Lys Leu Pro Asp Asp Phe	405	410	415
Met Gly Cys Val Leu Ala Trp Asn Thr Arg Asn Ile Asp Ala Thr Ser	420	425	430
Thr Gly Asn Tyr Asn Tyr Lys Tyr Arg Tyr Leu Arg His Gly Lys Leu	435	440	445
Arg Pro Phe Glu Arg Asp Ile Ser Asn Val Pro Phe Ser Pro Asp Gly	450	455	460
Lys Pro Cys Thr Pro Pro Ala Leu Asn Cys Tyr Trp Pro Leu Asn Asp	465	470	475
Tyr Gly Phe Tyr Thr Thr Thr Gly Ile Gly Tyr Gln Pro Tyr Arg Val	485	490	495
Val Val Leu Ser Phe Glu Leu Leu Asn Ala Pro Ala Thr Val Cys Gly	500	505	510
Pro Lys Leu Ser Thr Asp Leu Ile Lys Asn Gln Cys Val Asn Phe Asn	515	520	525
Phe Asn Gly Leu Thr Gly Thr Gly Val Leu Thr Pro Ser Ser Lys Arg	530	535	540
Phe Gln Pro Phe Gln Gln Phe Gly Arg Asp Val Ser Asp Phe Thr Asp	545	550	555
Ser Val Arg Asp Pro Lys Thr Ser Glu Ile Leu Asp Ile Ser Pro Cys	565	570	575
Ser Phe Gly Gly Val Ser Val Ile Thr Pro Gly Thr Asn Ala Ser Ser	580	585	590
Glu Val Ala Val Leu Tyr Gln Asp Val Asn Cys Thr Asp Val Ser Thr	595	600	605
Ala Ile His Ala Asp Gln Leu Thr Pro Ala Trp Arg Ile Tyr Ser Thr	610	615	620
Gly Asn Asn Val Phe Gln Thr Gln Ala Gly Cys Leu Ile Gly Ala Glu	625	630	635
His Val Asp Thr Ser Tyr Glu Cys Asp Ile Pro Ile Gly Ala Gly Ile	645	650	655
Cys Ala Ser Tyr His Thr Val Ser Leu Leu Arg Ser Thr Ser Gln Lys	660	665	670
Ser Ile Val Ala Tyr Thr Met Ser Leu Gly Ala Asp Ser Ser Ile Ala	675	680	685
Tyr Ser Asn Asn Thr Ile Ala Ile Pro Thr Asn Phe Ser Ile Ser Ile	690	695	700
Thr Thr Glu Val Met Pro Val Ser Met Ala Lys Thr Ser Val Asp Cys	705	710	715
Asn Met Tyr Ile Cys Gly Asp Ser Thr Glu Cys Ala Asn Leu Leu Leu	725	730	735
Gln Tyr Gly Ser Phe Cys Thr Gln Leu Asn Arg Ala Leu Ser Gly Ile	740	745	750
Ala Ala Glu Gln Asp Arg Asn Thr Arg Glu Val Phe Ala Gln Val Lys	755	760	765
Gln Met Tyr Lys Thr Pro Thr Leu Lys Tyr Phe Gly Gly Phe Asn Phe	770	775	780
Ser Gln Ile Leu Pro Asp Pro Leu Lys Pro Thr Lys Arg Ser Phe Ile	785	790	795
			800

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Glu Asp Leu Leu Phe Asn Lys Val Thr Leu Ala Asp Ala Gly Phe Met
 805 810 815
 Lys Gln Tyr Gly Glu Cys Leu Gly Asp Ile Asn Ala Arg Asp Leu Ile
 820 825 830
 Cys Ala Gln Lys Phe Asn Gly Leu Thr Val Leu Pro Pro Leu Leu Thr
 835 840 845
 Asp Asp Met Ile Ala Ala Tyr Thr Ala Ala Leu Val Ser Gly Thr Ala
 850 855 860
 Thr Ala Gly Trp Thr Phe Gly Ala Gly Ala Ala Leu Gln Ile Pro Phe
 865 870 875 880
 Ala Met Gln Met Ala Tyr Arg Phe Asn Gly Ile Gly Val Thr Gln Asn
 885 890 895
 Val Leu Tyr Glu Asn Gln Lys Gln Ile Ala Asn Gln Phe Asn Lys Ala
 900 905 910
 Ile Ser Gln Ile Gln Glu Ser Leu Thr Thr Thr Ser Thr Ala Leu Gly
 915 920 925
 Lys Leu Gln Asp Val Val Asn Gln Asn Ala Gln Ala Leu Asn Thr Leu
 930 935 940
 Val Lys Gln Leu Ser Ser Asn Phe Gly Ala Ile Ser Ser Val Leu Asn
 945 950 955 960
 Asp Ile Leu Ser Arg Leu Asp Lys Val Glu Ala Glu Val Gln Ile Asp
 965 970 975
 Arg Leu Ile Thr Gly Arg Leu Gln Ser Leu Gln Thr Tyr Val Thr Gln
 980 985 990
 Gln Leu Ile Arg Ala Ala Glu Ile Arg Ala Ser Ala Asn Leu Ala Ala
 995 1000 1005
 Thr Lys Met Ser Glu Cys Val Leu Gly Gln Ser Lys Arg Val Asp
 1010 1015 1020
 Phe Cys Gly Lys Gly Tyr His Leu Met Ser Phe Pro Gln Ala Ala
 1025 1030 1035
 Pro His Gly Val Val Phe Leu His Val Thr Tyr Val Pro Ser Gln
 1040 1045 1050
 Glu Arg Asn Phe Thr Thr Ala Pro Ala Ile Cys His Glu Gly Lys
 1055 1060 1065
 Ala Tyr Phe Pro Arg Glu Gly Val Phe Val Phe Asn Gly Thr Ser
 1070 1075 1080
 Trp Phe Ile Thr Gln Arg Asn Phe Phe Ser Pro Gln Ile Ile Thr
 1085 1090 1095
 Thr Asp Asn Thr Phe Val Ser Gly Asn Cys Asp Val Val Ile Gly
 1100 1105 1110
 Ile Ile Asn Asn Thr Val Tyr Asp Pro Leu Gln Pro Glu Leu Asp
 1115 1120 1125
 Ser Phe Lys Glu Glu Leu Asp Lys Tyr Phe Lys Asn His Thr Ser
 1130 1135 1140
 Pro Asp Val Asp Leu Gly Asp Ile Ser Gly Ile Asn Ala Ser Val
 1145 1150 1155
 Val Asn Ile Gln Lys Glu Ile Asp Arg Leu Asn Glu Val Ala Lys
 1160 1165 1170
 Asn Leu Asn Glu Ser Leu Ile Asp Leu Gln Glu Leu Gly Lys Tyr
 1175 1180 1185

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Glu	Gln	Tyr	Ile	Lys	Trp	Pro	Trp	Tyr	Val	Trp	Leu	Gly	Phe	Ile
1190						1195					1200			
Ala	Gly	Leu	Ile	Ala	Ile	Val	Met	Val	Thr	Ile	Leu	Leu	Cys	Cys
1205						1210					1215			
Met	Thr	Ser	Cys	Cys	Ser	Cys	Leu	Lys	Gly	Ala	Cys	Ser	Cys	Gly
1220						1225					1230			
Ser	Cys	Cys	Lys	Phe	Asp	Glu	Asp	Asp	Ser	Glu	Pro	Val	Leu	Lys
1235						1240					1245			
Gly	Val	Lys	Leu	His	Tyr	Thr								
1250						1255								

<210> SEQ ID NO 4

<211> LENGTH: 3943

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 4

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gtggtagtga ccttgaccgg tgcaccactt ttgatgatgt tcaagctcct aattacactc    180
aacatacttc atctatgagg ggggtttact atcctgatga aatttttaga tcagacactc    240
tttatttaac tcaggattta tttcttccat tttattctaa tgttacaggg tttcatacta    300
ttaatcatac gtttggcaac cctgtcatac cttttaagga tggattttat tttgctgcca    360
cagagaaatc aaatgttgtc cgtgggtggg tttttggttc taccatgaac aacaagtcac    420
agtcggtgat tattattaac aattctacta atgttggtat acgagcatgt aactttgaat    480
tgtgtgacaa ccttttcttt gctgtttcta aacctatggg tacacagaca catactatga    540
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gagccattct tacagccttt tcacctgctc aagacatttg gggcacgtca gctgcagcct    840
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caactggtaa ttataattat aaatataggt atcttagaca tggcaagctt aggccctttg   1440
agagagacat atctaattgt cctttctccc ctgatggcaa accttgcacc ccacctgctc   1500
ttaattgtta ttggccatta aatgattatg gtttttacac cactactggc attggctacc   1560
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gccgtgatgt ctctgatttc actgattccg ttcgagatcc taaaacatct gaaatattag	1800
acatttcacc ttgctctttt ggggggtgtaa gtgtaattac acctggaaca aatgcttcat	1860
ctgaagttgc tgttctatat caagatgtta actgcactga tgtttctaca gcaatccatg	1920
cagatcaact cacaccagct tggcgcatat attctactgg aaacaatgta ttccagactc	1980
aagcagcgtg tcttatagga gctgagcatg tcgacacttc ttatgagtgc gacattccta	2040
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caggcattaa cgtctctgtc gtcaacattc aaaaagaaat tgaccgcctc aatgaggtcg	3600
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gtggttcttg ctgcaagttt gatgaggatg actctgagcc agttctcaag ggtgtcaaat	3840
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<210> SEQ ID NO 5

<211> LENGTH: 2049

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 5

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gtggtagtga ccttgaccgg tgcaccactt ttgatgatgt tcaagctcct aattacactc 180
aacatacttc atctatgagg ggggtttact atcctgatga aatttttaga tcagacactc 240
tttatttaac tcaggattta tttcttccat tttattctaa tgttacaggg tttcatacta 300
ttaatcatac gtttggcaac cctgtcatac cttttaagga tggatttat tttgctgcca 360
cagagaaatc aaatgttgtc cgtgggtggg tttttggttc taccatgaac aacaagtcac 420
agtcggtgat tattattaac aattctacta atgttggtat acgagcatgt aactttgaat 480
tgtgtgacaa ccctttcttt gctgtttcta aacctatggg tacacagaca catactatga 540
tattcgataa tgcatttaat tgcactttcg agtacatato tcatgccttt tcgcttgatg 600
tttcagaaaa gtcaggtaat tttaaacact tacgagagtt tgtgtttaaa aataaagatg 660
ggtttctcta tgtttataag ggctatcaac ctatagatgt agttcgtgat ctaccttctg 720
gttttaacac tttgaaacct atttttaagt tgcctcttgg tattaacatt acaaatttta 780
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aaccttacag agttgtagta tttctttttg aacttttaaa tgcaccggcc acggttttgtg 1620
gaccaaaatt atccactgac cttattaaga accagtgtgt caattttaat tttaatggac 1680
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ctgaagttgc tgttctatat caagatgtta actgcactga tgtttctaca gcaatccatg 1920
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<210> SEQ ID NO 6
<211> LENGTH: 2027
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 6

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cctattggag ctggcatttg tgctagtac catacagttt ctttattacg tagtactagc 180
caaaaatcta ttgtggctta tactatgtct ttaggtgctg atagtccaat tgcttactct 240
aataacacca ttgctatacc tactaacttt tcaattagca ttactacaga agtaatgcct 300
gtttctatgg ctaaaacctc cgtagattgt aatatgtaca tctgcggaga ttctactgaa 360
tgtgctaatt tgcttctcca atatggtagc ttttgcacac aactaaatcg tgcactctca 420
ggattgctg ctgaacagga tcgcaacaca cgtgaagtgt tcgctcaagt caaacaatg 480
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gctgatgctg gcttcatgaa gcaatatggc gaatgcctag gtgatattaa tgctagagat 660
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atggttacaa tcttgcttgg ttgcatgact agttgttgca gttgcctcaa gggtgcatgc 1860
tcttgtgggt cttgctgcaa gtttgatgag gatgactctg agccagttct caagggtgtc 1920

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caattactgc acagccagta aaaattgaca atgcttctcc tgcaagt 2027

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<210> SEQ ID NO 7
<211> LENGTH: 1096
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 7

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acacataaac gaacttatgg atttgtttat gagatTTTTT actcttggat caattactgc 180
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accgctacaa gcctcactcc ctttcggatg gcttggtatt ggcgttgcat ttcttgctgt 300
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caagaacca ttactttatg atgccaacta ctttgtttgc tggcacacac ataactatga 600
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<210> SEQ ID NO 8
<211> LENGTH: 1135
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 8

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ctcaagggtg tcaattaca ttacacataa acgaacttat ggatttggtt atgagatttt 180
ttactcttgg atcaattact gcacagccag taaaattga caatgcttct cctgcaagta 240
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ttaccatcta ttacatctt ttgcttgctg ctgcaggatg ggaggcgcaa tttttgtacc 480
tctatgcctt gatataattt ctacaatgca tcaacgcatg tagaattatt atgagatgtt 540

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tcgttactga aggtgacggc atttcaacac caaaactcaa agaagactac caaattgggtg 720
gttattctga ggataggcac tcaggtgtta aagactatgt cgttgtacat ggctatttca 780
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tcgacggctc ttcaggagtt gctaattccag caatggatcc aatttatgat gagccgacga 960
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tttcggaaga aacaggtagc ttaatatgta atagcgtact tctttttctt gctttcgtgg 1080
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<210> SEQ ID NO 9
<211> LENGTH: 1096
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (137)..(958)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 9

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acacataaac gaactt atg gat ttg ttt atg aga ttt ttt act ctt gga tca 172
      Met Asp Leu Phe Met Arg Phe Phe Thr Leu Gly Ser
      1             5             10
att act gca cag cca gta aaa att gac aat gct tct cct gca agt act 220
Ile Thr Ala Gln Pro Val Lys Ile Asp Asn Ala Ser Pro Ala Ser Thr
      15             20             25
gtt cat gct aca gca acg ata ccg cta caa gcc tca ctc cct ttc gga 268
Val His Ala Thr Ala Thr Ile Pro Leu Gln Ala Ser Leu Pro Phe Gly
      30             35             40
tgg ctt gtt att ggc gtt gca ttt ctt gct gtt ttt cag agc gct acc 316
Trp Leu Val Ile Gly Val Ala Phe Leu Ala Val Phe Gln Ser Ala Thr
      45             50             55             60
aaa ata att gcg ctc aat aaa aga tgg cag cta gcc ctt tat aag ggc 364
Lys Ile Ile Ala Leu Asn Lys Arg Trp Gln Leu Ala Leu Tyr Lys Gly
      65             70             75
ttc cag ttc att tgc aat tta ctg ctg cta ttt gtt acc atc tat tca 412
Phe Gln Phe Ile Cys Asn Leu Leu Leu Leu Phe Val Thr Ile Tyr Ser
      80             85             90
cat ctt ttg ctt gtc gct gca ggt atg gag gcg caa ttt ttg tac ctc 460
His Leu Leu Leu Val Ala Ala Gly Met Glu Ala Gln Phe Leu Tyr Leu
      95             100             105
tat gcc ttg ata tat ttt cta caa tgc atc aac gca tgt aga att att 508
Tyr Ala Leu Ile Tyr Phe Leu Gln Cys Ile Asn Ala Cys Arg Ile Ile
      110             115             120
atg aga tgt tgg ctt tgt tgg aag tgc aaa tcc aag aac cca tta ctt 556
Met Arg Cys Trp Leu Cys Trp Lys Cys Lys Ser Lys Asn Pro Leu Leu
      125             130             135             140
tat gat gcc aac tac ttt gtt tgc tgg cac aca cat aac tat gac tac 604
Tyr Asp Ala Asn Tyr Phe Val Cys Trp His Thr His Asn Tyr Asp Tyr
      145             150             155

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gac ggc att tca aca cca aaa ctc aaa gaa gac tac caa att ggt ggt Asp Gly Ile Ser Thr Pro Lys Leu Lys Glu Asp Tyr Gln Ile Gly Gly 175 180 185	700
tat tct gag gat agg cac tca ggt gtt aaa gac tat gtc gtt gta cat Tyr Ser Glu Asp Arg His Ser Gly Val Lys Asp Tyr Val Val Val His 190 195 200	748
ggc tat ttc acc gaa gtt tac tac cag ctt gag tct aca caa att act Gly Tyr Phe Thr Glu Val Tyr Tyr Gln Leu Glu Ser Thr Gln Ile Thr 205 210 215 220	796
aca gac act ggt att gaa aat gct aca ttc ttc atc ttt aac aag ctt Thr Asp Thr Gly Ile Glu Asn Ala Thr Phe Phe Ile Phe Asn Lys Leu 225 230 235	844
gtt aaa gac cca ccg aat gtg caa ata cac aca atc gac ggc tct tca Val Lys Asp Pro Pro Asn Val Gln Ile His Thr Ile Asp Gly Ser Ser 240 245 250	892
gga gtt gct aat cca gca atg gat cca att tat gat gag ccg acg acg Gly Val Ala Asn Pro Ala Met Asp Pro Ile Tyr Asp Glu Pro Thr Thr 255 260 265	940
act act agc gtg cct ttg taagcacaag aaagttagta cgaacttatg Thr Thr Ser Val Pro Leu 270	988
tactcattcg ttccggaaga aacaggtacg ttaatagtta atagcgtact tctttttctt	1048
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<210> SEQ ID NO 10
 <211> LENGTH: 274
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 10

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Ala Thr Ile Pro Leu Gln Ala Ser Leu Pro Phe Gly Trp Leu Val Ile 35 40 45
Gly Val Ala Phe Leu Ala Val Phe Gln Ser Ala Thr Lys Ile Ile Ala 50 55 60
Leu Asn Lys Arg Trp Gln Leu Ala Leu Tyr Lys Gly Phe Gln Phe Ile 65 70 75 80
Cys Asn Leu Leu Leu Leu Phe Val Thr Ile Tyr Ser His Leu Leu Leu 85 90 95
Val Ala Ala Gly Met Glu Ala Gln Phe Leu Tyr Leu Tyr Ala Leu Ile 100 105 110
Tyr Phe Leu Gln Cys Ile Asn Ala Cys Arg Ile Ile Met Arg Cys Trp 115 120 125
Leu Cys Trp Lys Cys Lys Ser Lys Asn Pro Leu Leu Tyr Asp Ala Asn 130 135 140
Tyr Phe Val Cys Trp His Thr His Asn Tyr Asp Tyr Cys Ile Pro Tyr 145 150 155 160
Asn Ser Val Thr Asp Thr Ile Val Val Thr Glu Gly Asp Gly Ile Ser

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Ser Leu Thr Ser Leu Leu Lys Thr His Arg Met Cys Lys Tyr Thr Gln	
95 100 105	
tcg acg gct ctt cag gag ttg cta atc cag caa tgg atc caa ttt atg	926
Ser Thr Ala Leu Gln Glu Leu Leu Ile Gln Gln Trp Ile Gln Phe Met	
110 115 120	
atg agc cga cga cga cta cta gcg tgc ctt tgt aag cac aag aaa gtg	974
Met Ser Arg Arg Arg Leu Leu Ala Cys Leu Cys Lys His Lys Lys Val	
125 130 135	
agt acg aac tta tgt act cat tcg ttt cgg aag aaa cag gta cgt	1019
Ser Thr Asn Leu Cys Thr His Ser Phe Arg Lys Lys Gln Val Arg	
140 145 150	
taatagttaa tagcgtaactt ctttttcttg ctttcgtggt attcttgcta gtcacactag	1079
ccatccttac tgcgctt	1096

<210> SEQ ID NO 12
 <211> LENGTH: 154
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 12

Met Met Pro Thr Thr Leu Phe Ala Gly Thr His Ile Thr Met Thr Thr	
1 5 10 15	
Val Tyr His Ile Thr Val Ser Gln Ile Gln Leu Ser Leu Leu Lys Val	
20 25 30	
Thr Ala Phe Gln His Gln Asn Ser Lys Lys Thr Thr Lys Leu Val Val	
35 40 45	
Ile Leu Arg Ile Gly Thr Gln Val Leu Lys Thr Met Ser Leu Tyr Met	
50 55 60	
Ala Ile Ser Pro Lys Phe Thr Thr Ser Leu Ser Leu His Lys Leu Leu	
65 70 75 80	
Gln Thr Leu Val Leu Lys Met Leu His Ser Ser Ser Leu Thr Ser Leu	
85 90 95	
Leu Lys Thr His Arg Met Cys Lys Tyr Thr Gln Ser Thr Ala Leu Gln	
100 105 110	
Glu Leu Leu Ile Gln Gln Trp Ile Gln Phe Met Met Ser Arg Arg Arg	
115 120 125	
Leu Leu Ala Cys Leu Cys Lys His Lys Lys Val Ser Thr Asn Leu Cys	
130 135 140	
Thr His Ser Phe Arg Lys Lys Gln Val Arg	
145 150	

<210> SEQ ID NO 13
 <211> LENGTH: 332
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (36)..(263)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 13

tgcctttgta agcacaagaa agtgagtacg aactt atg tac tca ttc gtt tcg	53
Met Tyr Ser Phe Val Ser	
1 5	
gaa gaa aca ggt acg tta ata gtt aat agc gta ctt ctt ttt ctt gct	101
Glu Glu Thr Gly Thr Leu Ile Val Asn Ser Val Leu Leu Phe Leu Ala	

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10	15	20	
ttc gtg gta ttc ttg cta gtc aca cta gcc atc ctt act gcg ctt cga			149
Phe Val Val Phe Leu Leu Val Thr Leu Ala Ile Leu Thr Ala Leu Arg			
25	30	35	
ttg tgt gcg tac tgc tgc aat att gtt aac gtg agt tta gta aaa cca			197
Leu Cys Ala Tyr Cys Cys Asn Ile Val Asn Val Ser Leu Val Lys Pro			
40	45	50	
acg gtt tac gtc tac tcg cgt gtt aaa aat ctg aac tct tct gaa gga			245
Thr Val Tyr Val Tyr Ser Arg Val Lys Asn Leu Asn Ser Ser Glu Gly			
55	60	65	70
gtt cct gat ctt ctg gtc taaacgaact aactattatt attattctgt			293
Val Pro Asp Leu Leu Val			
75			
ttggaacttt aacattgctt atcatggcag acaacgga			332

<210> SEQ ID NO 14
 <211> LENGTH: 76
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 14

Met Tyr Ser Phe Val Ser Glu Glu Thr Gly Thr Leu Ile Val Asn Ser	
1	15
Val Leu Leu Phe Leu Ala Phe Val Val Phe Leu Leu Val Thr Leu Ala	
20	30
Ile Leu Thr Ala Leu Arg Leu Cys Ala Tyr Cys Cys Asn Ile Val Asn	
35	45
Val Ser Leu Val Lys Pro Thr Val Tyr Val Tyr Ser Arg Val Lys Asn	
50	60
Leu Asn Ser Ser Glu Gly Val Pro Asp Leu Leu Val	
65	75

<210> SEQ ID NO 15
 <211> LENGTH: 332
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 15

tgcctttgta agcacaagaa agtgagtacg aacttatgta ctcattcggt tcggaagaaa	60
caggtagctt aatagttaat agcgtacttc tttttcttgc tttcgtggtt ttcttgctag	120
tcacactagc catccttact gcgcttcgat tgtgtgcgta ctgctgcaat attgttaacg	180
tgagtttagt aaaaccaacg gtttacgtct actcgcgtgt taaaaatctg aactcttctg	240
aaggagttcc tgatcttctg gtctaaacga actaactatt attattattc tgtttggaac	300
tttaacattg cttatcatgg cagacaacgg ta	332

<210> SEQ ID NO 16
 <211> LENGTH: 708
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (41)..(703)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 16

tattattatt attctgtttg gaactttaac attgcttatt atg gca gac aac ggt	55
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Met Ala Asp Asn Gly Thr Ile Thr Val Glu Glu Leu Lys Gln Leu Leu
1 5 10 15

Glu Gln Trp Asn Leu Val Ile Gly Phe Leu Phe Leu Ala Trp Ile Met
20 25 30

Leu Leu Gln Phe Ala Tyr Ser Asn Arg Asn Arg Phe Leu Tyr Ile Ile
35 40 45

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Lys Leu Val Phe Leu Trp Leu Leu Trp Pro Val Thr Leu Ala Cys Phe
 50 55 60
 Val Leu Ala Ala Val Tyr Arg Ile Asn Trp Val Thr Gly Gly Ile Ala
 65 70 75 80
 Ile Ala Met Ala Cys Ile Val Gly Leu Met Trp Leu Ser Tyr Phe Val
 85 90 95
 Ala Ser Phe Arg Leu Phe Ala Arg Thr Arg Ser Met Trp Ser Phe Asn
 100 105 110
 Pro Glu Thr Asn Ile Leu Leu Asn Val Pro Leu Arg Gly Thr Ile Val
 115 120 125
 Thr Arg Pro Leu Met Glu Ser Glu Leu Val Ile Gly Ala Val Ile Ile
 130 135 140
 Arg Gly His Leu Arg Met Ala Gly His Ser Leu Gly Arg Cys Asp Ile
 145 150 155 160
 Lys Asp Leu Pro Lys Glu Ile Thr Val Ala Thr Ser Arg Thr Leu Ser
 165 170 175
 Tyr Tyr Lys Leu Gly Ala Ser Gln Arg Val Gly Thr Asp Ser Gly Phe
 180 185 190
 Ala Ala Tyr Asn Arg Tyr Arg Ile Gly Asn Tyr Lys Leu Asn Thr Asp
 195 200 205
 His Ala Gly Ser Asn Asp Asn Ile Ala Leu Leu Val Gln
 210 215 220

<210> SEQ ID NO 18

<211> LENGTH: 769

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 18

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cctgatcttc tggctctaaac gaactaacta ttattattat tctgtttgga actttaacat    60
tgcttatcat ggcagacaac ggtactatta ccgttgagga gcttaaacia ctccctggaac    120
aatggaacct agtaataggt ttcctattcc tagcctggat tatgttacta caatttgcct    180
attctaactg gaacaggttt ttgtacataa taaagcttgt tttcctctgg ctcttgtggc    240
cagtaacact tgcttgtttt gtgcttgctg ctgtctacag aattaattgg gtgactggcg    300
ggattgcgat tgcaatggct tgtattgtag gcttgatgtg gcttagctac ttcgttgctt    360
ccttcaggct gtttgcctgt acccgctcaa tgtggctatt caaccagaa acaaacatto    420
ttctcaatgt gcctctccgg gggacaattg tgaccagacc gctcatggaa agtgaacttg    480
tcattggtgc tgtgatcatt cgtggctact tgcgaatggo cggacactcc ctaggcgct    540
gtgacattaa ggacctgcca aaagagatca ctgtggctac atcacgaacg ctttcttatt    600
acaaattagg agcgtcgag cgtgtaggca ctgattcagg ttttgctgca tacaaccgct    660
accgtattgg aaactataaa ttaaatacag accacgccgg tagcaacgac aatattgctt    720
tgctagtaca gtaagtgaca acagatgttt catcttgttg acttocagg                769

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<210> SEQ ID NO 19

<211> LENGTH: 1231

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 19

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taccgtattg gaaactataa attaaataca gaccacgccg gtagcaacga caatattgct    60

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ttgctagtagac agtaagtac aacagatggt tcattctgtt gacttccagg ttacaatagc	120
agagatatgt attatcatta tgaggacttt caggattgct atttggaaac ttgacgttat	180
aataagttca atagttagac aattatttaa gcctctaact aagaagaatt attcggagtt	240
agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct	300
tcctgacatt gattgtattt acattctgag agctatatca ctatcaggag tgtgttagag	360
gtacgactgt actactaaaa gaaccttgcc catcaggaa atacgagggc aattcaccat	420
ttcaccctct tgctgacaat aaatttgac taacttgac tagcacacac ttgcttttg	480
cttggtgta cggtagctga catacctatc agctgcgtgc aagatcagtt tcacaaaac	540
ttttcatcag acaaggagg gttcaacaag agctctact gccacttttt ctattgttg	600
ctgcttagt atttttaata cttgtctta ccattaagag aaagacagaa tgaatgagct	660
cactttaatt gacttctatt tgtgttttt agcctttctg ctattccttg ttttaataat	720
gcttattata ttttggttt cactcgaaat ccaggatcta gaagaacct gtacaaaagt	780
ctaaacgaac atgaaacttc tcattgtttt gacttgatt tctctatgca gttgcatatg	840
cactgtagta cagcgctgtg catctaata acctcatgtg cttgaagatc cttgtaaggt	900
acaacactag gggtaatact tatagcactg cttggctttg tgctctagga aagggtttac	960
ctttcatag atggcacact atggttcaaa catgcacacc taatgttact atcaactgtc	1020
aagatccagc tgggtgtgag cttatagcta ggtgttgta ccttcagaa ggtcacaaa	1080
ctgctgcatt tagagacgta cttgtgttt taaataaacg aacaaattaa aatgtctgat	1140
aatggacccc aatcaaacca acgtagtgcc ccccgatta catttggtgg acccacagat	1200
tcaactgaca ataaccagaa tggaggacgc a	1231

<210> SEQ ID NO 20

<211> LENGTH: 1242

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 20

gcatacaacc gctaccgtat tggaaactat aaattaaata cagaccacgc cggtagcaac	60
gacaatatgt ctttgtagt acagtaagt acaacagatg tttcatcttg ttgacttcca	120
ggttacaata gcagagatat tgattatcat tatgaggact ttcaggattg ctatttgga	180
tcttgacgtt ataataagtt caatagttag acagttattt aagcctctaa ctaagaagaa	240
ttattcggag ttagatgatg aagaacctat ggagtttag tatccataaa acgaacatga	300
aaattattct cttcctgaca ttgattgtat ttacatcttg cgagctatat cactatcagg	360
agtgtgttag aggtacgact gtactactaa aagaaccttg cccatcagga acatacgagg	420
gcaattcacc atttcacct cttgctgaca ataaatttgc actaacttgc actagcacac	480
actttgcttt tgettgtgtg gacggtagt gacataccta tcagctgctg gcaagatcag	540
tttcacaaa acttttcac agacaagagg aggttcaaca agagctctac tcgccacttt	600
ttctcattgt tgctgctcta gtatttttaa tactttgctt caccattaag agaaagacag	660
aatgaatgag ctacttttaa ttgacttcta tttgtgcttt ttagcctttc tgctattcct	720
tgttttaata atgcttatta tattttggtt ttactcga atccaggatc tagaagaacc	780
ttgtacaaa gtctaaacga acatgaaact tctcattgtt ttgacttgta tttctctatg	840

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cagttgcata tgcactgtag tacagcgctg tgcactctaat aaacctcatg tgcttgaaga    900
tccttgtaag gtacaacact aggggtaata cttatagcac tgcttggcctt tgtgctctag    960
gaaagggtttt accttttcat agatggcaca ctatggttca aacatgcaca cctaattgta 1020
ctatcaactg tcaagatcca gctgggtggtg cgcttatagc taggtgttgg taccttcatg 1080
aaggtcacca aactgctgca tttagagacg tacttgttgt tttaaataaa cgaacgaatt 1140
aaaatgtctg ataatggacc ccaatcaaac caacgtagtg cccccgcat tacatttgg 1200
ggacccacag attcaactga caataaccag aatggaggac gc 1242

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<210> SEQ ID NO 21
<211> LENGTH: 1231
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (86)..(274)
<223> OTHER INFORMATION:

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<400> SEQUENCE: 21

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taccgtattg gaaactataa attaaataca gaccacgccg gtagcaacga caatattgct    60
ttgctagtag agtaagtac aacag atg ttt cat ctt gtt gac ttc cag gtt    112
                               Met Phe His Leu Val Asp Phe Gln Val
                               1                               5
aca ata gca gag ata ttg att atc att atg agg act ttc agg att gct    160
Thr Ile Ala Glu Ile Leu Ile Ile Ile Met Arg Thr Phe Arg Ile Ala
10                               15                               20                               25
att tgg aat ctt gac gtt ata ata agt tca ata gtg aga caa tta ttt    208
Ile Trp Asn Leu Asp Val Ile Ile Ser Ser Ile Val Arg Gln Leu Phe
30                               35                               40
aag cct cta act aag aag aat tat tcg gag tta gat gat gaa gaa cct    256
Lys Pro Leu Thr Lys Lys Asn Tyr Ser Glu Leu Asp Asp Glu Glu Pro
45                               50                               55
atg gag tta gat tat cca taaaacgaac atgaaaatta ttctcttcct    304
Met Glu Leu Asp Tyr Pro
60
gacattgatt gtattttacat cttgcgagct atatcactat caggagtgtg ttagaggtag    364
gactgtacta ctaaaagaac cttgccatc aggaacatac gagggcaatt caccatttca    424
ccctcttgct gacaataaat ttgcactaac ttgcactagc acacactttg cttttgcttg    484
tgctgacggt actcgacata cctatcagct gcgtgcaaga tcagtttcac caaaactttt    544
catcagacaa gaggaggttc aacaagagct ctactcgcca ctttttctca ttgttgctgc    604
tctagtatth ttaatacttt gcttcacat taagagaaag acagaatgaa tgagctcact    664
ttaattgact tctatttgtg ctttttagcc tttctgctat tccttgtttt aataatgctt    724
attatatttt gggttttcaat cgaaatccag gatctagaag aaccttgtag caaagtctaa    784
acgaacatga aactttctcat tgttttgact tgtattttct tatgcagttg catatgcact    844
gtagtacagc gctgtgcac taataaacct catgtgcttg aagatccttg taaggtacaa    904
cactaggggt aatacttata gcaactgctg gctttgtgct ctaggaaagg ttttaccttt    964
tcatagatgg cacactatgg ttcaaactg cacacctaata gttactatca actgtcaaga 1024
tccagctggt ggtgcgctta tagctaggtg ttggtacctt catgaaggtc accaaactgc 1084
tgcatthtag gacgtacttg ttgttttaaa taaacgaaca aattaaaatg tctgataatg 1144

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gacccaatc aaaccaacgt agtgccccc gcattacatt tggtaggaccc acagattcaa 1204
 ctgacaataa ccagaatgga ggacgca 1231

<210> SEQ ID NO 22
 <211> LENGTH: 63
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 22

Met Phe His Leu Val Asp Phe Gln Val Thr Ile Ala Glu Ile Leu Ile
 1 5 10 15
 Ile Ile Met Arg Thr Phe Arg Ile Ala Ile Trp Asn Leu Asp Val Ile
 20 25 30
 Ile Ser Ser Ile Val Arg Gln Leu Phe Lys Pro Leu Thr Lys Lys Asn
 35 40 45
 Tyr Ser Glu Leu Asp Asp Glu Glu Pro Met Glu Leu Asp Tyr Pro
 50 55 60

<210> SEQ ID NO 23
 <211> LENGTH: 1231
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (285)..(650)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 23

taccgtattg gaaactataa attaaatata gaccacgccg gtagcaacga caatattgct 60
 ttgctagtag agtaagttag aacagatggt tcatcttggt gacttccagg ttacaatagc 120
 agagatatg attatcatta tgaggacttt caggattgct atttggatc ttgacgttat 180
 aataagttca atagttagac aattatttaa gcctctaact aagaagaatt attcgaggtt 240
 agatgatgaa gaacctatgg agtttagatta tccataaaac gaac atg aaa att att 296
 Met Lys Ile Ile
 1
 ctc ttc ctg aca ttg att gta ttt aca tct tgc gag cta tat cac tat 344
 Leu Phe Leu Thr Leu Ile Val Phe Thr Ser Cys Glu Leu Tyr His Tyr
 5 10 15 20
 cag gag tgt gtt aga ggt acg act gta cta cta aaa gaa cct tgc cca 392
 Gln Glu Cys Val Arg Gly Thr Thr Val Leu Leu Lys Glu Pro Cys Pro
 25 30 35
 tca gga aca tac gag ggc aat tca cca ttt cac cct ctt gct gac aat 440
 Ser Gly Thr Tyr Glu Gly Asn Ser Pro Phe His Pro Leu Ala Asp Asn
 40 45 50
 aaa ttt gca cta act tgc act agc aca cac ttt gct ttt gct tgt gct 488
 Lys Phe Ala Leu Thr Cys Thr Ser Thr His Phe Ala Phe Ala Cys Ala
 55 60 65
 gac ggt act cga cat acc tat cag ctg cgt gca aga tca gtt tca cca 536
 Asp Gly Thr Arg His Thr Tyr Gln Leu Arg Ala Arg Ser Val Ser Pro
 70 75 80
 aaa ctt ttc atc aga caa gag gag gtt caa caa gag ctc tac tcg cca 584
 Lys Leu Phe Ile Arg Gln Glu Glu Val Gln Gln Glu Leu Tyr Ser Pro
 85 90 95 100
 ctt ttt ctc att gtt gct gct cta gta ttt tta ata ctt tgc ttc acc 632
 Leu Phe Leu Ile Val Ala Ala Leu Val Phe Leu Ile Leu Cys Phe Thr
 105 110 115

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att aag aga aag aca gaa tgaatgagct cactttaatt gacttctatt      680
Ile Lys Arg Lys Thr Glu
      120

tgtgcttttt agcctttctg ctattccttg ttttaataat gcttattata ttttggtttt      740
cactcgaaat ccaggatcta gaagaacctt gtaccaaagt ctaaacgaac atgaaacttc      800
tcattgtttt gacttgtatt tctctatgca gttgcatatg cactgtagta cagcgctgtg      860
catctaataa acctcatgtg cttgaagatc cttgtaaggt acaacactag gggtaatact      920
tatagcactg cttggctttg tgctctagga aagggttttac cttttcatag atggcacact      980
atggttcaaa catgcacacc taatgttact atcaactgtc aagatccagc tgggtgtgctg      1040
cttatagcta ggtgttggtta ctttcatgaa ggtcaccaaa ctgctgcatt tagagacgta      1100
cttggtgttt taaataaacg aacaaattaa aatgtctgat aatggacccc aatcaaacca      1160
acgtagtgcc ccccgcatca catttggtgg acccacagat tcaactgaca ataaccagaa      1220
tggaggacgc a                                                    1231

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<210> SEQ ID NO 24
 <211> LENGTH: 122
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 24

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Met Lys Ile Ile Leu Phe Leu Thr Leu Ile Val Phe Thr Ser Cys Glu
1           5           10          15

Leu Tyr His Tyr Gln Glu Cys Val Arg Gly Thr Thr Val Leu Leu Lys
      20           25           30

Glu Pro Cys Pro Ser Gly Thr Tyr Glu Gly Asn Ser Pro Phe His Pro
      35           40           45

Leu Ala Asp Asn Lys Phe Ala Leu Thr Cys Thr Ser Thr His Phe Ala
      50           55           60

Phe Ala Cys Ala Asp Gly Thr Arg His Thr Tyr Gln Leu Arg Ala Arg
      65           70           75           80

Ser Val Ser Pro Lys Leu Phe Ile Arg Gln Glu Glu Val Gln Gln Glu
      85           90           95

Leu Tyr Ser Pro Leu Phe Leu Ile Val Ala Ala Leu Val Phe Leu Ile
      100          105          110

Leu Cys Phe Thr Ile Lys Arg Lys Thr Glu
      115          120

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<210> SEQ ID NO 25
 <211> LENGTH: 1231
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (650)..(781)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 25

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taccgtattg gaaactataa attaaatata gaccacgccg gtagcaacga caatattgct      60
ttgctagtac agtaagtac aacagatggt tcattctgtt gacttccagg ttacaatagc      120
agagatatgt attatcatta tgaggacttt caggattgct atttggaaac ttgacgttat      180
aataagttca atagttagac aattatttaa gcctctaact aagaagaatt attcggagtt      240

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agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct 300
tcctgacatt gattgtattht acatcttgcg agctatatca ctatcaggag tgtgttagag 360
gtacgactgt actactaaaa gaaccttgcc catcaggaac atacgagggc aattcaccat 420
ttcaccctct tgctgacaat aaatttgcac taacttgcac tagcacacac ttgcttttg 480
cttgtgctga cggtactcga catacctatc agctgcgtgc aagatcagtt tcacaaaaac 540
ttttcatcag acaagaggag gttcaacaag agctctactc gccacttttt ctcatgttg 600
ctgctctagt atttttaata ctttgcttca ccattaagag aaagacaga atg aat gag 658
                               Met Asn Glu
                               1
ctc act tta att gac ttc tat ttg tgc ttt tta gcc ttt ctg cta ttc 706
Leu Thr Leu Ile Asp Phe Tyr Leu Cys Phe Leu Ala Phe Leu Leu Phe
  5                10                15

ctt gtt tta ata atg ctt att ata ttt tgg ttt tca ctc gaa atc cag 754
Leu Val Leu Ile Met Leu Ile Ile Phe Trp Phe Ser Leu Glu Ile Gln
 20                25                30                35

gat cta gaa gaa cct tgt acc aaa gtc taaacgaaca tgaaacttct 801
Asp Leu Glu Glu Pro Cys Thr Lys Val
 40

cattgttttg acttgattht ctctatgcag ttgcatatgc actgtagtac agcgtgtgac 861
atctaataaaa cctcatgtgc ttgaagatcc ttgtaaggta caacactagg ggtaataactt 921
atagcactgc ttggcttttg gctctaggaa aggttttacc ttttcataga tggcacacta 981
tgggtcaaac atgcacacct aatgttacta tcaactgtca agatccagct ggtggtgagc 1041
ttatagctag gtgttggtac cttcatgaag gtcaccaaac tgctgcattt agagacgtac 1101
ttgttgtttt aaataaacga acaaattaaa atgtctgata atggacccca atcaaaccaa 1161
cgtagtgccc cccgcattac atttggtgga cccacagatt caactgacaa taaccagaat 1221
ggaggacgca 1231

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<210> SEQ ID NO 26
 <211> LENGTH: 44
 <212> TYPE: PRT
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 26

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Met Asn Glu Leu Thr Leu Ile Asp Phe Tyr Leu Cys Phe Leu Ala Phe
1                5                10                15

Leu Leu Phe Leu Val Leu Ile Met Leu Ile Ile Phe Trp Phe Ser Leu
 20                25                30

Glu Ile Gln Asp Leu Glu Glu Pro Cys Thr Lys Val
 35                40

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<210> SEQ ID NO 27
 <211> LENGTH: 1231
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (791)..(907)
 <223> OTHER INFORMATION:

<400> SEQUENCE: 27

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taccgtattg gaaactataa attaaataca gaccacgccc gtagcaacga caatattgct 60

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ttgctagtagac agtaagtgac aacagatggtt tcatcttggt gacttccagg ttacaatagc	120
agagatatattg attatcatta tgaggacttt caggattgct atttggaaac ttgacgttat	180
aataagttca atagttagac aattatttaa gcctctaact aagaagaatt attcggagtt	240
agatgatgaa gaacctatgg agttagatta tccataaaac gaacatgaaa attattctct	300
tcctgacatt gattgtatgt acatcttgcg agctatatca ctatcaggag tgtgttagag	360
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<211> LENGTH: 39

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 28

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Asp Pro Cys Lys Val Gln His
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Ala Leu Gly Lys Val Leu Pro Phe His Arg Trp His Thr Met Val Gln
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Thr Cys Thr Pro Asn Val Thr Ile Asn Cys Gln Asp Pro Ala Gly Gly
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<211> LENGTH: 84

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 30

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Trp His Thr Met Val Gln Thr Cys Thr Pro Asn Val Thr Ile Asn Cys
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<213> ORGANISM: CORONAVIRUS

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Gln Gly Gln Asn Ser Ala Asp Pro Lys Val Tyr Pro Ile Ile Leu Arg
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Leu Gly Ser Gln Leu Ser Leu Ser Met Ala Arg Arg Asn Leu Asp Ser
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Leu Glu Ala Arg Ala Phe Gln Ser Thr Pro Ile Val Val Gln Met Thr
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Lys Leu Ala Thr Thr Glu Glu Leu Pro Asp Glu Phe Val Val Val Thr
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Ala Lys

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Pro His His Val Val Ala Val Ile Gln Glu Ile Gln Leu Leu Ala Ala
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50 55 60

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gag gaa ctt aga ttc cct cga ggc cag ggc gtt cca atc aac acc aat Glu Glu Leu Arg Phe Pro Arg Gly Gln Gly Val Pro Ile Asn Thr Asn 65 70 75	300
agt ggt cca gat gac caa att ggc tac tac cga aga gct acc cga cga Ser Gly Pro Asp Asp Gln Ile Gly Tyr Tyr Arg Arg Ala Thr Arg Arg 80 85 90	348
gtt cgt ggt ggt gac ggc aaa atg aaa gag ctc agc ccc aga tgg tac Val Arg Gly Gly Asp Gly Lys Met Lys Glu Leu Ser Pro Arg Trp Tyr 95 100 105 110	396
ttc tat tac cta gga act ggc cca gaa gct tca ctt ccc tac ggc gct Phe Tyr Tyr Leu Gly Thr Gly Pro Glu Ala Ser Leu Pro Tyr Gly Ala 115 120 125	444
aac aaa gaa ggc atc gta tgg gtt gca act gag gga gcc ttg aat aca Asn Lys Glu Gly Ile Val Trp Val Ala Thr Glu Gly Ala Leu Asn Thr 130 135 140	492
ccc aaa gac cac att ggc acc cgc aat cct aat aac aat gct gcc acc Pro Lys Asp His Ile Gly Thr Arg Asn Pro Asn Asn Ala Ala Thr 145 150 155	540
gtg cta caa ctt cct caa gga aca aca ttg cca aaa ggc ttc tac gca Val Leu Gln Leu Pro Gln Gly Thr Thr Leu Pro Lys Gly Phe Tyr Ala 160 165 170	588
gag gga agc aga ggc ggc agt caa gcc tct tct cgc tcc tca tca cgt Glu Gly Ser Arg Gly Gly Ser Gln Ala Ser Ser Arg Ser Ser Ser Arg 175 180 185 190	636
agt cgc ggt aat tca aga aat tca act cct ggc agc agt agg gga aat Ser Arg Gly Asn Ser Arg Asn Ser Thr Pro Gly Ser Ser Arg Gly Asn 195 200 205	684
tct cct gct cga atg gct agc gga ggt ggt gaa act gcc ctc gcg cta Ser Pro Ala Arg Met Ala Ser Gly Gly Gly Glu Thr Ala Leu Ala Leu 210 215 220	732
ttg ctg cta gac aga ttg aac cag ctt gag agc aaa gtt tct ggt aaa Leu Leu Leu Asp Arg Leu Asn Gln Leu Glu Ser Lys Val Ser Gly Lys 225 230 235	780
ggc caa caa caa caa ggc caa act gtc act aag aaa tct gct gct gag Gly Gln Gln Gln Gln Gly Gln Thr Val Thr Lys Lys Ser Ala Ala Glu 240 245 250	828
gca tct aaa aag cct cgc caa aaa cgt act gcc aca aaa cag tac aac Ala Ser Lys Lys Pro Arg Gln Lys Arg Thr Ala Thr Lys Gln Tyr Asn 255 260 265 270	876
gtc act caa gca ttt ggg aga cgt ggt cca gaa caa acc caa gga aat Val Thr Gln Ala Phe Gly Arg Arg Gly Pro Glu Gln Thr Gln Gly Asn 275 280 285	924
ttc ggg gac caa gac cta atc aga caa gga act gat tac aaa cat tgg Phe Gly Asp Gln Asp Leu Ile Arg Gln Gly Thr Asp Tyr Lys His Trp 290 295 300	972
ccg caa att gca caa ttt gct cca agt gcc tct gca ttc ttt gga atg Pro Gln Ile Ala Gln Phe Ala Pro Ser Ala Ser Ala Phe Phe Gly Met 305 310 315	1020
tca cgc att ggc atg gaa gtc aca cct tcg gga aca tgg ctg act tat Ser Arg Ile Gly Met Glu Val Thr Pro Ser Gly Thr Trp Leu Thr Tyr 320 325 330	1068
cat gga gcc att aaa ttg gat gac aaa gat cca caa ttc aaa gac aac His Gly Ala Ile Lys Leu Asp Asp Lys Asp Pro Gln Phe Lys Asp Asn 335 340 345 350	1116
gtc ata ctg ctg aac aag cac att gac gca tac aaa aca ttc cca cca Val Ile Leu Leu Asn Lys His Ile Asp Ala Tyr Lys Thr Phe Pro Pro 355 360 365	1164

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aca gag cct aaa aag gac aaa aag aaa aag act gat gaa gct cag cct    1212
Thr Glu Pro Lys Lys Asp Lys Lys Lys Lys Thr Asp Glu Ala Gln Pro
          370                      375                      380

ttg ccg cag aga caa aag aag cag ccc act gtg act ctt ctt cct gcg    1260
Leu Pro Gln Arg Gln Lys Lys Gln Pro Thr Val Thr Leu Leu Pro Ala
          385                      390                      395

gct gac atg gat gat ttc tcc aga caa ctt caa aat tcc atg agt gga    1308
Ala Asp Met Asp Asp Phe Ser Arg Gln Leu Gln Asn Ser Met Ser Gly
          400                      405                      410

gct tct gct gat tca act cag gca taa acactcatga tgaccacaca    1355
Ala Ser Ala Asp Ser Thr Gln Ala
415                      420

aggcagatgg gctatgtaaa cg    1377

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<210> SEQ ID NO 37
<211> LENGTH: 422
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 37

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Met Ser Asp Asn Gly Pro Gln Ser Asn Gln Arg Ser Ala Pro Arg Ile
1          5          10          15

Thr Phe Gly Gly Pro Thr Asp Ser Thr Asp Asn Asn Gln Asn Gly Gly
20          25          30

Arg Asn Gly Ala Arg Pro Lys Gln Arg Arg Pro Gln Gly Leu Pro Asn
35          40          45

Asn Thr Ala Ser Trp Phe Thr Ala Leu Thr Gln His Gly Lys Glu Glu
50          55          60

Leu Arg Phe Pro Arg Gly Gln Gly Val Pro Ile Asn Thr Asn Ser Gly
65          70          75          80

Pro Asp Asp Gln Ile Gly Tyr Tyr Arg Arg Ala Thr Arg Arg Val Arg
85          90          95

Gly Gly Asp Gly Lys Met Lys Glu Leu Ser Pro Arg Trp Tyr Phe Tyr
100         105         110

Tyr Leu Gly Thr Gly Pro Glu Ala Ser Leu Pro Tyr Gly Ala Asn Lys
115         120         125

Glu Gly Ile Val Trp Val Ala Thr Glu Gly Ala Leu Asn Thr Pro Lys
130         135         140

Asp His Ile Gly Thr Arg Asn Pro Asn Asn Asn Ala Ala Thr Val Leu
145         150         155         160

Gln Leu Pro Gln Gly Thr Thr Leu Pro Lys Gly Phe Tyr Ala Glu Gly
165         170         175

Ser Arg Gly Gly Ser Gln Ala Ser Ser Arg Ser Ser Ser Arg Ser Arg
180         185         190

Gly Asn Ser Arg Asn Ser Thr Pro Gly Ser Ser Arg Gly Asn Ser Pro
195         200         205

Ala Arg Met Ala Ser Gly Gly Gly Glu Thr Ala Leu Ala Leu Leu Leu
210         215         220

Leu Asp Arg Leu Asn Gln Leu Glu Ser Lys Val Ser Gly Lys Gly Gln
225         230         235         240

Gln Gln Gln Gly Gln Thr Val Thr Lys Lys Ser Ala Ala Glu Ala Ser
245         250         255

Lys Lys Pro Arg Gln Lys Arg Thr Ala Thr Lys Gln Tyr Asn Val Thr

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260	265	270
Gln Ala Phe Gly Arg Arg Gly Pro Glu Gln Thr Gln Gly Asn Phe Gly		
275	280	285
Asp Gln Asp Leu Ile Arg Gln Gly Thr Asp Tyr Lys His Trp Pro Gln		
290	295	300
Ile Ala Gln Phe Ala Pro Ser Ala Ser Ala Phe Phe Gly Met Ser Arg		
305	310	315
Ile Gly Met Glu Val Thr Pro Ser Gly Thr Trp Leu Thr Tyr His Gly		
325	330	335
Ala Ile Lys Leu Asp Asp Lys Asp Pro Gln Phe Lys Asp Asn Val Ile		
340	345	350
Leu Leu Asn Lys His Ile Asp Ala Tyr Lys Thr Phe Pro Pro Thr Glu		
355	360	365
Pro Lys Lys Asp Lys Lys Lys Lys Thr Asp Glu Ala Gln Pro Leu Pro		
370	375	380
Gln Arg Gln Lys Lys Gln Pro Thr Val Thr Leu Leu Pro Ala Ala Asp		
385	390	395
Met Asp Asp Phe Ser Arg Gln Leu Gln Asn Ser Met Ser Gly Ala Ser		
405	410	415
Ala Asp Ser Thr Gln Ala		
420		

<210> SEQ ID NO 38

<211> LENGTH: 1377

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 38

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atgaaggtca ccaactgct gcatttagag acgtacttgt tgttttaaat aaacgaacaa    60
attaaatgt ctgataatgg accccaatca aaccaacgta gtgcccccg cattacattt    120
ggtggacca cagattcaac tgacaataac cagaatggag gacgcaatgg ggcaaggcca    180
aaacagcgcc gacccaagg tttaccaat aatactgcgt cttggttcac agctctcact    240
cagcatggca aggaggaact tagattccct cgaggccagg gcgttccaat caacaccaat    300
agtgggccag atgaccaaatt tggctactac cgaagagcta ccgacgagtg tcgtggtggt    360
gacggcaaaa tgaaagagct cagcccaga tggctacttct attacctagg aactggccca    420
gaagcttcac ttccctacgg cgctaacaaa gaaggcatcg tatgggtgac aactgagggg    480
gccttgaata caccacaaga ccacattggc acccgcaatc ctaataacaa tgctgccacc    540
gtgctacaac ttctcaagg aacaacattg ccaaaaggct tctacgcaga gggaagcaga    600
ggcggcagtc aagcctcttc tcgctcctca tcacgtagtc gcggtaatc aagaaattca    660
actcctggca gcagtagggg aaattctcct gctcgaatgg ctacggaggg tggtgaaact    720
gccctcgccg tattgtctgt agacagattg aaccagcttg agagcaaatg ttctggtaaa    780
ggccaacaac aacaaggcca aactgtcact aagaaatctg ctgctgaggc atctaaaaag    840
cctcgccaaa aacgtactgc caaaaacag tacaacgtca ctcaagcatt tgggagacgt    900
ggtccagaac aaaccaagg aaatttcggg gaccaagacc taatcagaca aggaactgat    960
tacaacatt ggccgcaaatt tgcacaattt gctccaagtg cctctgcatt ctttggaatg   1020
tcacgcattg gcatggaagt cacaccttcg ggaacatggo tgacttatca tggagccatt   1080

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aaattggatg acaaagatcc acaattcaaa gacaacgtca tactgctgaa caagcacatt	1140
gacgcataca aaacattccc accaacagag cctaaaaagg acaaaaagaa aaagactgat	1200
gaagctcagc ctttgccgca gagacaaaag aagcagccca ctgtgactct tcttcctgcg	1260
gctgacatgg atgatttctc cagacaactt caaaattcca tgagtggagc ttctgctgat	1320
tcaactcagg cataaacact catgatgacc acacaaggca gatgggctat gtaaacy	1377

<210> SEQ ID NO 39
 <211> LENGTH: 204
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 39

atattaggtt ttacctacc caggaaaagc caaccaacct cgatctcttg tagatctgtt	60
ctctaaacga actttaaaat ctgtgtagct gtcgctcggc tgcagccta gtgcacctac	120
gcagtataaa caataataaa ttttactgtc gttgacaaga aacgagtaac tcgtccctct	180
tctgcagact gcttacgggt tcgt	204

<210> SEQ ID NO 40
 <211> LENGTH: 809
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 40

actcaagcat ttgggagacg tggtcagaa caaacccaag gaaatttcgg ggaccaagac	60
ctaactagac aaggaactga ttacaaacat tggccgcaaa ttgcacaatt tgctccaagt	120
gcctctgcat tctttggaat gtcacgcatt ggcatggaag tcacaccttc ggaacatgg	180
ctgacttatac atggagccat taaattggat gacaaagatc cacaattcaa agacaacgtc	240
atactgtga acaagcacat tgacgcatac aaaacattcc caccaacaga gcctaaaaag	300
gacaaaaaga aaaagactga tgaagctcag cttttgccgc agagacaaaa gaagcagccc	360
actgtgactc ttcttcctgc ggctgacatg gatgatttct ccagacaact tcaaaattcc	420
atgagtggag cttctgctga ttcaactcag gcataaacac tcatgatgac cacacaaggc	480
agatgggcta tgtaaacgtt ttgcgaattc cgtttacgat acatagtcta ctcttgtgca	540
gaatgaattc tcgtaactaa acagcacaag taggtttagt taactttaat ctcacatagc	600
aatctttaat caatgtgtaa cattagggag gacttgaaag agccaccaca ttttcacgca	660
ggccacgcgc agtacgatc aggggtacagt gaataatgct agggagagct gcctatatgg	720
aagagcccta atgtgtaaaa ttaattttag tagtgctatc cccatgtgat tttaatagct	780
tcttaggaga atgacaaaaa aaaaaaaaaa	809

<210> SEQ ID NO 41
 <211> LENGTH: 448
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 41

aatgaacaca tagggctgtt caagctgggg cagtacgcct ttttcagct ctactagacc	60
acaagtgccca tttttgaggt gttcacgtgc ctccgatagg gcctcttcca cagagtcgcc	120
gaagccacgc actagcacgt ctctaacctg aaggacaggc aaactgagtt ggacgtgtgt	180

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tttctcgttg acaccaagaa caaggctctc catcttacct ttcggtcaca cccggacgaa	240
acctaggtat gctgatgato gactgcaaca cggacgaaac cgtaagcagt ctgcagaaga	300
gggacgagtt actcgtttct tgtcaacgac agtaaaattt attattgttt atactgcgta	360
ggtgcactag gcatgcagcc gagcgacagc tacacagatt ttaaagttcg tttagagaac	420
agatctacaa gagatcgagg ttggttgg	448

<210> SEQ ID NO 42

<211> LENGTH: 2033

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 42

atacctaggt ttcgtccggg tgtgaccgaa aggtaagatg gagagccttg ttcttggtgt	60
caacgagaaa acacacgtcc aactcagttt gcctgtcctt caggttagag acgtgctagt	120
gcgtggcttc ggggactctg tggaagaggc cctatcggag gcacgtgaac acctcaaaaa	180
tggcacttgt ggtctagttag agctggaaaa aggcgtactg ccccgacttg aacagcccta	240
tgtgttcatt aaacgtttct atgccttaag caccaatcac ggccacaagg tcgttgagct	300
ggttgcagaa atggacggca ttcagtacgg tcgtagcggg ataacttg gagtactcgt	360
gccacatgtg ggcgaaaccc caattgcata ccgcaatgtt cttcttcgta agaacggtaa	420
taaggagacc ggtggtcata gctatggcat cgatctaaag tcttatgact taggtgacga	480
gcttggcact gatcccatgt aagattatga acaaaactgg aacactaagc atggcagtgg	540
tgcactccgt gaactcactc gtgagctcaa tggaggtgca gtcactcgct atgtcgacaa	600
caatttctgt ggcccagatg ggtaccctct tgattgcac aaagattttc tcgcacgcgc	660
gggcaagtca atgtgcactc tttccgaaca acttgattac atcgagtcga agagaggtgt	720
ctactgctgc cgtgaccatg agcatgaaat tgcctggttc actgagcgct ctgataagag	780
ctacgagcac cagacaccct tcgaaattaa gagtgcgaag aaatttgaca ctttcaaagg	840
ggaatgcccc aagtttgtgt ttcctcttaa ctcaaaagtc aaagtcattc aaccacgtgt	900
tgaaaagaaa aagactgagg gtttcattgg gcgtatacgc tctgtgtacc ctgttgcatc	960
tccacaggag tgtaacaata tgcacttgtc taccttgatg aaatgtaac attgcgatga	1020
agtttcatgg cagacgtgcg actttctgaa agccacttgt gaacattgtg gcactgaaaa	1080
tttagttatt gaaggaccta ctacatgtgg gtacctacct actaatgctg tagtgaaaat	1140
gccatgtcct gcctgtcaag acccagagat tggacctgag catagtgttg cagattatca	1200
caaccactca aacattgaaa ctcgactccg caaggagggt aggactagat gttttggagg	1260
ctgtgtgttt gcctatgttg gctgtataaa taagcgtgcc tactgggttc ctgctgctag	1320
tgtgtatatt ggctcaggcc atactggcat tactggtgac aatgtggaga ccttgaatga	1380
ggatctcctt gagatactga gtcgtgaacg tgtaacatt aacattgttg gcgattttca	1440
tttgaatgaa gaggttgcca tcattttggc atctttctct gctttacaa gtgcctttat	1500
tgacactata aagagtcttg attacaagtc tttcaaaacc attgttgagt cctgcggtaa	1560
ctataaagtt accaaggga agcccgtaaa aggtgcttgg aacattggac aacagagatc	1620
agttttaaca ccactgtgtg gttttccctc acaggctgct ggtgttatca gatcaatttt	1680
tgcgcgacac cttgatgcag caaaccactc aattcctgat ttgcaaagag cagctgtcac	1740

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catacttgat ggtatttctg aacagtcatt acgtcttgto gacgccatgg tttatacttc	1800
agacctgctc accaacagtg tcattattat ggcatatgta actggtggto ttgtacaaca	1860
gacttctcag tggttgtcta atcttttggg cactactgtt gaaaaactca ggcctatctt	1920
tgaatggatt gaggcgaaac ttagtgcagg agttgaattt ctcaaggatg cttgggagat	1980
tctcaaattt ctcattacag gtgtttttga catcgtcaag ggtcaaatac agg	2033

<210> SEQ ID NO 43

<211> LENGTH: 2018

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 43

ggattgaggc gaaacttagt gcaggagttg aatttctcaa ggatgcttg gagattctca	60
aatttctcat tacagggtgt tttgacatcg tcaagggtca aatacagggt gcttcagata	120
acatcaagga ttgtgtaaaa tgcttcattg atgttggtta caaggcactc gaaatgtgca	180
ttgatcaagt cactatcgct ggcgcgaaagt tgcgatcact caacttaggt gaagtcttca	240
tcgctcaaag caagggactt taccgtcagt gtatacgtgg caaggagcag ctgcaactac	300
tcatgcctct taaggcacca aaagaagtaa cctttcttga aggtgattca catgacacag	360
tacttacctc tgaggagggt gttctcaaga acggtgaact cgaagcactc gagacgcccg	420
ttgatagctt cacaaatgga gctatcgttg gcacaccagt ctgtgtaaat ggctcatgc	480
tcttagagat taaggacaaa gaacaatact gcgcattgto tccgtgttta ctggctacaa	540
acaatgtctt tcgcttaaaa ggggggtgcac caattaaagg tgtaaccttt ggagaagata	600
ctgtttggga agttcaagggt tacaagaatg tgagaatcac atttgagctt gatgaacgtg	660
ttgacaaagt gcttaatgaa aagtgtctct tctacactgt tgaatccggt accgaagtta	720
ctgagtttgc atgtgttgta gcaggagctg ttgtgaagac tttaacacca gtttctgac	780
tccttaccaa catgggtatt gatcttgatg agtggagtgt agctacattc tacttatttg	840
atgatgctgg tgaagaaaac ttttcatcac gtatgtattg ttccttttac cctccagatg	900
aggaagaaga ggacgatgca gagtgtgagg aagaagaaat tgatgaaacc tgtgaacatg	960
agtacggtac agaggatgat tatcaaggtc tccctctgga atttggtgcc tcagctgaaa	1020
cagttcgagt tgaggaagaa gaagaggaag actggctgga tgatactact gagcaatcag	1080
agattgagcc agaaccagaa cctacacctg aagaaccagt taatcagttt actggttatt	1140
taaaacttac tgacaatgtt gccattaaat gtgttgacat cgtaaggag gcacaaagtg	1200
ctaactctat ggtgattgta aatgctgcta acatacacct gaaacatggg ggtggtgtag	1260
cagggtgact caacaaggca accaatgggt ccatgcaaaa ggagagtgat gattacatta	1320
agctaaatgg ccctcttaca gtaggagggt cttgtttgct ttctggacat aatcttgcta	1380
agaagtgtct gcattgtgtt ggacctaac taaatgcagg tgaggacatc cagcttctta	1440
aggcagcata tgaattttt aattcacagg acatcttact tgcaccattg ttgtcagcag	1500
gcataatttg tgctaaacca cttcagcttt tacaagtgtg cgtgcagacg gttcgtacac	1560
aggtttatat tgacgtcaat gacaaagctc tttatgagca ggtgtcatg gattatcttg	1620
ataacctgaa gcctagagtg gaagcaccta aacaagagga gccaccaaac acagaagatt	1680
cacaaactga ggagaaatct gtcgtacaga agcctgtcga tgtgaagcca aaaattaagg	1740

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cctgcattga tgaggttacc acaacactgg aagaaactaa gtttcttacc aataagttac	1800
tcttgtttgc tgatatcaat ggtaagcttt accatgattc tcagaacatg cttagagggtg	1860
aagatatgtc tttccttgag aaggatgcac cttacatggt aggtgatgtt atcactagtgt	1920
gtgatatcac ttgtgttgta ataccctcca aaaaggctgg tggcactact gagatgctct	1980
caagagcttt gaagaaagtg ccagttgatg agtatata	2018

<210> SEQ ID NO 44
 <211> LENGTH: 1442
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 44

ttgatgaggt taccacaaca ctggaagaaa ctaagtttct taccaataag ttactcttgt	60
ttgctgatat caatggtaag ctttaccatg attctcagaa catgcttaga ggtgaagata	120
tgtctttcct tgagaaggat gcaccttaca tggtaggtga tgttatcact agtggtgata	180
tcacttgtgt tgtaataccc tccaaaaagg ctggtggcac tactgagatg ctctcaagag	240
ctttgaagaa agtgccagtt gatgagtata taaccacgta ccctggacaa ggatgtgctg	300
gttatcact tgaggaagct aagactgctc ttaagaaatg caaatctgca ttttatgtac	360
taccttcaga agcacctaag gctaaggaa agattctagg aactgtatcc tggaatttga	420
gagaaatgct tgctcatgct gaagagacaa gaaaattaat gcctatatgc atggatgtta	480
gagccataat ggcaaccatc caacgtaagt ataaaggaat taaaattcaa gagggcatcg	540
ttgactatgg tgtccgattc ttcttttata ctagttaaaga gcctgtagct tctattatta	600
cgaagctgaa ctctctaaat gagccgcttg tcacaatgcc aattggttat gtgacacatg	660
gttttaatct tgaagaggct gcgcgctgta tgcgttctct taaagctcct gccgtagtgt	720
cagtatcatc accagatgct gttactacat ataatggata cctcacttcg tcaccaaaga	780
catctgagga gcactttgta gaaacagttt ctttggtggt ctcttacaga gattggtcct	840
attcaggaca gcgtacagag ttaggtgttg aatttcttaa gcgtggtgac aaaattgtgt	900
accacactct ggagagcccc gtcgagtttc atcttgacgg tgaggttcct tcacttgaca	960
aactaaagag tctcttatcc ctgcgggagg ttaagactat aaaagtgttc acaactgtgg	1020
acaacactaa tctccacaca cagcttggtg atatgtctat gacatatgga cagcagtttg	1080
gtccaacata cttggatggt gctgatgta caaaaattaa acctcatgta aatcatgagg	1140
gtaagacttt ctttgtacta cctagtgatg acacactacg tagtgaagct ttcgagtact	1200
accatactct tgatgagagt tttcttggtg ggtacatgct tgctttaaac cacacaaaga	1260
aatggaaatt tcctcaagtt ggtgggttaa cttcaattaa atgggctgat aacaattgtt	1320
atttgtctag tgttttatta gcacttcaac agcttgaagt caaattcaat gcaccagcac	1380
ttcaagaggc ttattataga gcccgctgct gtgatgctgc taacttttgt gcactcatac	1440
tc	1442

<210> SEQ ID NO 45
 <211> LENGTH: 1050
 <212> TYPE: DNA
 <213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 45

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atatgtctat gacatatgga cagcagtttg gtccaacata cttggatggt gctgatgta	60
caaaaattaa acctcatgta aatcatgagg gtaagacttt ctttgtacta cctagtgatg	120
acacactacg tagtgaagct ttcgagtact accatactct tgatgagagt tttcttggtta	180
ggtacatgtc tgctttaaac cacacaaaga aatggaaatt tcctcaagtt ggtggtttaa	240
cttcaattaa atgggctgat aacaattggt atttgtctag tgttttatta gcacttcaac	300
agcttgaagt caaattcaat gcaccagcac ttcaagaggc ttattataga gcccggtgtg	360
gtgatgtgc taacttttgt gcactcatac tcgcttacag taataaaact gttggcgcgc	420
ttggtgatgt cagagaaact atgacccatc ttctacagca tgctaatttg gaatctgcaa	480
agcgagttct taatgtggtg tgtaaacatt gtggtcagaa aactactacc ttaacgggtg	540
tagaagctgt gatgtatatg ggtactctat cttatgataa tcttaagaca ggtgtttcca	600
ttccatgtgt gtgtggtcgt gatgctacac aatatctagt acaacaagag tcttcttttg	660
ttatgatgtc tgcaccacct gctgagtata aattacagca aggtacattc ttatgtgcga	720
atgagtacac tggtaactat cagtgtgggc attacactca tataactgct aaggagaccc	780
tctatcgtat tgacggagct caccttaca agatgtcaga gtacaaagga ccagtgaactg	840
atgttttcta caaggaaaca tcttactaca caaccatcaa gcctgtgtcg tataaactcg	900
atggagtac ttacacagag attgaaccaa aattggatgg gtattataaa aaggataatg	960
cttactatac agagcagcct atagacctg taccaactca accattacca aatgcgagtt	1020
ttgataattt caaactcaca tgttctaaca	1050

<210> SEQ ID NO 46

<211> LENGTH: 1995

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 46

tttgtgcact catactcgct tacagtaata aaactgttgg cgagcttggg gatgtcagag	60
aaactatgac ccatcttcta cagcatgcta atttgaatc tgcaaagcga gttcttaatg	120
tggtgtgtaa acattgtggg cagaaaaacta ctaccttaac ggggtgagaa gctgtgatgt	180
atatgggtac tctatcttat gataatctta agacaggtgt ttccattcca tgtgtgtgtg	240
gtcgtgatgc tacacaatat ctagtacaac aagagtcttc ttttgttatg atgtctgcac	300
cacctgctga gtataaatta cagcaaggta cattcttatg tgcgaatgag tacactggta	360
actatcagtg tggtcattac actcatataa ctgctaagga gacctctat cgtattgacg	420
gagctcacct tacaaagatg tcagagtaca aaggaccagt gactgatgtt ttctacaagg	480
aaacatctta cactacaacc atcaagcctg tgcgtataa actcgatgga gttacttaca	540
cagagattga accaaaattg gatgggtatt ataaaaagga taatgcttac tatacagagc	600
agcctataga ccttgtacca actcaacct taccaaatgc gagttttgat aatttcaaac	660
tcacatgttc taacacaaaa tttgctgatg atttaaatca aatgacaggc ttacaaaagc	720
cagcttcacg agagctatct gtcacattct tcccagactt gaattggcgat gtagtggcta	780
ttgactatag acactatttc gcgagtttca agaaagggtc taaattactg cataagccaa	840
ttgtttggca cattaaccag gctacaacca agacaacgtt caaaccaaac acttggtgtt	900
tacgttgtct ttggagtaca aagccagtag atacttcaaa ttcatttgaa gttctggcag	960

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tagaagacac acaaggaatg gacaatcttg cttgtgaaag tcaacaaccc acctctgaag	1020
aagtagtgga aaatcctacc atacagaagg aagtcataga gtgtgacgtg aaaactaccg	1080
aagttgtagg caatgtcata cttaaaccat cagatgaagg tgtaaagta acacaagagt	1140
taggtcatga ggatcttatg gctgcttatg tggaaaacac aagcattacc attaagaaac	1200
ctaagagct ttcactagcc ttaggtttaa aaacaattgc cactcatggt attgctgcaa	1260
ttaatagtgt tccttgaggt aaaattttgg cttatgtcaa accattctta ggacaagcag	1320
caattacaac atcaaattgc gctaagagat tagcacaacg tgtgtttaac aattatatgc	1380
cttatgtgtt tacattattg ttccaattgt gtacttttac taaaagtacc aattctagaa	1440
ttagagcttc actacctaca actattgcta aaaatagtgt taagagtgtt gctaaattat	1500
gtttggatgc cggcattaat tatgtgaagt cacccaaatt ttctaaattg ttcacaatcg	1560
ctatgtggct attgtttgta agtatttgct taggttctct aatctgtgta actgctgctt	1620
ttggtgtact cttatctaatt tttggtgctc cttcttattg taatggcgtt agagaattgt	1680
atcttaattc gtctaactgt actactatgg atttctgtga aggttctttt ccttgacgca	1740
tttgtttaag tggattagac tcccttgatt cttatccagc tcttgaaacc attcaggtga	1800
cgatttcacg gtacaagcta gacttgacaa ttttaggtct ggccgctgag tgggttttgg	1860
cataatgtt gttcacaaaa ttcttttatt tattaggtct ttcagctata atgcaggtgt	1920
tctttgcta ttttgctagt catttcatca gcaattcttg gctcatgtgg tttatcatta	1980
gtattgtaca aatgg	1995

<210> SEQ ID NO 47

<211> LENGTH: 1884

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 47

aattcttggc tcatgtggtt tatcattagt attgtacaaa tggcaccggt ttctgcaatg	60
gttaggatgt acatcttctt tgctcttttc tactacatat ggaagagcta tgttcatatc	120
atggatggtt gcacctcttc gacttgcatt atgtgctata agcgcaatcg tgccacacgc	180
gttgagtgt caactattgt taatggcatt aagagatctt tctatgtcta tgcaaatgga	240
ggccgtggct tctgcaagac tcacaattgg aattgtctca attgtgacac attttgcaat	300
ggtagtacat tcattagtga tgaagttgct cgtgatttgt cactccagtt taaaagacca	360
atcaacccta ctgaccagtc atcgtatatt gttgatagt ttgctgtgaa aaatggcgcg	420
cttcacctct actttgacaa ggctgggtcaa aagacctatg agagacatcc gctctcccat	480
tttgtcaatt tagacaattt gagagctaac aacactaaag gttcactgcc tattaatgtc	540
atagtttttg atggcaagtc caaatgcgac gagtctgctt ctaagtctgc ttctgtgtac	600
tacagtcagc tgatgtgcca acctattctg ttgcttgacc aagctcttgt atcagacgtt	660
ggagatagta ctgaagtctt cgtaaatggt tttgatgctt atgtogacac cttttcagca	720
acttttagtg ttcttatgga aaaacttaag gcacttgttg ctacagctca cagcgagtta	780
gcaaagggtg tagctttaga tgggtgcctt tctacattcg tgtcagctgc ccgacaaggt	840
gttgttgata ccgatgttga cacaaggat gttattgaat gtctcaaaact ttcacatcac	900
tctgacttag aagtgcacag tgacagttgt aacaatttca tgctcaccta taataaggtt	960

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gaaaacatga cgcccagaga tcttgccgca tgtattgact gtaatgcaag gcatatcaat	1020
gccaagtag caaaaagtca caatgtttca ctcatctgga atgtaaaaga ctacatgtct	1080
ttatctgaac agctgcgtaa acaaatctgt agtgctgcca agaagaacaa catacctttt	1140
agactaactt gtgctacaac tagacaggtt gtcaatgtca taactactaa aatctcactc	1200
aagggtggta agattgttag tacttgtttt aaacttatgc ttaagggcac attattgtgc	1260
gttcttgctg cattgggttg ttatatcggt atgccagtac atacattgtc aatccatgat	1320
ggttacacaa atgaaatcat tggttacaaa gccattcagg atgggtgtcac tcgtgacatc	1380
atttctactg atgattgttt tgcaataaaa catgctggtt ttgacgcatg gtttagccag	1440
cgtgggtggt catacaaaaa tgacaaaagc tgccctgtag tagctgctat cattacaaga	1500
gagattgggt tcatagtgcc tggcttaccg ggtactgtgc tgagagcaat caatgggtgac	1560
ttcttgcaat ttctacctcg tgtttttagt gctgttgga acatttgcta cacaccttcc	1620
aaactcattg agtatagtga ttttgctacc tctgcttgcg ttcttgctgc tgagtgtaca	1680
atttttaagg atgctatggg caaacctgtg ccatattgtt atgacactaa tttgctagag	1740
ggttctatatt cttatagtga gcttcgtcca gacactggt atgtgcttat ggatgggtcc	1800
atcatacagt ttcctaacac ttacctggag ggttctgtta gagtagtaac aacttttgat	1860
gctgagtact gtagacatgg taca	1884

<210> SEQ ID NO 48

<211> LENGTH: 2020

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 48

cactcgttat gtgcttatgg atggttccat catacagttt cctaacactt acctggaggg	60
ttctgttaga gtagtaacaa cttttgatgc tgagtactgt agacatggta catgcgaaag	120
gtcagaagta ggtatttgcc tatctaccag tggtagatgg gttcttaata atgagcatta	180
cagagctcta tcaggagttt tctgtgtgtg tgatgcgatg aatctcatag ctaacatctt	240
tactcctctt gtgcaacctg tgggtgcttt agatgtgtct gcttcagtag tggctgggtg	300
tattattgcc atattgggtga cttgtgctgc ctactacttt atgaaattca gacgtgtttt	360
tggtagtac aacctggtt ttgctgctaa tgcacttttg tttttgatgt ctttactat	420
actctgtctg gtaccagctt acagctttct gccgggagtc tactcagtct tttacttgta	480
cttgacattc tatttcacca atgatgtttc attcttggct caccttcaat ggtttgccat	540
gttttctcct attgtgcctt tttggataac agcaatctat gtattctgta tttctctgaa	600
gcactgccat tggttcttta acaactatct taggaaaaga gtcattgtta atggagtac	660
atttagtacc ttcgaggagg ctgctttgtg tacctttttg ctcaacaagg aaatgtacct	720
aaaattgcgt agcgagacac tggttgccact tacacagtat aacaggatc ttgctctata	780
taacaagtac aagtatttca gtggagcctt agatactacc agctatcgtg aagcagcttg	840
ctgccactta gcaaggctc taaatgactt tagcaactca ggtgctgatg ttctctacca	900
accaccacag acatcaatca cttctgctgt tctgcagagt ggttttagga aaatggcatt	960
cccgtcaggc aaagtgaag ggtgcctggt acaagtaacc tgtggaacta caactcttaa	1020
tggattgtgg ttggatgaca cagtatactg tccaagacat gtcatttgca cagcagaaga	1080

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catgcttaat cctaactatg aagatctgct cattcgcaaa toccaaccata gctttcttgt	1140
tcaggctggc aatgttcaac ttctgtttat tggccattct atgcaaaatt gtctgcttag	1200
gcttaaagtt gatacttcta accctaagac acccaagtat aaatttgtcc gtatccaacc	1260
tggtaaaca ttttcagttc tagcatgcta caatggttca ccactctggg tttatcagtg	1320
tgccatgaga cctaatacata ccattaaagg ttctttcctt aatggatcat gtggtagtgt	1380
tggttttaac attgattatg attgctgtgc ttctgtctat atgcatcata tggagcttcc	1440
aacaggagta cagctgggta ctgacttaga aggtaaatc tatgggtccat ttgttgacag	1500
acaaactgca caggctgcag gtacagacac aaccataaca ttaaatgttt tggcatggct	1560
gtatgctgct gttatcaatg gtgatagggt gtttcttaat agattcacca ctactttgaa	1620
tgactttaac cttgtggcaa tgaagtacaa ctatgaacct ttgacacaag atcatgttga	1680
catattggga cctctttctg ctcaaacagg aattgccgtc ttagatatgt gtgctgcttt	1740
gaaagagctg ctgcagaatg gtatgaatgg tcgtactatc cttggtagca ctattttaga	1800
agatgagttt acaccatttg atgtgtttag acaatgctct ggtgttacct tccaaggtaa	1860
gttcaagaaa attgttaagg gcactcatca ttggatgctt ttaactttct tgacatcact	1920
attgattctt gttcaaagta cacagtgggc actgttttct tttgtttacg agaatgcttt	1980
cttgccattt actcttggtt ttatggcaat tgctgcatgt	2020

<210> SEQ ID NO 49

<211> LENGTH: 2040

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 49

agcatttcca gcctgaagac gtactgtagc agctaaactg cccagcacca tacctctatt	60
taggttggtt aagcctttga tgaagtacaa gtatttcaact ttaggccctt ttggtgtgtc	120
tgtaacaaac ctacaagggt gttccagttc tgtgtaaatt gtacctgtac catcactctt	180
agggaaatcta gccattttga gatcttggtg gtctgatagt aatgccagca caaacctacc	240
tcccttcgaa ttgttatagt aggcaagtgc attgtcatca gtacaagctg tttgtgtggt	300
accagccgca caggacatct gtcgtagtgc tactggactc agttcattat tctgtagttt	360
aacagctgag ttggctctta gagctgtaac aataagaggo caagccaaat ttggtgaatt	420
gtccatgtta atttactaa gttgaacaat cttgctatcc gcatacaaa cttgctggat	480
ttcccagagt gcagatgcat atgtaaagggt gttaccatca caagtgttct ttaggtacc	540
ataatcaggg acaacaacca tgagtttggc tgctgtagtc aatggatga tgttagtggt	600
aacacaacca tcacgcgcat gtgtgataat gttgttaagt gcatacattat caagcttcct	660
aagcatagtg aagagcattg tttgcatagc actagttact tttgccctct tgcctcaga	720
tcttgccctgt ttgtacattt gggcatagc ctgatctgcc atcttttcca acttgctgtg	780
catggcagca tcacgggtcaa actcagattt agccacatto aaagatttct ttaacttttt	840
gagaacgact tcagaatcac cattagctac agcctgctca taggcctcct gggcagtggt	900
ataagcggca tatgatggtt aagaactaaa ttctgaagca atagcctgaa gagtagcacg	960
gttatcgagc atttctctgc acaacctatt aatgtctaca gcacctgca tggatagcaa	1020
aacagacaaa agagaaacca tcttctcgaa agcttcagtt gtgtcttttg caagaagaat	1080

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atcattgtgg agttgtacac attgtgccca caatttagaa gatgactcta ctctaagttg	1140
ttgaagaacc gagagcagta ccacagatgt gcactttacg tcagacattt tagactgtac	1200
agtagcaacc ttgatacatg gtttacctcc aataccaaac aacttaatgt taagcttgaa	1260
agcatcaata ctactcttag gaggcataag cccctgggag ttcataatac taaattcttg	1320
tgtagagacc aagtagtcat aaacaccaag agtaagcctg aagtaacggt tgagtaaaaca	1380
gaaaaggcca aagtagcagc agcaacaata gcctaagaaa caataaaca gcatgataca	1440
ctgtaaggtg ttgccagtaa taaataacaa tgggtaatac tcaacacaca caaacactat	1500
agctctagct aaaaacatga tagtcgtaac gacaccagaa tagttagagg ttacagaaat	1560
aactaaggcc cacatggaaa tagcttgatc taaagcatta ccatagtaga ctttgtaaac	1620
aagtgtaatg acattcatca gtgtccaaac acgtctagca gcacatcat aaacagtgcg	1680
agctgtcatg agaataagc aaactaaagc tgaagcatac ataacacaat ccttaagcct	1740
ataaccagac aagctagtgt cagccaattc aagccatgto atgatacgca tccccagct	1800
agcaggcatg tagaccatat taaagtaagc aactgttgca agagaaggta acagaaacaa	1860
gcacaagaat gcgtgcttat gcttaacaag cagcatagca catgcagcaa ttgccataat	1920
accaagagta aatggcaaga aagcattctc gtaacaaaag aaaaacagt accactgtgt	1980
actttgaaca agaatacaata gtgagtgtcaa gaaagttaaa agcatccaat gatgagtgtca	2040

<210> SEQ ID NO 50

<211> LENGTH: 2012

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 50

cttgtagggt tgttacagac acacccaaaag ggcctaaagt gaaatacttg tacttcatca	60
aaggcttaaa caacctaaat agaggtagtg tgctgggcag tttagctgct acagtacgto	120
ttcaggctgg aaatgtaca gaagtacctg ccaattcaac tgtgctttcc ttctgtgctt	180
ttgcagtaga cctgtctaaa gcatataagg attacctagc aagtgaggga caaccaatca	240
ccaactgtgt gaagatgttg tgtacacaca ctggtacagg acaggcaatt actgtaacac	300
cagaagctaa catggaccaa gagtcccttg gtggtgcttc atgttgtctg tattgtagat	360
gccacattga ccattccaaat cctaaaggat tctgtgactt gaaaggtaag tacgtccaaa	420
tacctaccac ttgtgctaag gaccagtggt gttttact tagaaacaca gtctgtaccg	480
tctgcggaat gtggaaaggt tatggctgta gttgtgacca actccgcgaa cccttgatgc	540
agtctgcgga tgcatacaacg tttttaaacg ggtttgcggt gtaagtgcag cccgtcttac	600
accgtgcggc acaggcacta gtactgatgt cgtctacagg gcttttgata ttacaacga	660
aaaagttgct ggttttgcaa agttccctaaa aactaattgc tgtcgcttcc aggagaagga	720
tgagggaaggc aatttattag actcttactt ttagttaaag aggcatacta tgtctaacta	780
ccaacatgaa gagactattt ataacttggg taaagattgt ccagcgggtg ctgtccatga	840
ctttttcaag tttagagtag atggtgacat ggtaccacat atatcacgto agcgtctaac	900
taaatacaca atggctgatt tagtctatgc tctacgtcat tttgatgagg gtaattgtga	960
tacattaaaa gaaatactcg tcacatacaa ttgctgtgat gatgattatt tcaataagaa	1020
ggattggtat gacttcgtag agaatcctga catcttacgc gtatatgcta acttaggtga	1080

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gcgtgtacgc caatcattat taaagactgt acaattctgc gatgctatgc gtgatgcagg	1140
cattgttaggc gtactgacat tagataatca ggatcttaat gggaactggg acgatttcgg	1200
tgatttcgta caagtagcac caggctgcgg agttcctatt gtggattcat attactcatt	1260
gctgatgccc atcctcactt tgactagggc attggctgct gagtcccata tggatgctga	1320
tctcgcaaaa ccacttatta agtgggattt gctgaaatat gattttacgg aagagagact	1380
ttgtctcttc gaccgttatt ttaaatattg ggaccagaca taccatccca attgtattaa	1440
ctgtttggat gatagtgta tccttcattg tgcaaacctt aatgtgttat tttctactgt	1500
gtttccacct acaagttttg gaccactagt aagaaaaata tttgtagatg gtgttccttt	1560
tggtgtttca actggatacc attttcgtga gttaggagtc gtacataatc aggatgtaaa	1620
cttacatagc tcgcgtctca gtttcaagga acttttagtg tatgctgctg atccagctat	1680
gcctgcagct tctggcaatt tattgctaga taaacgcact acatgctttt cagtagctgc	1740
actaacaac aatgttgctt ttcaaacgtt caaacccggg aattttaata aagactttta	1800
tgactttgct gtgtctaaag gtttctttaa ggaaggaagt tctgttgaac taaaacactt	1860
cttctttgct caggatggca acgctgctat cagtatttat gactattatc gttataatct	1920
gccacaatg tgtgatatca gacaactcct attcgtagtt gaagttgttg ataaatactt	1980
tgattgttac gatggtggct gtattaatgc ca	2012

<210> SEQ ID NO 51

<211> LENGTH: 1877

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 51

gtacttcgcg tacagtggca ataccatatg acagcttaaa tgtttcctca gtggctttga	60
gcgtttctgc tgcgaaaagc ttgagtctct cagtacaagt gttggcaagt atgtaatcgc	120
cagcattagt ccaatcacat gttgctatcg cattgaagtc agtgacattg tcactgccta	180
cacatgtgtt tttgtataaa ccaaaaaacct gaccattagc acataatgga aaactaatgg	240
gaggcttatg tgacttgcaa taatagctca tacctcctag atacagttgt gtcacatcag	300
tgacatcaca acctggggca ttgcaaact agggattaac agacaacact aattttgtgtg	360
atgttgaaat gacatggtca tagcagcact tgcaacatag gaatggtctc ctaatacagg	420
caccgcaacg aagtgaagtc tgtgaattgc acaatacaca agcacctaca gcctgcaaga	480
ctgtatgtgg tgtgtacata gcctcataaa actcagggtc ccagtaccgt gaggtgttat	540
cattagttag cattacggaa tacatgtcca acatgtggcc agtaagctca tcatgtaact	600
ttctaagtga ttgtaaatac aagtgaaga catcagcata ctctgatta ggatgttttg	660
taagtgggta agcatcaata gccagtgaac cgaacctttc aatcataagt gtaccatctg	720
ttttgacaat atcatcgaca aaacagcctg cgcctaatat tottgatgga tctgggtaag	780
gcaggtaac gtaatcatct cttgttttaa ctagcattgt atgctgtgag caaaattcgt	840
gaggctcttt agtaaggcta gtctcagtc aacattttgc ctgagacatg aacacattat	900
tttgataata aagaactgcc ttaaagtctt taatgctagc tactaaacct tgagccgcat	960
agttactgtt atagcacaca acggcatcat cagaaagaat catcatggag aaatgtttac	1020
gcaggtaagc gtaaaactca tccacgaatt catgatcaac atccctatct ctatagagac	1080

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actcatagag cctgtgttgt agattgcgga catacttgto agctatctta ttaccatcag	1140
ttgaaagaag tgcatttaca ttggctgtaa cagcttgaca aatgttaaag acactattag	1200
cataagcagt ttagcatca ccggatgatg ttccacctg tttacatat agtgagccgc	1260
cacacatgac catctcactt aatacttgcg cactctggt agctaacctg tagaaacggt	1320
gtgataagtt acagcaagtg ttatgtttgc gagcaagaac aagagaggcc attatcctaa	1380
gcatgttagg catggctctg tcacattttg gataatocca acccataagg tgtggagttt	1440
ctacatcact gtaaacagtt tttacatat tatgccagcc accgtaaac ttgcttggtc	1500
caattaccac agtagctcct ctatggcgg ctattgactt caataatttc tgatgaaact	1560
gtctatttgt catagtacta cagatagaga caccagctac ggtgcgagct ctattctttg	1620
cactaatggc atacttaaga ttcatgttag ttatagtagg gatgacatta cgcttagtat	1680
acgcgaaaag tgcactttga tcctcataac tcattgagtc ataataaagt ctagccttac	1740
cccatttatt aaatgggaaa ccagctgatt tatccagatt gttaacgatt acttggttg	1800
cattaataca gccaccatcg taacaatcaa agtatttato aacaacttca actacgaata	1860
ggagttgtct gatatca	1877

<210> SEQ ID NO 52

<211> LENGTH: 2051

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 52

tcaggtccaa tcttgacaaa gtacttcatt gatgtaagct caaagccatg cgcccaaagg	60
acgaacacga ctctgtctga caatcctttc agtgtatcac tgagcatttg tactatctta	120
atacgcacta cattccaggg caagccttta tacatgagtg gtataagatg tttaaactgg	180
tcacctggtg gaggttttgc attaaactct gtgaattctg tgttattttc agtgtcaaca	240
taaccagtcg gtacagctac taagttaaca cctgtagaaa atcctagctg gagaggtagg	300
ttagtacctc cagcatctct agttgcatga cagccctcta catcaaagcc aatccacgca	360
cgaacgtgac gaatagcttc ttgcggggtg ataaacatat tagggtaacc attgacttgg	420
taattcattt tgaaccat catagagatg agtctacggt aggtcatgtc ctttggtatg	480
cctggtagtg caacacataa tccttcagtc ttgaacttta tatcaacgct gaggtgtgta	540
ggtgcctgtg taggatgaag accagtaatg atcttactac agtccttaaa aagtcagtt	600
acattttctg cttgtaatgt agccacattg cgacgtggtt tttctagact tgtaaattgc	660
agtttgtcat aaagatctct atcagacatt atgcacaaaa tgccaatttt tgcccttggtg	720
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atgacatagt catattcaga accctgtgat gaatcaacag tctgcgtagg caatcctaag	840
atttttgaag ctacagcgtt ctgtgaatta taagtgaga taaaacagc tttctccaa	900
gcaggattgc gtgtaagaaa ttctottaca acgcctattt gaggtctggt gattgcagat	960
gaaacatcat gtgtaataac acctttgtag aacattttga agcattgagc tgacttatcc	1020
ttgtgtgctt ttgcttatt gtcataaact aaagcactca cagtgtcaac aatttcagca	1080
ggacaacggc gacaagttcc aaggaacatg tctggaccta ttgttttcat aagtctgcac	1140
actgaattaa aatattctgg ttctagtgtg cctttagtca gcaatgtgcg gggggctggt	1200

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aattgagcag gatcgccaat atagacgtag tgttttgcac gaagtctagc attgacaaca	1260
ctcaagtcac aattagtagc catagagatt tcatcaaaga ctacaatgac agcagttgtt	1320
tctggcaatg catttacagt gcagaaaaca tactgttcta gtgttgaaatt cactttgaat	1380
ttatcaaaac actctacgcg cgcacgcgca ggtatgatc tactacattt atctatgggc	1440
aaatatttta atgccttttc acatagggca tcaacagctg catgagagca tgccgtatac	1500
actatgcgag cagatgggta atagagagca agtccgatgg caaaatgact cttaccagta	1560
ccagggtggtc cttggagtgt agagtacttt tgcatgccga ccttttgata atttgaaca	1620
ttgctagaaa actcatctga gatgttgagt gttgggtaca agccagtaat tctcacatag	1680
tgctcttggt gcaactagat aggtgcacta agtggcatta cagtgtgaga tgtcaacaca	1740
aagtaatcac caacattcaa cttgtatgtc gtagtacctc tgtacacaac agcatcacca	1800
tagtcacctt tttcaaaggt gtactctcca atctgtactt tactattttt agttacacgg	1860
taaccagtaa agacatagtt tctgttcaat ggtggtctag gttttccaac ctcccatgaa	1920
agatgcaatt ctctgtcaga gagtacttcg cgtacagtgg caataccata tgacagctta	1980
aatgtttcct cagtggcttt gagcgtttct gctgcgaaaa gcttgagtct ctcagtacaa	2040
gtgttggtgca g	2051

<210> SEQ ID NO 53

<211> LENGTH: 2075

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 53

tgctttagt tttgggtaga aggtttcaac atgtccatcc ttacaccaa gcatgaatga	60
aatttcagca tagtcaattg taaccttgac cacttttgaa atcactgaca aatcttgtga	120
ctttattatc tcgacaaagt catcaagtaa aagatcaatc acagaacaca cacattttga	180
tgaacctgtt tgcgcactctg ttatgaagta atttttcaact gtgctgtcca tagggataaa	240
atcctctaata ttaagtgtg aatcttgtga gcgcttggt aagcctatca ttaaatgaag	300
accgccaagt tgtccatgac tgaatctcc ataaacgatg tgttcgaag catagccctc	360
gagcttatat cgctgtatga attcatccat agcagactcg agaaagtcag tttccatttg	420
tgatctgggc ttaaaatcct ctaagtctct gctctgagta aagtaggttt caggcaactg	480
ttgaataatg ccgtctactt tcttaaagta gttaaactgt gtttttactg attctccaat	540
taatgtgact ccattgacgc tagcttgtgc tgggtccctt gaagggttta gaccttgac	600
tgaaccttct gttattaaaa caccattacg ggcgtttcta aaaaggctta cctgtccttc	660
cactctacca tcaacaaga cagtaagtga agaacaagca ctctcagtag gtttcttggc	720
aatgtcagtc attgtgcaga cacctattgt agatacatgt gctggggctt ctcttttgta	780
gtcccagatt acagtattag cagcgatatc aacacccaaa ttattgagta tcttaatctc	840
tggcactggt ttaatgttac gcttagccca aagctcaaat gcaacattaa cagggaagtgt	900
tgtcttattt tcaaatgact ccacatcaat accatctacc tttgtgtaa cagcattatt	960
aatgatggaa acagggtgct cgccggcgtg tccatcaaag tgcctttat taacaacatt	1020
ataagccaca ttttctaaac tctgtaacct ggtaaatgta ttccacaggt tataagtatc	1080
aaattgtttg taaatccata ggctaaatcc agcagaaatc atcatattat atgcatccaa	1140

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gtactgtcgg tactcatttg catggtgtct gcaaacagca ccacctaaat tgcacgtgtg	1200
aatacacgta gcagatttga gtggaacata atcaatatcc gacactactt gtttgccatg	1260
agactcacia ggactatcag aatagtaaaa gaaaggcaat tgctttaaat tagtaaatgc	1320
acttttatcg aaagctggag tgtggaatgc atgcttattc acatacaaac taccaccatc	1380
acagcctggg aagttcaagt ttgacaagac tcttgtgtca aacctacaca caattgcatt	1440
ggctgggtaa cgatcaacgt tacaattcca aaacaacaa acaccatcag tgaatttato	1500
gtgatgtgta gcataagaat agaagagttc ctctattttg taagctttgt cactacatgg	1560
ctgagcatcg tagaatttcc attctacttc agcctgaggc acacacttga tagccttttg	1620
atttccaatg tcatgaagaa ctggaaactt atcagcaagc aatgcagact tcacaacct	1680
gtgtgttact tttctgcaag cagaattaac cctcagttca tctcctataa tagggtattc	1740
aacagaccaa tcaacgcgct taacaaagca ctcatggact gctaaacatc tagtcatgat	1800
agcatcacia ctgaccacat gtgcatttcc atgtacctgg caatgttggg catggttact	1860
ctgaagggtt cccgtaaaag cccactgctg aacatcaatc ataaatgggt tatagacata	1920
gtcaaaaccc acagaatgat tccagcaggc ataagtatct gatgaagtag aaaagcaagt	1980
tgcacgtttg tcacacagac aacacgttct ttcaggtcca atcttgacaa agtacttcat	2040
tgatgtaagc tcaaagccat gcgcccacaa gacga	2075

<210> SEQ ID NO 54

<211> LENGTH: 1891

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 54

aagattcacc acttaaatga gaggatttta tccctatgga cagcacagtg aaaaattact	60
tcataacaga tgcgcaaaca ggttcacaa aatgtgtgtg ttctgtgatt gatcttttac	120
ttgatgactt tgtcgagata ataaagtcac aagatttgcg agtgatttca aaagtgggtca	180
agggttacaat tgactatgct gaaatttcat tcatgctttg gtgtaaggat ggacatgttg	240
aaaccttcta cccaaaacta caagcaagtc aagcgtggca accaggtgtt gcgatgccta	300
acttgtacaa gatgcaaaga atgcttcttg aaaagtgtga ccttcagaat tatgggtgaa	360
atgctgttat accaaaagga ataatgatga atgtcgcaaa gtatactcaa ctgtgtcaat	420
acttaaatac acttacttta gctgtacctt acaacatgag agttattcac tttgggtgctg	480
gctctgataa aggagttgca ccaggtacag ctgtgctcag acaatgggtg ccaactggca	540
cactacttgt cgattcagat cttaatgact tcgtctccga cgcagattct actttaattg	600
gagactgtgc aacagtacat acggctaata aatgggacct tattattagc gatatgtatg	660
accctaggac caaacatgtg acaaaagaga atgactctaa agaaggggtt ttcacttatt	720
tgtgtggatt tataaagcaa aaactagccc tgggtgggtc tatagctgta aagataacag	780
agcattcttg gaatgtgac ctttacaagc ttatgggcca tttctcatgg tggacagctt	840
ttgttacaaa tgtaaatgca tcatcatcgg aagcattttt aattggggct aactatcttg	900
gcaagccgaa ggaacaaatt gatggctata ccatgcatgc taactacatt ttctggagga	960
acacaaatcc tatccagttg tcttctctatt cactctttga catgagcaaa tttctcttta	1020
aattaagagg aactgtctga atgtctctta aggagaatca aatcaatgat atgatttatt	1080

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ctcttctgga aaaaggtagg cttatcatta gagaaaaca cagagttgtg gtttcaagtg 1140
atattcttgt taacaactaa acgaacatgt ttattttctt attatttctt actctcacta 1200
gtggtagtga ccttgaccgg tgcaccactt ttgatgatgt tcaagctcct aattacactc 1260
aacatacttc atctatgagg ggggtttact atcctgatga aattttttaga tcagacactc 1320
tttatttaac tcaggattta tttcttccat tttattctaa tggtacaggg tttcatacta 1380
ttaatcatat gtttggaac cctgtcatat cttttaagga tgggtattat tttgctgcca 1440
cagagaaatc aaatgttgtc cgtgggtggg tttttgggtc taccatgaac aacaagtcac 1500
agtcggtgat tattattaac aattctacta atgttgttat acgagcatgt aactttgaat 1560
tgtgtgacaa ccttttcttt gctgtttcta aacctatggg tacacagaca catactatga 1620
tattcgataa tgcatttaac tgcactttcg agtacatata tgatgccttt tcgcttgatg 1680
tttcagaaaa gtcaggtaac tttaaacact tacgagagtt tgtgtttaaa aataaagatg 1740
ggtttctcta tgtttataag ggctatcaac ctatagatgt agttcgtgat ctaccttctg 1800
gttttaacac tttgaaacct atttttaagt tgcctcttgg tattaacatt acaaatttta 1860
gagccattct tacagccttt tcacctgtc a 1891

```

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<210> SEQ ID NO 55
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: N sens primer

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<400> SEQUENCE: 55

```

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cccatatgtc tgataatgga cccaatcaa ac 32

```

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<210> SEQ ID NO 56
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: N antisens primer

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<400> SEQUENCE: 56

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cccccggtg cctgagttga atcagcagaa gc 32

```

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<210> SEQ ID NO 57
<211> LENGTH: 31
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: Sc sens primer

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<400> SEQUENCE: 57

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cccatatgag tgaccttgac cggtgcacca c 31

```

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<210> SEQ ID NO 58
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SL sens primer

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<400> SEQUENCE: 58

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```

cccatatgaa accttgacc ccacctgtc 30

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<210> SEQ ID NO 59
<211> LENGTH: 33
<212> TYPE: DNA
<213> ORGANISM: Sc and SL antisens primer

<400> SEQUENCE: 59

cccccggtt taatatattg ctcatatttt ccc 33

<210> SEQ ID NO 60
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Sens set 1 primer

<400> SEQUENCE: 60

ggcatcgtat gggttg 16

<210> SEQ ID NO 61
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Antisens set 2 (28774-28759) primer

<400> SEQUENCE: 61

cagtttcacc acctcc 16

<210> SEQ ID NO 62
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Sens set 2 (28375-28390) primer

<400> SEQUENCE: 62

ggctactacc gaagag 16

<210> SEQ ID NO 63
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Antisens set 2 (28702-28687)primer

<400> SEQUENCE: 63

aattaccgag actacg 16

<210> SEQ ID NO 64
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Probe 1/set 1 (28561-28586)

<400> SEQUENCE: 64

ggcaccgcga atcctaataa caatgc 26

<210> SEQ ID NO 65
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Probe 2/set 1 (28588-28608)

<400> SEQUENCE: 65

gccaccgtgc tacaacttcc t 21

<210> SEQ ID NO 66
<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Probe 1/set 2 /probe N/FL (28541-28563)

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<400> SEQUENCE: 66

atacacccaa agaccacatt ggc 23

<210> SEQ ID NO 67

<211> LENGTH: 25

<212> TYPE: DNA

<213> ORGANISM: Probe 2/set 2/probe SARS/N/LC705 (28565-28589)

<400> SEQUENCE: 67

cccgcaatcc taataacaat gctgc 25

<210> SEQ ID NO 68

<211> LENGTH: 30

<212> TYPE: DNA

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: Anchor primer 14T

<400> SEQUENCE: 68

agatgaattc ggtacctttt tttttttttt 30

<210> SEQ ID NO 69

<211> LENGTH: 13

<212> TYPE: PRT

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: M2-14 peptide

<400> SEQUENCE: 69

Ala Asp Asn Gly Thr Ile Thr Val Glu Glu Leu Lys Gln
1 5 10

<210> SEQ ID NO 70

<211> LENGTH: 12

<212> TYPE: PRT

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: E1-12 peptide

<400> SEQUENCE: 70

Met Tyr Ser Phe Val Ser Glu Glu Thr Gly Thr Leu
1 5 10

<210> SEQ ID NO 71

<211> LENGTH: 24

<212> TYPE: PRT

<213> ORGANISM: artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: E53-72 peptide

<400> SEQUENCE: 71

Lys Pro Thr Val Tyr Val Tyr Ser Arg Val Lys Asn Leu Asn Ser Ser
1 5 10 15Glu Gly Val Pro Asp Leu Leu Val
20

<210> SEQ ID NO 72

<211> LENGTH: 153

<212> TYPE: DNA

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 72

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gatattaggt ttttacctac ccaggaaaag ccaaccaacc togatctctt gtagatctgt      60
tctctaaacg aactttaaaa tctgtgtagc tgtcgctcgg ctgcatgcct agtgcaccta      120
cgcagtataa acaataataa attttactgt cgt                                     153

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<210> SEQ ID NO 73
<211> LENGTH: 410
<212> TYPE: DNA
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 73

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ttctccagac aacttcaaaa ttccatgagt ggagcttctg ctgattcaac tcaggcataa      60
aactcatga tgaccacaca aggcagatgg gctatgtaaa cgttttcgca attccgttta      120
cgatacatag tctactcttg tgcagaatga attctcgtaa ctaaacagca caagtaggtt      180
tagttaactt taatctcaca tagcaatctt taatcaatgt gtaacattag ggaggacttg      240
aaagagccac cacattttca tcgaggccac gcggagtacg atcgagggta cagtgaataa      300
tgctaggggag agctgcctat atggaagagc cctaattgtgt aaaattaatt ttagtagtgc      360
tatcccatg tgattttaat agcttcttag gagaatgaca aaaaaaaaaa                  410

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<210> SEQ ID NO 74
<211> LENGTH: 4382
<212> TYPE: PRT
<213> ORGANISM: CORONAVIRUS

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<400> SEQUENCE: 74

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Met Glu Ser Leu Val Leu Gly Val Asn Glu Lys Thr His Val Gln Leu
 1             5             10             15

Ser Leu Pro Val Leu Gln Val Arg Asp Val Leu Val Arg Gly Phe Gly
      20             25             30

Asp Ser Val Glu Glu Ala Leu Ser Glu Ala Arg Glu His Leu Lys Asn
      35             40             45

Gly Thr Cys Gly Leu Val Glu Leu Glu Lys Gly Val Leu Pro Gln Leu
      50             55             60

Glu Gln Pro Tyr Val Phe Ile Lys Arg Ser Asp Ala Leu Ser Thr Asn
      65             70             75             80

His Gly His Lys Val Val Glu Leu Val Ala Glu Met Asp Gly Ile Gln
      85             90             95

Tyr Gly Arg Ser Gly Ile Thr Leu Gly Val Leu Val Pro His Val Gly
      100            105            110

Glu Thr Pro Ile Ala Tyr Arg Asn Val Leu Leu Arg Lys Asn Gly Asn
      115            120            125

Lys Gly Ala Gly Gly His Ser Tyr Gly Ile Asp Leu Lys Ser Tyr Asp
      130            135            140

Leu Gly Asp Glu Leu Gly Thr Asp Pro Ile Glu Asp Tyr Glu Gln Asn
      145            150            155            160

Trp Asn Thr Lys His Gly Ser Gly Ala Leu Arg Glu Leu Thr Arg Glu
      165            170            175

Leu Asn Gly Gly Ala Val Thr Arg Tyr Val Asp Asn Asn Phe Cys Gly
      180            185            190

Pro Asp Gly Tyr Pro Leu Asp Cys Ile Lys Asp Phe Leu Ala Arg Ala
      195            200            205

Gly Lys Ser Met Cys Thr Leu Ser Glu Gln Leu Asp Tyr Ile Glu Ser

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210					215					220					
Lys 225	Arg	Gly	Val	Tyr	Cys 230	Cys	Arg	Asp	His	Glu 235	His	Glu	Ile	Ala	Trp 240
Phe	Thr	Glu	Arg	Ser 245	Asp	Lys	Ser	Tyr	Glu 250	His	Gln	Thr	Pro	Phe 255	Glu
Ile	Lys	Ser	Ala 260	Lys	Lys	Phe	Asp 265	Thr	Phe	Lys	Gly	Glu	Cys 270	Pro	Lys
Phe	Val	Phe 275	Pro	Leu	Asn	Ser	Lys 280	Val	Lys	Val	Ile	Gln 285	Pro	Arg	Val
Glu 290	Lys	Lys	Lys	Thr	Glu	Gly 295	Phe	Met	Gly	Arg	Ile 300	Arg	Ser	Val	Tyr
Pro 305	Val	Ala	Ser	Pro	Gln 310	Glu	Cys	Asn	Asn	Met 315	His	Leu	Ser	Thr	Leu 320
Met	Lys	Cys	Asn 325	His	Cys	Asp	Glu	Val	Ser 330	Trp	Gln	Thr	Cys 335	Asp	Phe
Leu	Lys	Ala 340	Thr	Cys	Glu	His	Cys	Gly 345	Thr	Glu	Asn	Leu 350	Val	Ile	Glu
Gly	Pro	Thr 355	Thr	Cys	Gly	Tyr	Leu 360	Pro	Thr	Asn	Ala 365	Val	Val	Lys	Met
Pro 370	Cys	Pro	Ala	Cys	Gln	Asp 375	Pro	Glu	Ile	Gly	Pro 380	Glu	His	Ser	Val
Ala 385	Asp	Tyr	His	Asn 390	His	Ser	Asn	Ile	Glu	Thr 395	Arg	Leu	Arg	Lys	Gly 400
Gly	Arg	Thr	Arg 405	Cys	Phe	Gly	Gly	Cys	Val 410	Phe	Ala	Tyr	Val	Gly 415	Cys
Tyr	Asn	Lys	Arg 420	Ala	Tyr	Trp	Val	Pro	Arg 425	Ala	Ser	Ala	Asp 430	Ile	Gly
Ser	Gly 435	His	Thr	Gly	Ile	Thr	Gly 440	Asp	Asn	Val	Glu	Thr 445	Leu	Asn	Glu
Asp 450	Leu	Leu	Glu	Ile	Leu	Ser 455	Arg	Glu	Arg	Val	Asn 460	Ile	Asn	Ile	Val
Gly 465	Asp	Phe	His	Leu 470	Asn	Glu	Glu	Val	Ala 475	Ile	Ile	Leu	Ala	Ser	Phe 480
Ser	Ala	Ser	Thr 485	Ser	Ala	Phe	Ile	Asp	Thr 490	Ile	Lys	Ser	Leu	Asp 495	Tyr
Lys	Ser	Phe	Lys 500	Thr	Ile	Val	Glu	Ser 505	Cys	Gly	Asn	Tyr	Lys 510	Val	Thr
Lys	Gly 515	Lys	Pro	Val	Lys	Gly	Ala 520	Trp	Asn	Ile	Gly	Gln 525	Gln	Arg	Ser
Val 530	Leu	Thr	Pro	Leu	Cys	Gly 535	Phe	Pro	Ser	Gln	Ala 540	Ala	Gly	Val	Ile
Arg 545	Ser	Ile	Phe	Ala 550	Arg	Thr	Leu	Asp	Ala	Ala 555	Asn	His	Ser	Ile	Pro 560
Asp	Leu	Gln	Arg 565	Ala	Ala	Val	Thr	Ile	Leu 570	Asp	Gly	Ile	Ser	Glu 575	Gln
Ser	Leu	Arg 580	Leu	Val	Asp	Ala	Met	Val 585	Tyr	Thr	Ser	Asp 590	Leu	Leu	Thr
Asn 595	Ser	Val	Ile	Ile	Met	Ala	Tyr 600	Val	Thr	Gly	Gly 605	Leu	Val	Gln	Gln
Thr 610	Ser	Gln	Trp	Leu	Ser 615	Asn	Leu	Leu	Gly	Thr 620	Thr	Val	Glu	Lys	Leu

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Arg	Pro	Ile	Phe	Glu	Trp	Ile	Glu	Ala	Lys	Leu	Ser	Ala	Gly	Val	Glu	
625					630					635					640	
Phe	Leu	Lys	Asp	Ala	Trp	Glu	Ile	Leu	Lys	Phe	Leu	Ile	Thr	Gly	Val	
			645						650					655		
Phe	Asp	Ile	Val	Lys	Gly	Gln	Ile	Gln	Val	Ala	Ser	Asp	Asn	Ile	Lys	
			660					665					670			
Asp	Cys	Val	Lys	Cys	Phe	Ile	Asp	Val	Val	Asn	Lys	Ala	Leu	Glu	Met	
			675				680					685				
Cys	Ile	Asp	Gln	Val	Thr	Ile	Ala	Gly	Ala	Lys	Leu	Arg	Ser	Leu	Asn	
	690					695					700					
Leu	Gly	Glu	Val	Phe	Ile	Ala	Gln	Ser	Lys	Gly	Leu	Tyr	Arg	Gln	Cys	
	705				710					715					720	
Ile	Arg	Gly	Lys	Glu	Gln	Leu	Gln	Leu	Leu	Met	Pro	Leu	Lys	Ala	Pro	
				725					730					735		
Lys	Glu	Val	Thr	Phe	Leu	Glu	Gly	Asp	Ser	His	Asp	Thr	Val	Leu	Thr	
			740					745					750			
Ser	Glu	Glu	Val	Val	Leu	Lys	Asn	Gly	Glu	Leu	Glu	Ala	Leu	Glu	Thr	
		755					760					765				
Pro	Val	Asp	Ser	Phe	Thr	Asn	Gly	Ala	Ile	Val	Gly	Thr	Pro	Val	Cys	
	770					775					780					
Val	Asn	Gly	Leu	Met	Leu	Leu	Glu	Ile	Lys	Asp	Lys	Glu	Gln	Tyr	Cys	
	785				790					795					800	
Ala	Leu	Ser	Pro	Gly	Leu	Leu	Ala	Thr	Asn	Asn	Val	Phe	Arg	Leu	Lys	
				805					810					815		
Gly	Gly	Ala	Pro	Ile	Lys	Gly	Val	Thr	Phe	Gly	Glu	Asp	Thr	Val	Trp	
			820					825					830			
Glu	Val	Gln	Gly	Tyr	Lys	Asn	Val	Arg	Ile	Thr	Phe	Glu	Leu	Asp	Glu	
		835					840					845				
Arg	Val	Asp	Lys	Val	Leu	Asn	Glu	Lys	Cys	Ser	Val	Tyr	Thr	Val	Glu	
						855					860					
Ser	Gly	Thr	Glu	Val	Thr	Glu	Phe	Ala	Cys	Val	Val	Ala	Glu	Ala	Val	
	865				870					875					880	
Val	Lys	Thr	Leu	Gln	Pro	Val	Ser	Asp	Leu	Leu	Thr	Asn	Met	Gly	Ile	
				885					890					895		
Asp	Leu	Asp	Glu	Trp	Ser	Val	Ala	Thr	Phe	Tyr	Leu	Phe	Asp	Asp	Ala	
			900					905					910			
Gly	Glu	Glu	Asn	Phe	Ser	Ser	Arg	Met	Tyr	Cys	Ser	Phe	Tyr	Pro	Pro	
		915					920					925				
Asp	Glu	Glu	Glu	Glu	Asp	Asp	Ala	Glu	Cys	Glu	Glu	Glu	Glu	Ile	Asp	
	930				935						940					
Glu	Thr	Cys	Glu	His	Glu	Tyr	Gly	Thr	Glu	Asp	Asp	Tyr	Gln	Gly	Leu	
	945				950					955					960	
Pro	Leu	Glu	Phe	Gly	Ala	Ser	Ala	Glu	Thr	Val	Arg	Val	Glu	Glu	Glu	
				965					970					975		
Glu	Glu	Glu	Asp	Trp	Leu	Asp	Asp	Thr	Thr	Glu	Gln	Ser	Glu	Ile	Glu	
			980					985					990			
Pro	Glu	Pro	Glu	Pro	Thr	Pro	Glu	Glu	Pro	Val	Asn	Gln	Phe	Thr	Gly	
		995					1000					1005				
Tyr	Leu	Lys	Leu	Thr	Asp	Asn	Val	Ala	Ile	Lys	Cys	Val	Asp	Ile		
	1010					1015						1020				

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Val	Lys	Glu	Ala	Gln	Ser	Ala	Asn	Pro	Met	Val	Ile	Val	Asn	Ala
1025						1030					1035			
Ala	Asn	Ile	His	Leu	Lys	His	Gly	Gly	Gly	Val	Ala	Gly	Ala	Leu
1040						1045					1050			
Asn	Lys	Ala	Thr	Asn	Gly	Ala	Met	Gln	Lys	Glu	Ser	Asp	Asp	Tyr
1055						1060					1065			
Ile	Lys	Leu	Asn	Gly	Pro	Leu	Thr	Val	Gly	Gly	Ser	Cys	Leu	Leu
1070						1075					1080			
Ser	Gly	His	Asn	Leu	Ala	Lys	Lys	Cys	Leu	His	Val	Val	Gly	Pro
1085						1090					1095			
Asn	Leu	Asn	Ala	Gly	Glu	Asp	Ile	Gln	Leu	Leu	Lys	Ala	Ala	Tyr
1100						1105					1110			
Glu	Asn	Phe	Asn	Ser	Gln	Asp	Ile	Leu	Leu	Ala	Pro	Leu	Leu	Ser
1115						1120					1125			
Ala	Gly	Ile	Phe	Gly	Ala	Lys	Pro	Leu	Gln	Ser	Leu	Gln	Val	Cys
1130						1135					1140			
Val	Gln	Thr	Val	Arg	Thr	Gln	Val	Tyr	Ile	Ala	Val	Asn	Asp	Lys
1145						1150					1155			
Ala	Leu	Tyr	Glu	Gln	Val	Val	Met	Asp	Tyr	Leu	Asp	Asn	Leu	Lys
1160						1165					1170			
Pro	Arg	Val	Glu	Ala	Pro	Lys	Gln	Glu	Glu	Pro	Pro	Asn	Thr	Glu
1175						1180					1185			
Asp	Ser	Lys	Thr	Glu	Glu	Lys	Ser	Val	Val	Gln	Lys	Pro	Val	Asp
1190						1195					1200			
Val	Lys	Pro	Lys	Ile	Lys	Ala	Cys	Ile	Asp	Glu	Val	Thr	Thr	Thr
1205						1210					1215			
Leu	Glu	Glu	Thr	Lys	Phe	Leu	Thr	Asn	Lys	Leu	Leu	Leu	Phe	Ala
1220						1225					1230			
Asp	Ile	Asn	Gly	Lys	Leu	Tyr	His	Asp	Ser	Gln	Asn	Met	Leu	Arg
1235						1240					1245			
Gly	Glu	Asp	Met	Ser	Phe	Leu	Glu	Lys	Asp	Ala	Pro	Tyr	Met	Val
1250						1255					1260			
Gly	Asp	Val	Ile	Thr	Ser	Gly	Asp	Ile	Thr	Cys	Val	Val	Ile	Pro
1265						1270					1275			
Ser	Lys	Lys	Ala	Gly	Gly	Thr	Thr	Glu	Met	Leu	Ser	Arg	Ala	Leu
1280						1285					1290			
Lys	Lys	Val	Pro	Val	Asp	Glu	Tyr	Ile	Thr	Thr	Tyr	Pro	Gly	Gln
1295						1300					1305			
Gly	Cys	Ala	Gly	Tyr	Thr	Leu	Glu	Glu	Ala	Lys	Thr	Ala	Leu	Lys
1310						1315					1320			
Lys	Cys	Lys	Ser	Ala	Phe	Tyr	Val	Leu	Pro	Ser	Glu	Ala	Pro	Asn
1325						1330					1335			
Ala	Lys	Glu	Glu	Ile	Leu	Gly	Thr	Val	Ser	Trp	Asn	Leu	Arg	Glu
1340						1345					1350			
Met	Leu	Ala	His	Ala	Glu	Glu	Thr	Arg	Lys	Leu	Met	Pro	Ile	Cys
1355						1360					1365			
Met	Asp	Val	Arg	Ala	Ile	Met	Ala	Thr	Ile	Gln	Arg	Lys	Tyr	Lys
1370						1375					1380			
Gly	Ile	Lys	Ile	Gln	Glu	Gly	Ile	Val	Asp	Tyr	Gly	Val	Arg	Phe
1385						1390					1395			
Phe	Phe	Tyr	Thr	Ser	Lys	Glu	Pro	Val	Ala	Ser	Ile	Ile	Thr	Lys

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1400	1405	1410
Leu Asn Ser Leu Asn Glu Pro	Leu Val Thr Met Pro	Ile Gly Tyr
1415	1420	1425
Val Thr His Gly Phe Asn Leu	Glu Glu Ala Ala Arg	Cys Met Arg
1430	1435	1440
Ser Leu Lys Ala Pro Ala Val	Val Ser Val Ser Ser	Pro Asp Ala
1445	1450	1455
Val Thr Thr Tyr Asn Gly Tyr	Leu Thr Ser Ser Ser	Lys Thr Ser
1460	1465	1470
Glu Glu His Phe Val Glu Thr	Val Ser Leu Ala Gly	Ser Tyr Arg
1475	1480	1485
Asp Trp Ser Tyr Ser Gly Gln	Arg Thr Glu Leu Gly	Val Glu Phe
1490	1495	1500
Leu Lys Arg Gly Asp Lys Ile	Val Tyr His Thr Leu	Glu Ser Pro
1505	1510	1515
Val Glu Phe His Leu Asp Gly	Glu Val Leu Ser Leu	Asp Lys Leu
1520	1525	1530
Lys Ser Leu Leu Ser Leu Arg	Glu Val Lys Thr Ile	Lys Val Phe
1535	1540	1545
Thr Thr Val Asp Asn Thr Asn	Leu His Thr Gln Leu	Val Asp Met
1550	1555	1560
Ser Met Thr Tyr Gly Gln Gln	Phe Gly Pro Thr Tyr	Leu Asp Gly
1565	1570	1575
Ala Asp Val Thr Lys Ile Lys	Pro His Val Asn His	Glu Gly Lys
1580	1585	1590
Thr Phe Phe Val Leu Pro Ser	Asp Asp Thr Leu Arg	Ser Glu Ala
1595	1600	1605
Phe Glu Tyr Tyr His Thr Leu	Asp Glu Ser Phe Leu	Gly Arg Tyr
1610	1615	1620
Met Ser Ala Leu Asn His Thr	Lys Lys Trp Lys Phe	Pro Gln Val
1625	1630	1635
Gly Gly Leu Thr Ser Ile Lys	Trp Ala Asp Asn Asn	Cys Tyr Leu
1640	1645	1650
Ser Ser Val Leu Leu Ala Leu	Gln Gln Leu Glu Val	Lys Phe Asn
1655	1660	1665
Ala Pro Ala Leu Gln Glu Ala	Tyr Tyr Arg Ala Arg	Ala Gly Asp
1670	1675	1680
Ala Ala Asn Phe Cys Ala Leu	Ile Leu Ala Tyr Ser	Asn Lys Thr
1685	1690	1695
Val Gly Glu Leu Gly Asp Val	Arg Glu Thr Met Thr	His Leu Leu
1700	1705	1710
Gln His Ala Asn Leu Glu Ser	Ala Lys Arg Val Leu	Asn Val Val
1715	1720	1725
Cys Lys His Cys Gly Gln Lys	Thr Thr Thr Leu Thr	Gly Val Glu
1730	1735	1740
Ala Val Met Tyr Met Gly Thr	Leu Ser Tyr Asp Asn	Leu Lys Thr
1745	1750	1755
Gly Val Ser Ile Pro Cys Val	Cys Gly Arg Asp Ala	Thr Gln Tyr
1760	1765	1770
Leu Val Gln Gln Glu Ser Ser	Phe Val Met Met Ser	Ala Pro Pro
1775	1780	1785

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Ala Glu Tyr Lys Leu Gln Gln Gly Thr Phe Leu Cys Ala Asn Glu	1790	1795	1800
Tyr Thr Gly Asn Tyr Gln Cys Gly His Tyr Thr His Ile Thr Ala	1805	1810	1815
Lys Glu Thr Leu Tyr Arg Ile Asp Gly Ala His Leu Thr Lys Met	1820	1825	1830
Ser Glu Tyr Lys Gly Pro Val Thr Asp Val Phe Tyr Lys Glu Thr	1835	1840	1845
Ser Tyr Thr Thr Thr Ile Lys Pro Val Ser Tyr Lys Leu Asp Gly	1850	1855	1860
Val Thr Tyr Thr Glu Ile Glu Pro Lys Leu Asp Gly Tyr Tyr Lys	1865	1870	1875
Lys Asp Asn Ala Tyr Tyr Thr Glu Gln Pro Ile Asp Leu Val Pro	1880	1885	1890
Thr Gln Pro Leu Pro Asn Ala Ser Phe Asp Asn Phe Lys Leu Thr	1895	1900	1905
Cys Ser Asn Thr Lys Phe Ala Asp Asp Leu Asn Gln Met Thr Gly	1910	1915	1920
Phe Thr Lys Pro Ala Ser Arg Glu Leu Ser Val Thr Phe Phe Pro	1925	1930	1935
Asp Leu Asn Gly Asp Val Val Ala Ile Asp Tyr Arg His Tyr Ser	1940	1945	1950
Ala Ser Phe Lys Lys Gly Ala Lys Leu Leu His Lys Pro Ile Val	1955	1960	1965
Trp His Ile Asn Gln Ala Thr Thr Lys Thr Thr Phe Lys Pro Asn	1970	1975	1980
Thr Trp Cys Leu Arg Cys Leu Trp Ser Thr Lys Pro Val Asp Thr	1985	1990	1995
Ser Asn Ser Phe Glu Val Leu Ala Val Glu Asp Thr Gln Gly Met	2000	2005	2010
Asp Asn Leu Ala Cys Glu Ser Gln Gln Pro Thr Ser Glu Glu Val	2015	2020	2025
Val Glu Asn Pro Thr Ile Gln Lys Glu Val Ile Glu Cys Asp Val	2030	2035	2040
Lys Thr Thr Glu Val Val Gly Asn Val Ile Leu Lys Pro Ser Asp	2045	2050	2055
Glu Gly Val Lys Val Thr Gln Glu Leu Gly His Glu Asp Leu Met	2060	2065	2070
Ala Ala Tyr Val Glu Asn Thr Ser Ile Thr Ile Lys Lys Pro Asn	2075	2080	2085
Glu Leu Ser Leu Ala Leu Gly Leu Lys Thr Ile Ala Thr His Gly	2090	2095	2100
Ile Ala Ala Ile Asn Ser Val Pro Trp Ser Lys Ile Leu Ala Tyr	2105	2110	2115
Val Lys Pro Phe Leu Gly Gln Ala Ala Ile Thr Thr Ser Asn Cys	2120	2125	2130
Ala Lys Arg Leu Ala Gln Arg Val Phe Asn Asn Tyr Met Pro Tyr	2135	2140	2145
Val Phe Thr Leu Leu Phe Gln Leu Cys Thr Phe Thr Lys Ser Thr	2150	2155	2160

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Asn	Ser	Arg	Ile	Arg	Ala	Ser	Leu	Pro	Thr	Thr	Ile	Ala	Lys	Asn
2165						2170					2175			
Ser	Val	Lys	Ser	Val	Ala	Lys	Leu	Cys	Leu	Asp	Ala	Gly	Ile	Asn
2180						2185					2190			
Tyr	Val	Lys	Ser	Pro	Lys	Phe	Ser	Lys	Leu	Phe	Thr	Ile	Ala	Met
2195						2200					2205			
Trp	Leu	Leu	Leu	Leu	Ser	Ile	Cys	Leu	Gly	Ser	Leu	Ile	Cys	Val
2210						2215					2220			
Thr	Ala	Ala	Phe	Gly	Val	Leu	Leu	Ser	Asn	Phe	Gly	Ala	Pro	Ser
2225						2230					2235			
Tyr	Cys	Asn	Gly	Val	Arg	Glu	Leu	Tyr	Leu	Asn	Ser	Ser	Asn	Val
2240						2245					2250			
Thr	Thr	Met	Asp	Phe	Cys	Glu	Gly	Ser	Phe	Pro	Cys	Ser	Ile	Cys
2255						2260					2265			
Leu	Ser	Gly	Leu	Asp	Ser	Leu	Asp	Ser	Tyr	Pro	Ala	Leu	Glu	Thr
2270						2275					2280			
Ile	Gln	Val	Thr	Ile	Ser	Ser	Tyr	Lys	Leu	Asp	Leu	Thr	Ile	Leu
2285						2290					2295			
Gly	Leu	Ala	Ala	Glu	Trp	Val	Leu	Ala	Tyr	Met	Leu	Phe	Thr	Lys
2300						2305					2310			
Phe	Phe	Tyr	Leu	Leu	Gly	Leu	Ser	Ala	Ile	Met	Gln	Val	Phe	Phe
2315						2320					2325			
Gly	Tyr	Phe	Ala	Ser	His	Phe	Ile	Ser	Asn	Ser	Trp	Leu	Met	Trp
2330						2335					2340			
Phe	Ile	Ile	Ser	Ile	Val	Gln	Met	Ala	Pro	Val	Ser	Ala	Met	Val
2345						2350					2355			
Arg	Met	Tyr	Ile	Phe	Phe	Ala	Ser	Phe	Tyr	Tyr	Ile	Trp	Lys	Ser
2360						2365					2370			
Tyr	Val	His	Ile	Met	Asp	Gly	Cys	Thr	Ser	Ser	Thr	Cys	Met	Met
2375						2380					2385			
Cys	Tyr	Lys	Arg	Asn	Arg	Ala	Thr	Arg	Val	Glu	Cys	Thr	Thr	Ile
2390						2395					2400			
Val	Asn	Gly	Met	Lys	Arg	Ser	Phe	Tyr	Val	Tyr	Ala	Asn	Gly	Gly
2405						2410					2415			
Arg	Gly	Phe	Cys	Lys	Thr	His	Asn	Trp	Asn	Cys	Leu	Asn	Cys	Asp
2420						2425					2430			
Thr	Phe	Cys	Thr	Gly	Ser	Thr	Phe	Ile	Ser	Asp	Glu	Val	Ala	Arg
2435						2440					2445			
Asp	Leu	Ser	Leu	Gln	Phe	Lys	Arg	Pro	Ile	Asn	Pro	Thr	Asp	Gln
2450						2455					2460			
Ser	Ser	Tyr	Ile	Val	Asp	Ser	Val	Ala	Val	Lys	Asn	Gly	Ala	Leu
2465						2470					2475			
His	Leu	Tyr	Phe	Asp	Lys	Ala	Gly	Gln	Lys	Thr	Tyr	Glu	Arg	His
2480						2485					2490			
Pro	Leu	Ser	His	Phe	Val	Asn	Leu	Asp	Asn	Leu	Arg	Ala	Asn	Asn
2495						2500					2505			
Thr	Lys	Gly	Ser	Leu	Pro	Ile	Asn	Val	Ile	Val	Phe	Asp	Gly	Lys
2510						2515					2520			
Ser	Lys	Cys	Asp	Glu	Ser	Ala	Ser	Lys	Ser	Ala	Ser	Val	Tyr	Tyr
2525						2530					2535			
Ser	Gln	Leu	Met	Cys	Gln	Pro	Ile	Leu	Leu	Leu	Asp	Gln	Ala	Leu

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2540	2545	2550
Val Ser Asp Val Gly Asp	Ser Thr Glu Val Ser	Val Lys Met Phe
2555	2560	2565
Asp Ala Tyr Val Asp Thr	Phe Ser Ala Thr Phe	Ser Val Pro Met
2570	2575	2580
Glu Lys Leu Lys Ala Leu	Val Ala Thr Ala His	Ser Glu Leu Ala
2585	2590	2595
Lys Gly Val Ala Leu Asp	Gly Val Leu Ser Thr	Phe Val Ser Ala
2600	2605	2610
Ala Arg Gln Gly Val Val	Asp Thr Asp Val Asp	Thr Lys Asp Val
2615	2620	2625
Ile Glu Cys Leu Lys Leu	Ser His His Ser Asp	Leu Glu Val Thr
2630	2635	2640
Gly Asp Ser Cys Asn Asn	Phe Met Leu Thr Tyr	Asn Lys Val Glu
2645	2650	2655
Asn Met Thr Pro Arg Asp	Leu Gly Ala Cys Ile	Asp Cys Asn Ala
2660	2665	2670
Arg His Ile Asn Ala Gln	Val Ala Lys Ser His	Asn Val Ser Leu
2675	2680	2685
Ile Trp Asn Val Lys Asp	Tyr Met Ser Leu Ser	Glu Gln Leu Arg
2690	2695	2700
Lys Gln Ile Arg Ser Ala	Ala Lys Lys Asn Asn	Ile Pro Phe Arg
2705	2710	2715
Leu Thr Cys Ala Thr Thr	Arg Gln Val Val Asn	Val Ile Thr Thr
2720	2725	2730
Lys Ile Ser Leu Lys Gly	Gly Lys Ile Val Ser	Thr Cys Phe Lys
2735	2740	2745
Leu Met Leu Lys Ala Thr	Leu Leu Cys Val Leu	Ala Ala Leu Val
2750	2755	2760
Cys Tyr Ile Val Met Pro	Val His Thr Leu Ser	Ile His Asp Gly
2765	2770	2775
Tyr Thr Asn Glu Ile Ile	Gly Tyr Lys Ala Ile	Gln Asp Gly Val
2780	2785	2790
Thr Arg Asp Ile Ile Ser	Thr Asp Asp Cys Phe	Ala Asn Lys His
2795	2800	2805
Ala Gly Phe Asp Ala Trp	Phe Ser Gln Arg Gly	Gly Ser Tyr Lys
2810	2815	2820
Asn Asp Lys Ser Cys Pro	Val Val Ala Ala Ile	Ile Thr Arg Glu
2825	2830	2835
Ile Gly Phe Ile Val Pro	Gly Leu Pro Gly Thr	Val Leu Arg Ala
2840	2845	2850
Ile Asn Gly Asp Phe Leu	His Phe Leu Pro Arg	Val Phe Ser Ala
2855	2860	2865
Val Gly Asn Ile Cys Tyr	Thr Pro Ser Lys Leu	Ile Glu Tyr Ser
2870	2875	2880
Asp Phe Ala Thr Ser Ala	Cys Val Leu Ala Ala	Glu Cys Thr Ile
2885	2890	2895
Phe Lys Asp Ala Met Gly	Lys Pro Val Pro Tyr	Cys Tyr Asp Thr
2900	2905	2910
Asn Leu Leu Glu Gly Ser	Ile Ser Tyr Ser Glu	Leu Arg Pro Asp
2915	2920	2925

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Thr	Arg	Tyr	Val	Leu	Met	Asp	Gly	Ser	Ile	Ile	Gln	Phe	Pro	Asn
2930						2935					2940			
Thr	Tyr	Leu	Glu	Gly	Ser	Val	Arg	Val	Val	Thr	Thr	Phe	Asp	Ala
2945						2950					2955			
Glu	Tyr	Cys	Arg	His	Gly	Thr	Cys	Glu	Arg	Ser	Glu	Val	Gly	Ile
2960						2965					2970			
Cys	Leu	Ser	Thr	Ser	Gly	Arg	Trp	Val	Leu	Asn	Asn	Glu	His	Tyr
2975						2980					2985			
Arg	Ala	Leu	Ser	Gly	Val	Phe	Cys	Gly	Val	Asp	Ala	Met	Asn	Leu
2990						2995					3000			
Ile	Ala	Asn	Ile	Phe	Thr	Pro	Leu	Val	Gln	Pro	Val	Gly	Ala	Leu
3005						3010					3015			
Asp	Val	Ser	Ala	Ser	Val	Val	Ala	Gly	Gly	Ile	Ile	Ala	Ile	Leu
3020						3025					3030			
Val	Thr	Cys	Ala	Ala	Tyr	Tyr	Phe	Met	Lys	Phe	Arg	Arg	Val	Phe
3035						3040					3045			
Gly	Glu	Tyr	Asn	His	Val	Val	Ala	Ala	Asn	Ala	Leu	Leu	Phe	Leu
3050						3055					3060			
Met	Ser	Phe	Thr	Ile	Leu	Cys	Leu	Val	Pro	Ala	Tyr	Ser	Phe	Leu
3065						3070					3075			
Pro	Gly	Val	Tyr	Ser	Val	Phe	Tyr	Leu	Tyr	Leu	Thr	Phe	Tyr	Phe
3080						3085					3090			
Thr	Asn	Asp	Val	Ser	Phe	Leu	Ala	His	Leu	Gln	Trp	Phe	Ala	Met
3095						3100					3105			
Phe	Ser	Pro	Ile	Val	Pro	Phe	Trp	Ile	Thr	Ala	Ile	Tyr	Val	Phe
3110						3115					3120			
Cys	Ile	Ser	Leu	Lys	His	Cys	His	Trp	Phe	Phe	Asn	Asn	Tyr	Leu
3125						3130					3135			
Arg	Lys	Arg	Val	Met	Phe	Asn	Gly	Val	Thr	Phe	Ser	Thr	Phe	Glu
3140						3145					3150			
Glu	Ala	Ala	Leu	Cys	Thr	Phe	Leu	Leu	Asn	Lys	Glu	Met	Tyr	Leu
3155						3160					3165			
Lys	Leu	Arg	Ser	Glu	Thr	Leu	Leu	Pro	Leu	Thr	Gln	Tyr	Asn	Arg
3170						3175					3180			
Tyr	Leu	Ala	Leu	Tyr	Asn	Lys	Tyr	Lys	Tyr	Phe	Ser	Gly	Ala	Leu
3185						3190					3195			
Asp	Thr	Thr	Ser	Tyr	Arg	Glu	Ala	Ala	Cys	Cys	His	Leu	Ala	Lys
3200						3205					3210			
Ala	Leu	Asn	Asp	Phe	Ser	Asn	Ser	Gly	Ala	Asp	Val	Leu	Tyr	Gln
3215						3220					3225			
Pro	Pro	Gln	Thr	Ser	Ile	Thr	Ser	Ala	Val	Leu	Gln	Ser	Gly	Phe
3230						3235					3240			
Arg	Lys	Met	Ala	Phe	Pro	Ser	Gly	Lys	Val	Glu	Gly	Cys	Met	Val
3245						3250					3255			
Gln	Val	Thr	Cys	Gly	Thr	Thr	Thr	Leu	Asn	Gly	Leu	Trp	Leu	Asp
3260						3265					3270			
Asp	Thr	Val	Tyr	Cys	Pro	Arg	His	Val	Ile	Cys	Thr	Ala	Glu	Asp
3275						3280					3285			
Met	Leu	Asn	Pro	Asn	Tyr	Glu	Asp	Leu	Leu	Ile	Arg	Lys	Ser	Asn
3290						3295					3300			

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His	Ser	Phe	Leu	Val	Gln	Ala	Gly	Asn	Val	Gln	Leu	Arg	Val	Ile
3305						3310					3315			
Gly	His	Ser	Met	Gln	Asn	Cys	Leu	Leu	Arg	Leu	Lys	Val	Asp	Thr
3320						3325					3330			
Ser	Asn	Pro	Lys	Thr	Pro	Lys	Tyr	Lys	Phe	Val	Arg	Ile	Gln	Pro
3335						3340					3345			
Gly	Gln	Thr	Phe	Ser	Val	Leu	Ala	Cys	Tyr	Asn	Gly	Ser	Pro	Ser
3350						3355					3360			
Gly	Val	Tyr	Gln	Cys	Ala	Met	Arg	Pro	Asn	His	Thr	Ile	Lys	Gly
3365						3370					3375			
Ser	Phe	Leu	Asn	Gly	Ser	Cys	Gly	Ser	Val	Gly	Phe	Asn	Ile	Asp
3380						3385					3390			
Tyr	Asp	Cys	Val	Ser	Phe	Cys	Tyr	Met	His	His	Met	Glu	Leu	Pro
3395						3400					3405			
Thr	Gly	Val	His	Ala	Gly	Thr	Asp	Leu	Glu	Gly	Lys	Phe	Tyr	Gly
3410						3415					3420			
Pro	Phe	Val	Asp	Arg	Gln	Thr	Ala	Gln	Ala	Ala	Gly	Thr	Asp	Thr
3425						3430					3435			
Thr	Ile	Thr	Leu	Asn	Val	Leu	Ala	Trp	Leu	Tyr	Ala	Ala	Val	Ile
3440						3445					3450			
Asn	Gly	Asp	Arg	Trp	Phe	Leu	Asn	Arg	Phe	Thr	Thr	Thr	Leu	Asn
3455						3460					3465			
Asp	Phe	Asn	Leu	Val	Ala	Met	Lys	Tyr	Asn	Tyr	Glu	Pro	Leu	Thr
3470						3475					3480			
Gln	Asp	His	Val	Asp	Ile	Leu	Gly	Pro	Leu	Ser	Ala	Gln	Thr	Gly
3485						3490					3495			
Ile	Ala	Val	Leu	Asp	Met	Cys	Ala	Ala	Leu	Lys	Glu	Leu	Leu	Gln
3500						3505					3510			
Asn	Gly	Met	Asn	Gly	Arg	Thr	Ile	Leu	Gly	Ser	Thr	Ile	Leu	Glu
3515						3520					3525			
Asp	Glu	Phe	Thr	Pro	Phe	Asp	Val	Val	Arg	Gln	Cys	Ser	Gly	Val
3530						3535					3540			
Thr	Phe	Gln	Gly	Lys	Phe	Lys	Lys	Ile	Val	Lys	Gly	Thr	His	His
3545						3550					3555			
Trp	Met	Leu	Leu	Thr	Phe	Leu	Thr	Ser	Leu	Leu	Ile	Leu	Val	Gln
3560						3565					3570			
Ser	Thr	Gln	Trp	Ser	Leu	Phe	Phe	Phe	Val	Tyr	Glu	Asn	Ala	Phe
3575						3580					3585			
Leu	Pro	Phe	Thr	Leu	Gly	Ile	Met	Ala	Ile	Ala	Ala	Cys	Ala	Met
3590						3595					3600			
Leu	Leu	Val	Lys	His	Lys	His	Ala	Phe	Leu	Cys	Leu	Phe	Leu	Leu
3605						3610					3615			
Pro	Ser	Leu	Ala	Thr	Val	Ala	Tyr	Phe	Asn	Met	Val	Tyr	Met	Pro
3620						3625					3630			
Ala	Ser	Trp	Val	Met	Arg	Ile	Met	Thr	Trp	Leu	Glu	Leu	Ala	Asp
3635						3640					3645			
Thr	Ser	Leu	Ser	Gly	Tyr	Arg	Leu	Lys	Asp	Cys	Val	Met	Tyr	Ala
3650						3655					3660			
Ser	Ala	Leu	Val	Leu	Leu	Ile	Leu	Met	Thr	Ala	Arg	Thr	Val	Tyr
3665						3670					3675			
Asp	Asp	Ala	Ala	Arg	Arg	Val	Trp	Thr	Leu	Met	Asn	Val	Ile	Thr

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3680	3685	3690
Leu Val Tyr Lys Val Tyr Tyr	Gly Asn Ala Leu Asp	Gln Ala Ile
3695	3700	3705
Ser Met Trp Ala Leu Val Ile	Ser Val Thr Ser Asn Tyr Ser Gly	
3710	3715	3720
Val Val Thr Thr Ile Met Phe	Leu Ala Arg Ala Ile	Val Phe Val
3725	3730	3735
Cys Val Glu Tyr Tyr Pro Leu	Leu Phe Ile Thr Gly	Asn Thr Leu
3740	3745	3750
Gln Cys Ile Met Leu Val Tyr	Cys Phe Leu Gly Tyr	Cys Cys Cys
3755	3760	3765
Cys Tyr Phe Gly Leu Phe Cys	Leu Leu Asn Arg Tyr	Phe Arg Leu
3770	3775	3780
Thr Leu Gly Val Tyr Asp Tyr	Leu Val Ser Thr Gln	Glu Phe Arg
3785	3790	3795
Tyr Met Asn Ser Gln Gly Leu	Leu Pro Pro Lys Ser	Ser Ile Asp
3800	3805	3810
Ala Phe Lys Leu Asn Ile Lys	Leu Leu Gly Ile Gly	Gly Lys Pro
3815	3820	3825
Cys Ile Lys Val Ala Thr Val	Gln Ser Lys Met Ser	Asp Val Lys
3830	3835	3840
Cys Thr Ser Val Val Leu Leu	Ser Val Leu Gln Gln	Leu Arg Val
3845	3850	3855
Glu Ser Ser Ser Lys Leu Trp	Ala Gln Cys Val Gln	Leu His Asn
3860	3865	3870
Asp Ile Leu Leu Ala Lys Asp	Thr Thr Glu Ala Phe	Glu Lys Met
3875	3880	3885
Val Ser Leu Leu Ser Val Leu	Leu Ser Met Gln Gly	Ala Val Asp
3890	3895	3900
Ile Asn Arg Leu Cys Glu Glu	Met Leu Asp Asn Arg	Ala Thr Leu
3905	3910	3915
Gln Ala Ile Ala Ser Glu Phe	Ser Ser Leu Pro Ser	Tyr Ala Ala
3920	3925	3930
Tyr Ala Thr Ala Gln Glu Ala	Tyr Glu Gln Ala Val	Ala Asn Gly
3935	3940	3945
Asp Ser Glu Val Val Leu Lys	Lys Leu Lys Lys Ser	Leu Asn Val
3950	3955	3960
Ala Lys Ser Glu Phe Asp Arg	Asp Ala Ala Met Gln	Arg Lys Leu
3965	3970	3975
Glu Lys Met Ala Asp Gln Ala	Met Thr Gln Met Tyr	Lys Gln Ala
3980	3985	3990
Arg Ser Glu Asp Lys Arg Ala	Lys Val Thr Ser Ala	Met Gln Thr
3995	4000	4005
Met Leu Phe Thr Met Leu Arg	Lys Leu Asp Asn Asp	Ala Leu Asn
4010	4015	4020
Asn Ile Ile Asn Asn Ala Arg	Asp Gly Cys Val Pro	Leu Asn Ile
4025	4030	4035
Ile Pro Leu Thr Thr Ala Ala	Lys Leu Met Val Val	Val Pro Asp
4040	4045	4050
Tyr Gly Thr Tyr Lys Asn Thr	Cys Asp Gly Asn Thr	Phe Thr Tyr
4055	4060	4065

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Ala Ser	Ala Leu Trp Glu Ile	Gln Gln Val Val Asp	Ala Asp Ser
4070	4075	4080	
Lys Ile	Val Gln Leu Ser Glu	Ile Asn Met Asp Asn	Ser Pro Asn
4085	4090	4095	
Leu Ala	Trp Pro Leu Ile Val	Thr Ala Leu Arg Ala	Asn Ser Ala
4100	4105	4110	
Val Lys	Leu Gln Asn Asn Glu	Leu Ser Pro Val Ala	Leu Arg Gln
4115	4120	4125	
Met Ser	Cys Ala Ala Gly Thr	Thr Gln Thr Ala Cys	Thr Asp Asp
4130	4135	4140	
Asn Ala	Leu Ala Tyr Tyr Asn	Asn Ser Lys Gly Gly	Arg Phe Val
4145	4150	4155	
Leu Ala	Leu Leu Ser Asp His	Gln Asp Leu Lys Trp	Ala Arg Phe
4160	4165	4170	
Pro Lys	Ser Asp Gly Thr Gly	Thr Ile Tyr Thr Glu	Leu Glu Pro
4175	4180	4185	
Pro Cys	Arg Phe Val Thr Asp	Thr Pro Lys Gly Pro	Lys Val Lys
4190	4195	4200	
Tyr Leu	Tyr Phe Ile Lys Gly	Leu Asn Asn Leu Asn	Arg Gly Met
4205	4210	4215	
Val Leu	Gly Ser Leu Ala Ala	Thr Val Arg Leu Gln	Ala Gly Asn
4220	4225	4230	
Ala Thr	Glu Val Pro Ala Asn	Ser Thr Val Leu Ser	Phe Cys Ala
4235	4240	4245	
Phe Ala	Val Asp Pro Ala Lys	Ala Tyr Lys Asp Tyr	Leu Ala Ser
4250	4255	4260	
Gly Gly	Gln Pro Ile Thr Asn	Cys Val Lys Met Leu	Cys Thr His
4265	4270	4275	
Thr Gly	Thr Gly Gln Ala Ile	Thr Val Thr Pro Glu	Ala Asn Met
4280	4285	4290	
Asp Gln	Glu Ser Phe Gly Gly	Ala Ser Cys Cys Leu	Tyr Cys Arg
4295	4300	4305	
Cys His	Ile Asp His Pro Asn	Pro Lys Gly Phe Cys	Asp Leu Lys
4310	4315	4320	
Gly Lys	Tyr Val Gln Ile Pro	Thr Thr Cys Ala Asn	Asp Pro Val
4325	4330	4335	
Gly Phe	Thr Leu Arg Asn Thr	Val Cys Thr Val Cys	Gly Met Trp
4340	4345	4350	
Lys Gly	Tyr Gly Cys Ser Cys	Asp Gln Leu Arg Glu	Pro Leu Met
4355	4360	4365	
Gln Ser	Ala Asp Ala Ser Thr	Phe Leu Asn Gly Phe	Ala Val
4370	4375	4380	

<210> SEQ ID NO 75

<211> LENGTH: 2695

<212> TYPE: PRT

<213> ORGANISM: CORONAVIRUS

<400> SEQUENCE: 75

Arg Val Cys Gly Val Ser Ala Ala Arg Leu Thr Pro Cys Gly Thr Gly
1 5 10 15

Thr Ser Thr Asp Val Val Tyr Arg Ala Phe Asp Ile Tyr Asn Glu Lys

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20							25					30				
Val	Ala	Gly	Phe	Ala	Lys	Phe	Leu	Lys	Thr	Asn	Cys	Cys	Arg	Phe	Gln	
	35						40					45				
Glu	Lys	Asp	Glu	Glu	Gly	Asn	Leu	Leu	Asp	Ser	Tyr	Phe	Val	Val	Lys	
	50					55					60					
Arg	His	Thr	Met	Ser	Asn	Tyr	Gln	His	Glu	Glu	Thr	Ile	Tyr	Asn	Leu	
65					70				75					80		
Val	Lys	Asp	Cys	Pro	Ala	Val	Ala	Val	His	Asp	Phe	Phe	Lys	Phe	Arg	
				85					90					95		
Val	Asp	Gly	Asp	Met	Val	Pro	His	Ile	Ser	Arg	Gln	Arg	Leu	Thr	Lys	
			100					105					110			
Tyr	Thr	Met	Ala	Asp	Leu	Val	Tyr	Ala	Leu	Arg	His	Phe	Asp	Glu	Gly	
		115					120					125				
Asn	Cys	Asp	Thr	Leu	Lys	Glu	Ile	Leu	Val	Thr	Tyr	Asn	Cys	Cys	Asp	
	130					135					140					
Asp	Asp	Tyr	Phe	Asn	Lys	Lys	Asp	Trp	Tyr	Asp	Phe	Val	Glu	Asn	Pro	
145					150					155					160	
Asp	Ile	Leu	Arg	Val	Tyr	Ala	Asn	Leu	Gly	Glu	Arg	Val	Arg	Gln	Ser	
				165					170					175		
Leu	Leu	Lys	Thr	Val	Gln	Phe	Cys	Asp	Ala	Met	Arg	Asp	Ala	Gly	Ile	
			180						185				190			
Val	Gly	Val	Leu	Thr	Leu	Asp	Asn	Gln	Asp	Leu	Asn	Gly	Asn	Trp	Tyr	
	195						200					205				
Asp	Phe	Gly	Asp	Phe	Val	Gln	Val	Ala	Pro	Gly	Cys	Gly	Val	Pro	Ile	
	210					215					220					
Val	Asp	Ser	Tyr	Tyr	Ser	Leu	Leu	Met	Pro	Ile	Leu	Thr	Leu	Thr	Arg	
225					230					235					240	
Ala	Leu	Ala	Ala	Glu	Ser	His	Met	Asp	Ala	Asp	Leu	Ala	Lys	Pro	Leu	
				245					250					255		
Ile	Lys	Trp	Asp	Leu	Leu	Lys	Tyr	Asp	Phe	Thr	Glu	Glu	Arg	Leu	Cys	
			260					265					270			
Leu	Phe	Asp	Arg	Tyr	Phe	Lys	Tyr	Trp	Asp	Gln	Thr	Tyr	His	Pro	Asn	
	275						280					285				
Cys	Ile	Asn	Cys	Leu	Asp	Asp	Arg	Cys	Ile	Leu	His	Cys	Ala	Asn	Phe	
	290					295					300					
Asn	Val	Leu	Phe	Ser	Thr	Val	Phe	Pro	Pro	Thr	Ser	Phe	Gly	Pro	Leu	
305					310					315					320	
Val	Arg	Lys	Ile	Phe	Val	Asp	Gly	Val	Pro	Phe	Val	Val	Ser	Thr	Gly	
			325						330					335		
Tyr	His	Phe	Arg	Glu	Leu	Gly	Val	Val	His	Asn	Gln	Asp	Val	Asn	Leu	
			340					345					350			
His	Ser	Ser	Arg	Leu	Ser	Phe	Lys	Glu	Leu	Leu	Val	Tyr	Ala	Ala	Asp	
	355						360					365				
Pro	Ala	Met	His	Ala	Ala	Ser	Gly	Asn	Leu	Leu	Leu	Asp	Lys	Arg	Thr	
	370					375					380					
Thr	Cys	Phe	Ser	Val	Ala	Ala	Leu	Thr	Asn	Asn	Val	Ala	Phe	Gln	Thr	
385					390					395					400	
Val	Lys	Pro	Gly	Asn	Phe	Asn	Lys	Asp	Phe	Tyr	Asp	Phe	Ala	Val	Ser	
			405						410				415			
Lys	Gly	Phe	Phe	Lys	Glu	Gly	Ser	Ser	Val	Glu	Leu	Lys	His	Phe	Phe	
			420					425					430			

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Phe Ala Gln Asp Gly Asn Ala Ala Ile Ser Asp Tyr Asp Tyr Tyr Arg
 435 440 445
 Tyr Asn Leu Pro Thr Met Cys Asp Ile Arg Gln Leu Leu Phe Val Val
 450 455 460
 Glu Val Val Asp Lys Tyr Phe Asp Cys Tyr Asp Gly Gly Cys Ile Asn
 465 470 475 480
 Ala Asn Gln Val Ile Val Asn Asn Leu Asp Lys Ser Ala Gly Phe Pro
 485 490 495
 Phe Asn Lys Trp Gly Lys Ala Arg Leu Tyr Tyr Asp Ser Met Ser Tyr
 500 505 510
 Glu Asp Gln Asp Ala Leu Phe Ala Tyr Thr Lys Arg Asn Val Ile Pro
 515 520 525
 Thr Ile Thr Gln Met Asn Leu Lys Tyr Ala Ile Ser Ala Lys Asn Arg
 530 535 540
 Ala Arg Thr Val Ala Gly Val Ser Ile Cys Ser Thr Met Thr Asn Arg
 545 550 555 560
 Gln Phe His Gln Lys Leu Leu Lys Ser Ile Ala Ala Thr Arg Gly Ala
 565 570 575
 Thr Val Val Ile Gly Thr Ser Lys Phe Tyr Gly Gly Trp His Asn Met
 580 585 590
 Leu Lys Thr Val Tyr Ser Asp Val Glu Thr Pro His Leu Met Gly Trp
 595 600 605
 Asp Tyr Pro Lys Cys Asp Arg Ala Met Pro Asn Met Leu Arg Ile Met
 610 615 620
 Ala Ser Leu Val Leu Ala Arg Lys His Asn Thr Cys Cys Asn Leu Ser
 625 630 635 640
 His Arg Phe Tyr Arg Leu Ala Asn Glu Cys Ala Gln Val Leu Ser Glu
 645 650 655
 Met Val Met Cys Gly Gly Ser Leu Tyr Val Lys Pro Gly Gly Thr Ser
 660 665 670
 Ser Gly Asp Ala Thr Thr Ala Tyr Ala Asn Ser Val Phe Asn Ile Cys
 675 680 685
 Gln Ala Val Thr Ala Asn Val Asn Ala Leu Leu Ser Thr Asp Gly Asn
 690 695 700
 Lys Ile Ala Asp Lys Tyr Val Arg Asn Leu Gln His Arg Leu Tyr Glu
 705 710 715 720
 Cys Leu Tyr Arg Asn Arg Asp Val Asp His Glu Phe Val Asp Glu Phe
 725 730 735
 Tyr Ala Tyr Leu Arg Lys His Phe Ser Met Met Ile Leu Ser Asp Asp
 740 745 750
 Ala Val Val Cys Tyr Asn Ser Asn Tyr Ala Ala Gln Gly Leu Val Ala
 755 760 765
 Ser Ile Lys Asn Phe Lys Ala Val Leu Tyr Tyr Gln Asn Asn Val Phe
 770 775 780
 Met Ser Glu Ala Lys Cys Trp Thr Glu Thr Asp Leu Thr Lys Gly Pro
 785 790 795 800
 His Glu Phe Cys Ser Gln His Thr Met Leu Val Lys Gln Gly Asp Asp
 805 810 815
 Tyr Val Tyr Leu Pro Tyr Pro Asp Pro Ser Arg Ile Leu Gly Ala Gly
 820 825 830

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Cys	Phe	Val	Asp	Asp	Ile	Val	Lys	Thr	Asp	Gly	Thr	Leu	Met	Ile	Glu	835	840	845
Arg	Phe	Val	Ser	Leu	Ala	Ile	Asp	Ala	Tyr	Pro	Leu	Thr	Lys	His	Pro	850	855	860
Asn	Gln	Glu	Tyr	Ala	Asp	Val	Phe	His	Leu	Tyr	Leu	Gln	Tyr	Ile	Arg	865	870	875
Lys	Leu	His	Asp	Glu	Leu	Thr	Gly	His	Met	Leu	Asp	Met	Tyr	Ser	Val	885	890	895
Met	Leu	Thr	Asn	Asp	Asn	Thr	Ser	Arg	Tyr	Trp	Glu	Pro	Glu	Phe	Tyr	900	905	910
Glu	Ala	Met	Tyr	Thr	Pro	His	Thr	Val	Leu	Gln	Ala	Val	Gly	Ala	Cys	915	920	925
Val	Leu	Cys	Asn	Ser	Gln	Thr	Ser	Leu	Arg	Cys	Gly	Ala	Cys	Ile	Arg	930	935	940
Arg	Pro	Phe	Leu	Cys	Cys	Lys	Cys	Cys	Tyr	Asp	His	Val	Ile	Ser	Thr	945	950	955
Ser	His	Lys	Leu	Val	Leu	Ser	Val	Asn	Pro	Tyr	Val	Cys	Asn	Ala	Pro	965	970	975
Gly	Cys	Asp	Val	Thr	Asp	Val	Thr	Gln	Leu	Tyr	Leu	Gly	Gly	Met	Ser	980	985	990
Tyr	Tyr	Cys	Lys	Ser	His	Lys	Pro	Pro	Ile	Ser	Phe	Pro	Leu	Cys	Ala	995	1000	1005
Asn	Gly	Gln	Val	Phe	Gly	Leu	Tyr	Lys	Asn	Thr	Cys	Val	Gly	Ser		1010	1015	1020
Asp	Asn	Val	Thr	Asp	Phe	Asn	Ala	Ile	Ala	Thr	Cys	Asp	Trp	Thr		1025	1030	1035
Asn	Ala	Gly	Asp	Tyr	Ile	Leu	Ala	Asn	Thr	Cys	Thr	Glu	Arg	Leu		1040	1045	1050
Lys	Leu	Phe	Ala	Ala	Glu	Thr	Leu	Lys	Ala	Thr	Glu	Glu	Thr	Phe		1055	1060	1065
Lys	Leu	Ser	Tyr	Gly	Ile	Ala	Thr	Val	Arg	Glu	Val	Leu	Ser	Asp		1070	1075	1080
Arg	Glu	Leu	His	Leu	Ser	Trp	Glu	Val	Gly	Lys	Pro	Arg	Pro	Pro		1085	1090	1095
Leu	Asn	Arg	Asn	Tyr	Val	Phe	Thr	Gly	Tyr	Arg	Val	Thr	Lys	Asn		1100	1105	1110
Ser	Lys	Val	Gln	Ile	Gly	Glu	Tyr	Thr	Phe	Glu	Lys	Gly	Asp	Tyr		1115	1120	1125
Gly	Asp	Ala	Val	Val	Tyr	Arg	Gly	Thr	Thr	Thr	Tyr	Lys	Leu	Asn		1130	1135	1140
Val	Gly	Asp	Tyr	Phe	Val	Leu	Thr	Ser	His	Thr	Val	Met	Pro	Leu		1145	1150	1155
Ser	Ala	Pro	Thr	Leu	Val	Pro	Gln	Glu	His	Tyr	Val	Arg	Ile	Thr		1160	1165	1170
Gly	Leu	Tyr	Pro	Thr	Leu	Asn	Ile	Ser	Asp	Glu	Phe	Ser	Ser	Asn		1175	1180	1185
Val	Ala	Asn	Tyr	Gln	Lys	Val	Gly	Met	Gln	Lys	Tyr	Ser	Thr	Leu		1190	1195	1200
Gln	Gly	Pro	Pro	Gly	Thr	Gly	Lys	Ser	His	Phe	Ala	Ile	Gly	Leu		1205	1210	1215
Ala	Leu	Tyr	Tyr	Pro	Ser	Ala	Arg	Ile	Val	Tyr	Thr	Ala	Cys	Ser				

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1220	1225	1230
His Ala Ala Val Asp Ala Leu Cys Glu Lys Ala Leu Lys Tyr Leu		
1235	1240	1245
Pro Ile Asp Lys Cys Ser Arg Ile Ile Pro Ala Arg Ala Arg Val		
1250	1255	1260
Glu Cys Phe Asp Lys Phe Lys Val Asn Ser Thr Leu Glu Gln Tyr		
1265	1270	1275
Val Phe Cys Thr Val Asn Ala Leu Pro Glu Thr Thr Ala Asp Ile		
1280	1285	1290
Val Val Phe Asp Glu Ile Ser Met Ala Thr Asn Tyr Asp Leu Ser		
1295	1300	1305
Val Val Asn Ala Arg Leu Arg Ala Lys His Tyr Val Tyr Ile Gly		
1310	1315	1320
Asp Pro Ala Gln Leu Pro Ala Pro Arg Thr Leu Leu Thr Lys Gly		
1325	1330	1335
Thr Leu Glu Pro Glu Tyr Phe Asn Ser Val Cys Arg Leu Met Lys		
1340	1345	1350
Thr Ile Gly Pro Asp Met Phe Leu Gly Thr Cys Arg Arg Cys Pro		
1355	1360	1365
Ala Glu Ile Val Asp Thr Val Ser Ala Leu Val Tyr Asp Asn Lys		
1370	1375	1380
Leu Lys Ala His Lys Asp Lys Ser Ala Gln Cys Phe Lys Met Phe		
1385	1390	1395
Tyr Lys Gly Val Ile Thr His Asp Val Ser Ser Ala Ile Asn Arg		
1400	1405	1410
Pro Gln Ile Gly Val Val Arg Glu Phe Leu Thr Arg Asn Pro Ala		
1415	1420	1425
Trp Arg Lys Ala Val Phe Ile Ser Pro Tyr Asn Ser Gln Asn Ala		
1430	1435	1440
Val Ala Ser Lys Ile Leu Gly Leu Pro Thr Gln Thr Val Asp Ser		
1445	1450	1455
Ser Gln Gly Ser Glu Tyr Asp Tyr Val Ile Phe Thr Gln Thr Thr		
1460	1465	1470
Glu Thr Ala His Ser Cys Asn Val Asn Arg Phe Asn Val Ala Ile		
1475	1480	1485
Thr Arg Ala Lys Ile Gly Ile Leu Cys Ile Met Ser Asp Arg Asp		
1490	1495	1500
Leu Tyr Asp Lys Leu Gln Phe Thr Ser Leu Glu Ile Pro Arg Arg		
1505	1510	1515
Asn Val Ala Thr Leu Gln Ala Glu Asn Val Thr Gly Leu Phe Lys		
1520	1525	1530
Asp Cys Ser Lys Ile Ile Thr Gly Leu His Pro Thr Gln Ala Pro		
1535	1540	1545
Thr His Leu Ser Val Asp Ile Lys Phe Lys Thr Glu Gly Leu Cys		
1550	1555	1560
Val Asp Ile Pro Gly Ile Pro Lys Asp Met Thr Tyr Arg Arg Leu		
1565	1570	1575
Ile Ser Met Met Gly Phe Lys Met Asn Tyr Gln Val Asn Gly Tyr		
1580	1585	1590
Pro Asn Met Phe Ile Thr Arg Glu Glu Ala Ile Arg His Val Arg		
1595	1600	1605

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Ala Trp	Ile Gly Phe Asp	Val	Glu Gly Cys His	Ala	Thr Arg Asp
1610		1615		1620	
Ala Val	Gly Thr Asn Leu	Pro	Leu Gln Leu Gly Phe	Ser Thr Gly	
1625		1630		1635	
Val Asn	Leu Val Ala Val	Pro	Thr Gly Tyr Val	Asp	Thr Glu Asn
1640		1645		1650	
Asn Thr	Glu Phe Thr Arg	Val	Asn Ala Lys Pro	Pro	Pro Gly Asp
1655		1660		1665	
Gln Phe	Lys His Leu Ile	Pro	Leu Met Tyr Lys	Gly	Leu Pro Trp
1670		1675		1680	
Asn Val	Val Arg Ile Lys	Ile	Val Gln Met Leu	Ser	Asp Thr Leu
1685		1690		1695	
Lys Gly	Leu Ser Asp Arg	Val	Val Phe Val Leu	Trp	Ala His Gly
1700		1705		1710	
Phe Glu	Leu Thr Ser Met	Lys	Tyr Phe Val Lys	Ile	Gly Pro Glu
1715		1720		1725	
Arg Thr	Cys Cys Leu Cys	Asp	Lys Arg Ala Thr	Cys	Phe Ser Thr
1730		1735		1740	
Ser Ser	Asp Thr Tyr Ala	Cys	Trp Asn His Ser	Val	Gly Phe Asp
1745		1750		1755	
Tyr Val	Tyr Asn Pro Phe	Met	Ile Asp Val Gln	Gln	Trp Gly Phe
1760		1765		1770	
Thr Gly	Asn Leu Gln Ser	Asn	His Asp Gln His	Cys	Gln Val His
1775		1780		1785	
Gly Asn	Ala His Val Ala	Ser	Cys Asp Ala Ile	Met	Thr Arg Cys
1790		1795		1800	
Leu Ala	Val His Glu Cys	Phe	Val Lys Arg Val	Asp	Trp Ser Val
1805		1810		1815	
Glu Tyr	Pro Ile Ile Gly	Asp	Glu Leu Arg Val	Asn	Ser Ala Cys
1820		1825		1830	
Arg Lys	Val Gln His Met	Val	Val Lys Ser Ala	Leu	Leu Ala Asp
1835		1840		1845	
Lys Phe	Pro Val Leu His	Asp	Ile Gly Asn Pro	Lys	Ala Ile Lys
1850		1855		1860	
Cys Val	Pro Gln Ala Glu	Val	Glu Trp Lys Phe	Tyr	Asp Ala Gln
1865		1870		1875	
Pro Cys	Ser Asp Lys Ala	Tyr	Lys Ile Glu Glu	Leu	Phe Tyr Ser
1880		1885		1890	
Tyr Ala	Thr His His Asp	Lys	Phe Thr Asp Gly	Val	Cys Leu Phe
1895		1900		1905	
Trp Asn	Cys Asn Val Asp	Arg	Tyr Pro Ala Asn	Ala	Ile Val Cys
1910		1915		1920	
Arg Phe	Asp Thr Arg Val	Leu	Ser Asn Leu Asn	Leu	Pro Gly Cys
1925		1930		1935	
Asp Gly	Gly Ser Leu Tyr	Val	Asn Lys His Ala	Phe	His Thr Pro
1940		1945		1950	
Ala Phe	Asp Lys Ser Ala	Phe	Thr Asn Leu Lys	Gln	Leu Pro Phe
1955		1960		1965	
Phe Tyr	Tyr Ser Asp Ser	Pro	Cys Glu Ser His	Gly	Lys Gln Val
1970		1975		1980	

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Val 1985	Ser	Asp	Ile	Asp	Tyr	Val 1990	Pro	Leu	Lys	Ser	Ala 1995	Thr	Cys	Ile
Thr 2000	Arg	Cys	Asn	Leu	Gly	Gly 2005	Ala	Val	Cys	Arg	His 2010	His	Ala	Asn
Glu 2015	Tyr	Arg	Gln	Tyr	Leu	Asp 2020	Ala	Tyr	Asn	Met	Met 2025	Ile	Ser	Ala
Gly 2030	Phe	Ser	Leu	Trp	Ile	Tyr 2035	Lys	Gln	Phe	Asp	Thr 2040	Tyr	Asn	Leu
Trp 2045	Asn	Thr	Phe	Thr	Arg	Leu 2050	Gln	Ser	Leu	Glu	Asn 2055	Val	Ala	Tyr
Asn 2060	Val	Val	Asn	Lys	Gly	His 2065	Phe	Asp	Gly	His	Ala 2070	Gly	Glu	Ala
Pro 2075	Val	Ser	Ile	Ile	Asn	Asn 2080	Ala	Val	Tyr	Thr	Lys 2085	Val	Asp	Gly
Ile 2090	Asp	Val	Glu	Ile	Phe	Glu 2095	Asn	Lys	Thr	Thr	Leu 2100	Pro	Val	Asn
Val 2105	Ala	Phe	Glu	Leu	Trp	Ala 2110	Lys	Arg	Asn	Ile	Lys 2115	Pro	Val	Pro
Glu 2120	Ile	Lys	Ile	Leu	Asn	Asn 2125	Leu	Gly	Val	Asp	Ile 2130	Ala	Ala	Asn
Thr 2135	Val	Ile	Trp	Asp	Tyr	Lys 2140	Arg	Glu	Ala	Pro	Ala 2145	His	Val	Ser
Thr 2150	Ile	Gly	Val	Cys	Thr	Met 2155	Thr	Asp	Ile	Ala	Lys 2160	Lys	Pro	Thr
Glu 2165	Ser	Ala	Cys	Ser	Ser	Leu 2170	Thr	Val	Leu	Phe	Asp 2175	Gly	Arg	Val
Glu 2180	Gly	Gln	Val	Asp	Leu	Phe 2185	Arg	Asn	Ala	Arg	Asn 2190	Gly	Val	Leu
Ile 2195	Thr	Glu	Gly	Ser	Val	Lys 2200	Gly	Leu	Thr	Pro	Ser 2205	Lys	Gly	Pro
Ala 2210	Gln	Ala	Ser	Val	Asn	Gly 2215	Val	Thr	Leu	Ile	Gly 2220	Glu	Ser	Val
Lys 2225	Thr	Gln	Phe	Asn	Tyr	Phe 2230	Lys	Lys	Val	Asp	Gly 2235	Ile	Ile	Gln
Gln 2240	Leu	Pro	Glu	Thr	Tyr	Phe 2245	Thr	Gln	Ser	Arg	Asp 2250	Leu	Glu	Asp
Phe 2255	Lys	Pro	Arg	Ser	Gln	Met 2260	Glu	Thr	Asp	Phe	Leu 2265	Glu	Leu	Ala
Met 2270	Asp	Glu	Phe	Ile	Gln	Arg 2275	Tyr	Lys	Leu	Glu	Gly 2280	Tyr	Ala	Phe
Glu 2285	His	Ile	Val	Tyr	Gly	Asp 2290	Phe	Ser	His	Gly	Gln 2295	Leu	Gly	Gly
Leu 2300	His	Leu	Met	Ile	Gly	Leu 2305	Ala	Lys	Arg	Ser	Gln 2310	Asp	Ser	Pro
Leu 2315	Lys	Leu	Glu	Asp	Phe	Ile 2320	Pro	Met	Asp	Ser	Thr 2325	Val	Lys	Asn
Tyr 2330	Phe	Ile	Thr	Asp	Ala	Gln 2335	Thr	Gly	Ser	Ser	Lys 2340	Cys	Val	Cys
Ser 2345	Val	Ile	Asp	Leu	Leu	Leu 2350	Asp	Asp	Phe	Val	Glu 2355	Ile	Ile	Lys
Ser	Gln	Asp	Leu	Ser	Val	Ile	Ser	Lys	Val	Val	Lys	Val	Thr	Ile

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2360	2365	2370
Asp Tyr Ala Glu Ile Ser Phe	Met Leu Trp Cys Lys	Asp Gly His
2375	2380	2385
Val Glu Thr Phe Tyr Pro Lys	Leu Gln Ala Ser Gln	Ala Trp Gln
2390	2395	2400
Pro Gly Val Ala Met Pro Asn	Leu Tyr Lys Met Gln	Arg Met Leu
2405	2410	2415
Leu Glu Lys Cys Asp Leu Gln	Asn Tyr Gly Glu Asn	Ala Val Ile
2420	2425	2430
Pro Lys Gly Ile Met Met Asn	Val Ala Lys Tyr Thr	Gln Leu Cys
2435	2440	2445
Gln Tyr Leu Asn Thr Leu Thr	Leu Ala Val Pro Tyr	Asn Met Arg
2450	2455	2460
Val Ile His Phe Gly Ala Gly	Ser Asp Lys Gly Val	Ala Pro Gly
2465	2470	2475
Thr Ala Val Leu Arg Gln Trp	Leu Pro Thr Gly Thr	Leu Leu Val
2480	2485	2490
Asp Ser Asp Leu Asn Asp Phe	Val Ser Asp Ala Asp	Ser Thr Leu
2495	2500	2505
Ile Gly Asp Cys Ala Thr Val	His Thr Ala Asn Lys	Trp Asp Leu
2510	2515	2520
Ile Ile Ser Asp Met Tyr Asp	Pro Arg Thr Lys His	Val Thr Lys
2525	2530	2535
Glu Asn Asp Ser Lys Glu Gly	Phe Phe Thr Tyr Leu	Cys Gly Phe
2540	2545	2550
Ile Lys Gln Lys Leu Ala Leu	Gly Gly Ser Ile Ala	Val Lys Ile
2555	2560	2565
Thr Glu His Ser Trp Asn Ala	Asp Leu Tyr Lys Leu	Met Gly His
2570	2575	2580
Phe Ser Trp Trp Thr Ala Phe	Val Thr Asn Val Asn	Ala Ser Ser
2585	2590	2595
Ser Glu Ala Phe Leu Ile Gly	Ala Asn Tyr Leu Gly	Lys Pro Lys
2600	2605	2610
Glu Gln Ile Asp Gly Tyr Thr	Met His Ala Asn Tyr	Ile Phe Trp
2615	2620	2625
Arg Asn Thr Asn Pro Ile Gln	Leu Ser Ser Tyr Ser	Leu Phe Asp
2630	2635	2640
Met Ser Lys Phe Pro Leu Lys	Leu Arg Gly Thr Ala	Val Met Ser
2645	2650	2655
Leu Lys Glu Asn Gln Ile Asn	Asp Met Ile Tyr Ser	Leu Leu Glu
2660	2665	2670
Lys Gly Arg Leu Ile Ile Arg	Glu Asn Asn Arg Val	Val Val Ser
2675	2680	2685
Ser Asp Ile Leu Val Asn Asn		
2690	2695	

<210> SEQ ID NO 76

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L3/+/4932 primer

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<400> SEQUENCE: 76
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<210> SEQ ID NO 77
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<212> TYPE: DNA
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<220> FEATURE:
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<400> SEQUENCE: 77
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<210> SEQ ID NO 78
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
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<400> SEQUENCE: 78
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<210> SEQ ID NO 79
<211> LENGTH: 20
<212> TYPE: DNA
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<400> SEQUENCE: 79
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<210> SEQ ID NO 80
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<400> SEQUENCE: 80
tggtcagtag ggttgattgg 20

<210> SEQ ID NO 81
<211> LENGTH: 20
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<213> ORGANISM: Artificial sequence
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<400> SEQUENCE: 81
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<210> SEQ ID NO 82
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L5/-8633 primer

<400> SEQUENCE: 82
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<210> SEQ ID NO 83
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<400> SEQUENCE: 83

atgcgacgag tctgcttcta 20

<210> SEQ ID NO 84
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<212> TYPE: DNA
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<400> SEQUENCE: 84

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<210> SEQ ID NO 85
<211> LENGTH: 20
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<223> OTHER INFORMATION: S/L5/+8255 primer

<400> SEQUENCE: 85

atcttggcgc atgtattgac 20

<210> SEQ ID NO 86
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/-9422 primer

<400> SEQUENCE: 86

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<210> SEQ ID NO 87
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/-9966 primer

<400> SEQUENCE: 87

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<210> SEQ ID NO 88
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/-10542 primer

<400> SEQUENCE: 88

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<210> SEQ ID NO 89
<211> LENGTH: 20
<212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L6/+10677 primer

<400> SEQUENCE: 89
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<210> SEQ ID NO 90
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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: S/L6/+10106 primer

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<210> SEQ ID NO 91
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<212> TYPE: DNA
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<223> OTHER INFORMATION: S/L6/+9571 primer

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<210> SEQ ID NO 92
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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: S/L7/-/11271 primer

<400> SEQUENCE: 92
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<210> SEQ ID NO 93
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<210> SEQ ID NO 94
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<210> SEQ ID NO 95
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<210> SEQ ID NO 96

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<223> OTHER INFORMATION: S/L7/+11551 primer

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<210> SEQ ID NO 98

<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L8/-13160 primer

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<210> SEQ ID NO 99

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<212> TYPE: DNA

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<220> FEATURE:

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<210> SEQ ID NO 100

<211> LENGTH: 20

<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: S/L8/-14284 primer

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<210> SEQ ID NO 104
<211> LENGTH: 20
<212> TYPE: DNA
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<223> OTHER INFORMATION: S/L9/-/15098 primer

<400> SEQUENCE: 104

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<210> SEQ ID NO 105
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/-/15677 primer

<400> SEQUENCE: 105

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<210> SEQ ID NO 106
<211> LENGTH: 20
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<220> FEATURE:
<223> OTHER INFORMATION: S/L9/-/16247 primer

<400> SEQUENCE: 106

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<210> SEQ ID NO 107
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L9/+/16323 primer

<400> SEQUENCE: 107

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<210> SEQ ID NO 108
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<212> TYPE: DNA

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<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: S/L9/+15858 primer

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<223> OTHER INFORMATION: S/L9/+15288 primer

<400> SEQUENCE: 109
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<223> OTHER INFORMATION: S/L10/-16914 primer

<400> SEQUENCE: 110
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<210> SEQ ID NO 111
<211> LENGTH: 20
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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L10/-17466 primer

<400> SEQUENCE: 111
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<210> SEQ ID NO 112
<211> LENGTH: 20
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<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: S/L10/-18022 primer

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<210> SEQ ID NO 113
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
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<400> SEQUENCE: 113
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<210> SEQ ID NO 114
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<223> OTHER INFORMATION: S/L10/+17061 primer

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<211> LENGTH: 20

<212> TYPE: DNA

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<223> OTHER INFORMATION: S/L11/-18877 primer

<400> SEQUENCE: 116

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<210> SEQ ID NO 117

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<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L11/-19396 primer

<400> SEQUENCE: 117

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<223> OTHER INFORMATION: S/L11/-20002 primer

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<211> LENGTH: 20

<212> TYPE: DNA

<213> ORGANISM: Artificial sequence

<220> FEATURE:

<223> OTHER INFORMATION: S/L11/+20245 primer

<400> SEQUENCE: 119

tcgaacacat cgtttatgga

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<210> SEQ ID NO 120

<211> LENGTH: 20

<212> TYPE: DNA

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<220> FEATURE:

<223> OTHER INFORMATION: S/L11/+19611 primer

<400> SEQUENCE: 120

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<210> SEQ ID NO 121
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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: S/L11/+ /19021 primer

<400> SEQUENCE: 121

acgatgctca gccatgtagt 20

<210> SEQ ID NO 122
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<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: SARS/L1/F3/+ /800 primer

<400> SEQUENCE: 122

gaggtgcagt cactcgctat 20

<210> SEQ ID NO 123
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<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L1/F4/+ /1391 primer

<400> SEQUENCE: 123

cagagattgg acctgagcat 20

<210> SEQ ID NO 124
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L1/F5/+ /1925 primer

<400> SEQUENCE: 124

cagcaaacca ctcaattcct 20

<210> SEQ ID NO 125
<211> LENGTH: 20
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<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L1/R3/- /1674 primer

<400> SEQUENCE: 125

aaatgatggc aacctcttca 20

<210> SEQ ID NO 126
<211> LENGTH: 20
<212> TYPE: DNA
<213> ORGANISM: Artificial sequence
<220> FEATURE:
<223> OTHER INFORMATION: SARS/L1/R4/- /1107 primer

<400> SEQUENCE: 126

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<210> SEQ ID NO 127
<211> LENGTH: 20
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<213> ORGANISM: Artificial sequence
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<223> OTHER INFORMATION: SARS/L1/R5/-/520 primer

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<210> SEQ ID NO 128
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<223> OTHER INFORMATION: SARS/L2/F3/+2664 primer

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<210> SEQ ID NO 129
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<400> SEQUENCE: 130
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<210> SEQ ID NO 132
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<213> ORGANISM: Artificial sequence

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<223> OTHER INFORMATION: SRAS/L3/F4/+ /5305 primer

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<210> SEQ ID NO 136

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<212> TYPE: DNA

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<212> TYPE: DNA

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<223> OTHER INFORMATION: SARS/L3/R4/- /4988 primer

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<210> SEQ ID NO 139

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<223> OTHER INFORMATION: SARS/L3/R5/- /4437 primer

<400> SEQUENCE: 139

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<210> SEQ ID NO 140
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<213> ORGANISM: Artificial sequence
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<212> TYPE: DNA
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<400> SEQUENCE: 157

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1. An isolated or purified strain of severe acute respiratory syndrome-associated human coronavirus, characterized in that its genome has, in the form of complementary DNA, a serine codon at position 23220-23222 of the gene for the S protein or a glycine codon at position 25298-25300 of the gene for ORF3, and an alanine codon at position 7918-7920 of ORF1a or a serine codon at position 26857-26859 of the gene for the M protein, said positions being indicated in terms of reference to the Genbank sequence AY274119.3.

2. The isolated or purified coronavirus strain as claimed in claim 1, characterized in that the DNA equivalent of its genome has a sequence corresponding to the sequence SEQ ID NO: 1.

3. An isolated or purified polynucleotide, characterized in that its sequence is that of the genome of the isolated coronavirus strain as claimed in claim 1 or claim 2.

4. The isolated or purified polynucleotide as claimed in claim 3, characterized in that its sequence is SEQ ID NO: 1.

5. A pair of primers capable of amplifying a fragment of the sequence of the genome of a SARS-associated coronavirus or of its DNA equivalent, characterized in that it is selected from the group consisting of:

the pair of primers No. 1 corresponding respectively to positions 28507 to 28522 (sense primer, SEQ ID NO: 60) and 28774 to 28759 (antisense primer, SEQ ID NO: 61) of the sequence of the polynucleotide as claimed in claim 3 or claim 4,

the pair of primers No. 2 corresponding respectively to positions 28375 to 28390 (sense primer, SEQ ID NO: 62) and 28702 to 28687 (antisense primer, SEQ ID NO: 63) of the sequence of the polynucleotide as claimed in claim 3 or claim 4, and

the pair of primers consisting of the primers SEQ ID Nos: 55 and 56.

6. A probe capable of detecting the presence of the genome of a SARS-associated coronavirus or of a fragment thereof, characterized in that it is selected from the group consisting of the fragments corresponding to the following positions of the polynucleotide sequence as claimed in claim 3 or claim 4: 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 (SEQ ID NO: 64 to 67).

7. A recombinant cloning and/or expression vector, characterized in that it comprises an insert having the sequence SEQ ID NO: 38 and it is contained in a bacterial strain and it was deposited under the No. I-3048, on Jun. 5, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

8. A recombinant cloning and/or expression vector, characterized in that it contains a cDNA fragment selected from the group consisting of:

a cDNA fragment encoding a C-terminal fusion of the N protein (SEQ ID NO: 37) with a polyhistidine tag, and

a cDNA fragment encoding an N-terminal fusion of the N protein (SEQ ID NO: 37) with a polyhistidine tag.

9. The recombinant expression vector as claimed in claim 8, characterized in that it is contained in a bacterial strain which was deposited under the No. I-3117, on Oct. 23, 2003, at the Collection Nationale de Cultures de Microorganismes, 25 rue du Docteur Roux, 75724 Paris Cedex 15.

10. A cell modified with a vector as claimed in any one of claims 7 to 9.

11. A hybridoma producing a monoclonal antibody against the N protein, characterized in that it is chosen from the following hybridomas:

the hybridoma producing the monoclonal antibody 87, deposited at the CNCM on Dec. 1, 2004 under the number I-3328,

the hybridoma producing the monoclonal antibody 86, deposited at the CNCM on Dec. 1, 2004 under the number I-3329,

the hybridoma producing the monoclonal antibody 57, deposited at the CNCM on Dec. 1, 2004 under the number I-3330, and

the hybridoma producing the monoclonal antibody 156, deposited at the CNCM on Dec. 1, 2004 under the number I-3331.

12. A polyclonal or monoclonal antibody or antibody fragment directed against the N protein, characterized in that it is produced by a hybridoma as claimed in claim 11.

13. A chip or filter, characterized in that it comprises an antibody or an antibody fragment as claimed in claim 12.

14. An immunocapture test intended to detect a SARS-associated coronavirus infection, characterized in that it uses a monoclonal antibody specific for the native viral nucleoprotein (N protein).

15. The immunocapture test as claimed in claim 14, characterized in that the antibody used for the capture of the native viral nucleoprotein is a monoclonal antibody specific for the central region and/or for a conformational epitope.

16. The immunocapture test as claimed in claim 14 or 15, characterized in that the antibody used for the capture of the N protein is the monoclonal antibody mAb87, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3328.

17. The immunocapture test as claimed in claim 14 or 15, characterized in that the antibody used for the capture of the N protein is the monoclonal antibody mAb86, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3329.

18. The immunocapture test as claimed in claim 14 or 15, characterized in that the monoclonal antibodies mAb86 and mAb87 are used for the capture of the N protein.

19. The immunocapture test as claimed in any one of claims 14 to 18, characterized in that the antibody used for the visualization of the N protein is the monoclonal antibody mAb57, produced by the hybridoma deposited at the CNCM on Dec. 1, 2004 under the number I-3330, said antibody being conjugated with a visualizing molecule or particle.

20. The immunocapture test as claimed in any one of claims 14 to 18, characterized in that a combination of the mAb57 and mAb87 antibodies, conjugated with a visualizing molecule or particle, is used for the visualization of the N protein.

21. A reagent for the detection of a SARS-associated coronavirus, characterized in that it is selected from the group consisting of:

- (a) a pair of primers as claimed in claim 5, or a probe as claimed in claim 6,
- (b) a recombinant vector as claimed in any one of claims 7 to 9 or a modified cell as claimed in claim 10,
- (c) an isolated coronavirus strain as claimed in claim 1 or claim 2 or a polynucleotide as claimed in either of claims 3 and 4,
- (d) an antibody or an antibody fragment as claimed in claim 12,

(e) a combination of antibodies comprising the monoclonal antibodies mAb86 and/or mAb87, and the monoclonal antibody mAb57;

(f) a chip or a filter as claimed in claim 13.

22. The use of a product selected from the group consisting of: a pair of primers as claimed in claim 5, a probe as claimed in claim 6, a recombinant vector as claimed in any one of claims 7 to 9, a modified cell as claimed in claim 10, an isolated coronavirus strain as claimed in claim 1 or claim 2, a polynucleotide as claimed in claim 3 or claim 4, for the preparation of a reagent for the detection and optionally genotyping of a SARS-associated coronavirus.

23. A method for the detection of a SARS-associated coronavirus, from a biological sample, which method is characterized in that it comprises at least:

- (a) the extraction of nucleic acids present in said biological sample,
- (b) the amplification of a fragment of ORF-N by RT-PCR with the aid of a pair of primers as claimed in claim 5, and
- (c) the detection, by any appropriate means, of the amplification products obtained in (b).

24. The method as claimed in claim 23, characterized in that step (b) of detection is carried out with the aid of at least one probe corresponding to positions 28561 to 28586, 28588 to 28608, 28541 to 28563 and 28565 to 28589 of the sequence of the polynucleotide as claimed in claim 3 or claim 4.

25. A method for the detection of a SARS-associated coronavirus infection, from a biological sample, by indirect IgG ELISA using the N protein, which method is characterized in that the plates are sensitized with an N protein solution at a concentration of between 0.5 and 4 µg/ml, preferably 2 µg/ml, in a 10 mM PBS buffer, pH 7.2, phenol red at 0.25 ml/l.

26. A method for the detection of a SARS-associated coronavirus infection, from a biological sample, by double epitope ELISA, characterized in that the serum to be tested is mixed with the visualizing antigen, said mixture then being brought into contact with the antigen attached to a solid support.

27. An immune complex formed of a polyclonal or monoclonal antibody or antibody fragment as claimed in claim 11, and of a SARS-associated coronavirus protein or peptide.

28. A SARS-associated coronavirus detection kit or box, characterized in that it comprises at least one reagent selected from the group consisting of: a pair of primers as claimed in claim 5, a probe as claimed in claim 6, a recombinant vector as claimed in any one of claims 7 to 9, a modified cell as claimed in claim 10, an isolated coronavirus strain as claimed in claim 1 or claim 2 and a polynucleotide as claimed in claim 3 or claim 4.

29. A fragment of the polynucleotide as claimed in claim 3, characterized in that it includes at least one pair of bases or pairs of bases corresponding to the following positions: 7919 and 23220, 7919 and 25298, 16622 and 23220, 19064 and 23220, 16622 and 25298, 19064 and 25298, 23220 and 24872, 23220 and 26857, 24872 and 25298, 25298 and 26857.